

# Perceiving the epigenetic landscape through histone readers

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**Post-translational modifications (PTMs) of histones provide a fine-tuned mechanism for regulating chromatin structure and dynamics. PTMs can alter direct interactions between histones and DNA and serve as docking sites for protein effectors, or readers, of these PTMs. Binding of the readers recruits or stabilizes various components of the nuclear signaling machinery at specific genomic sites, mediating fundamental DNA-templated processes, including gene transcription and DNA recombination, replication and repair. In this review, we highlight the latest advances in characterizing histone-binding mechanisms and identifying new epigenetic readers and summarize the functional significance of PTM recognition.**

The genetic material of eukaryotic cells is packaged into the nucleus in the form of chromatin. Chromatin is made up of building blocks called nucleosomes. Each nucleosomal particle contains an octamer of four histone proteins—H2A, H2B, H3 and H4—around which genomic DNA is wound almost twice<sup>1</sup>. The nucleosomes undergo recurrent structural rearrangements through DNA unwrapping and rewrapping and histone core disassembly and assembly, and they are subject to covalent modifications. The modifications, or epigenetic marks, have been identified on both DNA and histones. Whereas DNA can primarily be methylated, histones are capable of carrying a wide array of PTMs<sup>2</sup>. A particularly large number of PTMs have been discovered on the histone tails that protrude from the nucleosomal core and are freely accessible to enzymes for the deposition or removal of PTMs (Fig. 1). The mechanisms by which histone PTMs affect chromatin structure and dynamics can generally be divided into two categories. Histone PTMs can directly influence histone-DNA and histone-histone interactions, or they can be targeted by protein effectors (also referred to as histone-binding domains or readers of PTMs).

The specific recognition of PTMs by readers recruits various components of the nuclear signaling network to chromatin, mediating fundamental processes such as gene transcription, DNA replication and recombination, DNA damage response and chromatin remodeling. Chromatin-associating complexes often contain multiple readers within one or several subunits that show specificities for distinct PTMs. Coordinated binding to multiple PTMs can provide a lock-and-key-type mechanism for targeting particular genomic sites and ensuring the proper biological outcomes. Misreading of epigenetic marks has been shown to underlie a host of human diseases, including autoimmune and developmental abnormalities and cancer<sup>3,4</sup>. More recently, misregulation of epigenetic pathways has been implicated in addiction, schizophrenia and other mental disorders<sup>5,6</sup>. Thus, understanding

the molecular mechanism and functional significance of reader-PTM interactions is essential to understanding not only the basic mechanisms of epigenetic regulation but also the etiology of epimutation-induced human diseases.

In this review, we outline known readers of histone PTMs, detail their mechanisms of action and discuss cross-talk between protein effectors and consequences of the combinatorial readout of PTMs.

## History of histone PTMs

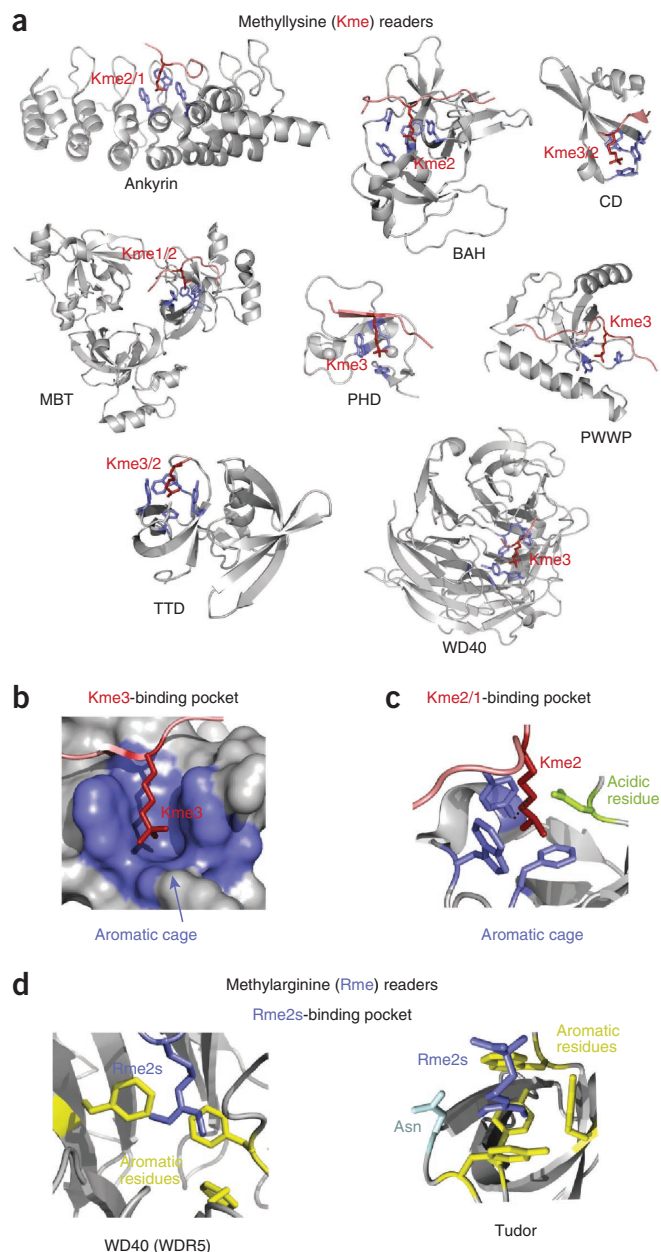
The first PTMs of histones were discovered nearly 50 years ago<sup>7</sup>; however, it was not until 30 years later that enzymatic activities of a histone acetyltransferase (HAT) and a deacetylase were directly linked to transcriptional regulation<sup>8,9</sup>. In addition to lysine acetylation, numerous PTMs have now been identified, including methylation, ubiquitination, SUMOylation, crotonylation, butyrylation and propionylation of lysine residues; methylation, citrullination and ADP-ribosylation of arginine residues; and phosphorylation and glycosylation of serine and threonine residues. As the substantial degree of interplay between PTMs began to unfold, it led Strahl and Allis to propose the histone code hypothesis, which states that “multiple histone modifications, acting in a combinatorial or sequential fashion on one or multiple histone tails, specify unique downstream function”<sup>10</sup>. Soon afterward, the terms ‘writer’, ‘eraser’ and ‘reader’ were formulated to describe proteins that deposit, remove and recognize PTMs, respectively<sup>11,12</sup>. In addition to histone PTMs, other factors—including methylation and hydroxymethylation of DNA, histone variants, nucleosome positioning, noncoding RNAs and histone chaperones—are necessary for fine-tuning chromatin structure and function, and together they constitute the powerful and dynamic epigenetic machinery.

The term ‘epigenetics’ was originally associated with heritable changes in gene activity that occur without alterations in the genetic code, sometimes defined as ‘soft’ inheritance<sup>13,14</sup>. However, more and more often this term is used to describe DNA-related regulatory mechanisms that do not involve changes in the nucleotide sequence, regardless of whether such imprinting is strictly heritable (‘epi-’ is derived from the Greek for ‘above’; hence, ‘above genetics’). The intricate relationship among the epigenetic elements represents one of the most intriguing concepts in modern chromatin biology, which we

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**Figure 2** Molecular mechanisms for the recognition of methyllysine and methylarginine. **(a)** Structures of the readers in complex with histone peptides methylated at lysine residues. The aromatic cage residues and the histone peptides are colored blue and red, respectively. **(b–d)** The binding sites for trimethylated lysine (Kme3) **(b)**, dimethylated lysine (Kme2) **(c)** and symmetrically dimethylated arginine (Rme2s) **(d)**. PDB codes: 3B95, 4DOW, 1KNE, 2PQW, 2G6Q, 2X4X, 2IG0, 31IW, 2F6J, 4A7J and 4A4E.

and dimethylated H3K9 (H3K9me2) (refs. 18,19). Upon binding, the histone peptide adopts a  $\beta$ -strand conformation and inserts between two  $\beta$ -strands of the chromodomain, completing the five-stranded antiparallel  $\beta$ -barrel. This induced-fit binding mode is stabilized through backbone hydrogen bonds and electrostatic contacts involving up to seven residues preceding trimethylated lysine and one residue following trimethylated lysine of the histone peptide. A small hydrophobic pocket, in which the  $n-2$  (with respect to methyllysine) residue resides, precludes binding of H3K4me3,

whereas interactions at the  $n-4$  and  $n-5$  positions are important for distinguishing between H3K9me3 and H3K27me3 (ref. 22).

Other Royal superfamily members have a complete five-stranded  $\beta$ -barrel, which prevents the insertion of a peptide between  $\beta$ -strands of the barrel. Instead, the histone tail lies across an open edge of the  $\beta$ -barrel, with the methyllysine occupying the aromatic cage located near the upper rim. Tudor is the classic module that utilizes this binding mechanism. Tudor can exist as a single domain or in tandem, containing two  $\beta$ -barrels. The single Tudor domains of PHF1 and PHF19 have been shown to recognize H3K36me3, whereas the canonical TTD of 53BP1 associates with H4K20me2, and the hybrid TTD of JMJD2A binds H3K4me3 and H4K20me3 (refs. 23–27,153). The aromatic cage is generally seen in only one  $\beta$ -barrel of TTDs, and the majority of contacts with the histone tail are found within that barrel. A notable exception to this is the TTD of Sgf29 (ref. 28). In the complex of the Sgf29 TTD with H3K4me3, the peptide spans both Tudor domains, with A1 and K4me3 being bound in separate  $\beta$ -barrels<sup>28</sup>. Here, the specificity toward H3K4 is largely defined by A1 recognition, a common determinant of H3K4me3 binding.

MBT reads lower methylation states, recognizing mono- and dimethylated lysines. All MBTs characterized at present contain two to four repeats of  $\sim 100$  amino acids. The most common three-repeat 3MBT module folds in a propeller-like structure, and the four-repeat 4MBT module has an asymmetric rhombus architecture<sup>29–32</sup>. Notably, although each MBT repeat contains an aromatic cage, which also includes an acidic residue, only the second repeat binds methyllysine<sup>29–32</sup>. Very little sequence specificity is observed, as few contacts are made with the histone residues beyond mono- or dimethylated lysine (refs. 29–33). The other two members of the Royal family, PWWP and chromo-barrel, show some specificity, preferring tri- or dimethylated H3K36 and monomethylated H4K20 (H4K20me1); however, they bind rather weakly<sup>34–39</sup>.

Distinct from the Royal family is the PHD finger, a well-characterized reader of H3K4me3 (refs. 40–43). It contains a  $C_4HC_3$  motif that coordinates two zinc ions in a cross-brace manner. Such an arrangement produces a globular domain with a small  $\beta$ -sheet and an  $\alpha$ -helix. PHD fingers make extensive contacts with H3K4me3, imparting a high degree of specificity<sup>44</sup>. The peptide forms the third antiparallel  $\beta$ -strand, pairing with the existing double-stranded  $\beta$ -sheet of the protein. While K4me3 inserts into the aromatic cage, R2 occupies the adjacent binding pocket. This pocket often contains acidic residues, which restrain the guanidinium group of R2 through ionic and hydrogen-bonding interactions. A1 is bound in a small hydrophobic cavity, and its N-terminal amino group donates two or three hydrogen bonds to the backbone carbonyl groups of the protein. This strictly conserved coordination of A1 and a frequent constraining of R2, as well as the fitting of T3 into a small pocket, provide the specificity for K4me3. A similar mode of recognition of the histone A1R2T3K4me3 sequence is seen in a single zinc finger (zf-CW)<sup>45</sup> and in the DCD of human CHD1, in which two chromodomains fold in a unique module, forming a conjoint binding site for the peptide<sup>46</sup>.

The WD40 domain of EED binds trimethylated lysines through an aromatic cage positioned at the top of the channel of a seven-bladed  $\beta$ -propeller<sup>47,48</sup>. It is fairly promiscuous and interacts with H3K27me3, H3K9me3, H4K20me3 and H1K26me3. The structure of the EED WD40 in complex with H3K27me3 reveals an important role for the residues flanking K27me3 (refs. 47,48). A small hydrophobic residue, a solvent-exposed residue and another hydrophobic residue at positions  $-2$ ,  $-1$  and  $+2$ , respectively, are responsible for the preferential recognition of repressive chromatin marks and the disfavoring of H3K4me3. Owing to extensive intermolecular



contacts, the ankyrin repeats of G9a and GLP and the BAH domain of ORC1 show high specificity for di- or monomethylated H3K9 and H4K20me2, respectively<sup>49,50</sup>. The H3K9me2 peptide is sandwiched between the  $\beta$ -turns and  $\alpha$ -helices of the fourth and fifth ankyrin repeats, whereas K9me2 fits into the aromatic cage also containing a glutamate. Residues S10–G13 of the H3K9me2 peptide are involved in multiple interactions that impart specificity. The ATRX ADD domain and the PHD fingers of CHD4 and TRIM33 are examples of readers lacking the aromatic cage<sup>51–55</sup>. Binding of these modules to the histone H3 tail is enhanced by methylation of H3K9. K9me3 is uniquely coordinated through formation of a nonconventional carbon-oxygen hydrogen bond or hydrophobic and cation- $\pi$  contacts with a single aromatic residue.

**Functional significance of Kme recognition.** Methylation of lysine residues is the most characterized PTM in terms of cellular functions, hallmarked by its major role in transcriptional regulation. On a global chromatin level, H3K4me is considered to be a gene-activation mark; however, the consequence of its readout by histone effectors is highly context dependent. Lysine trimethylation is generally located in the 5' end region of actively transcribed genes, and this trait is conserved from yeast to higher eukaryotes<sup>56–58</sup>. The TAF3 subunit of the basal transcription complex TFIID binds to H3K4me3 through its PHD finger, directly implicating this mark in transcriptional activation<sup>59</sup>. In contrast, recognition of H3K4me3 by the PHD finger of ING2, a subunit of the mSin3a histone deacetylase complex, links this PTM to rapid gene repression<sup>42</sup>. Binding of the PHD finger of the PHF8 histone demethylase<sup>60</sup> and the DCD of the CHD1 ATPase<sup>46</sup> to H3K4me3 mediates enzymatic activities of these proteins, coupling the readout of H3K4me to the chromatin-modifying and chromatin-remodeling mechanisms, respectively. The chromo-barrel domain of the Tip60 acetyltransferase associates with H3K4me1, a marker of active and poised enhancer elements, tethering Tip60 to the enhancer regions and leading to estrogen-induced transcription<sup>61</sup>.

H3K36me also has a role in the transcriptional process. Genome-wide profiling of H3K36me stages shows a progressive shift from monomethylation to trimethylation of K36 between the 5' and 3' ends of genes<sup>62</sup>. The H3K36me-specific enzyme Set2 binds to elongating RNA polymerase II and methylates the coding regions of genes, preventing spurious transcription initiation and histone exchange in budding yeast by recruiting the Rpd3S deacetylase complex via its Eaf3 subunit, which binds to K36me through its chromo-barrel domain<sup>63–65</sup>. This complex deacetylates nucleosomes on gene bodies during elongation and thus provides nucleosome stability in the wake of elongating RNA polymerase II<sup>66</sup>. Recognition of H3K36me3 by the Tudor domain of PHF1 inhibits Polycomb repressive complex 2 (PRC2)-mediated H3K27 methylation<sup>27</sup>. This interaction may represent a mechanism to guide the deposition of H3K27me3, restricting activity of the PRC2 complex at the boundaries of active and repressed regions and preventing the Polycomb-linked repressive environment from spreading into neighboring transcriptionally active chromatin<sup>67,68</sup>. Interaction of another component of the PRC2 complex, PHF19, with H3K36me3 is important for localization of PHF19 at a subset of PRC2 target genes and is required for their repression and H3K27me3 deposition in mouse embryonic stem cells<sup>23</sup>, as well as for the recruitment of demethylase NO66 to embryonic stem cell genes during differentiation (ref. 153).

The importance of H3K36me recognition for other fundamental processes has recently been established. Association of the chromo-barrel domains of MRG15 and the MSL3 subunit of the MOF complex with H3K36me is essential in regulating alternative splicing

of mRNAs<sup>69</sup> and upregulating genes for dosage compensation in *Drosophila melanogaster*<sup>70</sup>, respectively. The PWWP domain present in the IOC4 subunit of the ISW1b chromatin-remodeling complex also recognizes H3K36me<sup>71,72</sup>, bridging readout of this PTM with ATP-dependent nucleosome remodeling activity. An additional role for H3K36me in DNA repair and recombination through the LEDGF PWWP domain has been suggested<sup>73</sup>; however, it remains unclear whether this PTM has a positive or negative regulatory function in DNA replication<sup>74,75</sup>. Recently, the BAH domain of ORC1 has been shown to bind H4K20me2, and this interaction aids in DNA replication licensing<sup>50</sup>. The H4K20me2 mark also has a crucial role in the DNA repair pathway in which it is targeted by the TTD of 53BP1 (ref. 24). This interaction is in direct competition with the JMJD2A and JMJD2B histone demethylases, which bind to H4K20me2 through their TTDs before DNA damage<sup>76</sup>. Upon DNA damage, JMJD2 is degraded through the RNF8- and RNF168-dependent pathways, exposing H4K20me2 to the 53BP1 TTD<sup>76</sup>.

Recognition of the H3K9 and H3K27 methylation marks is largely associated with formation of constitutive and facultative heterochromatin and gene silencing. H3K27me1 and H3K9me3 are found in pericentromeric heterochromatin regions<sup>77,78</sup>, whereas H3K27me3 and H3K9me2 colocalize in repressed euchromatin regions. The K27me and K9me marks are both targets for chromodomain-containing proteins. The former recruits Polycomb proteins and their host PRC1 complexes to repressed chromatin<sup>20,21</sup>, whereas the latter recruits HP1 proteins involved in heterochromatin formation and spreading<sup>17,79,80</sup>.

**Arginine methylation.** The omega nitrogen atoms of arginine can be monomethylated or dimethylated symmetrically (Rme2s) or asymmetrically (Rme2a). To date, methylation of H3R2, H3R8, H3R17, H3R26, H4R3, H2AR11 and H2AR29 has been reported. As compared to lysine methylation, information on writing and especially erasing and recognition of methylated arginine is rather limited; however, there are some indications that this mark is read by histone effectors, and a number of readers show sensitivity to the methylation state of arginine.

The Tudor domain is known to bind methylated arginines in nonhistone ligands, particularly PIWI<sup>81</sup>, and recent studies suggest that the TDRD3 Tudor domain recognizes H3R17me2a and H4R3me2a<sup>82,83</sup>. Although no structure has yet been determined of a Tudor bound to a histone peptide containing a methylated arginine, the structures of the SMN and SPF30 Tudor domains in complex with a dimethylated arginine amino acid provide insight into how this mark can be specifically recognized<sup>84</sup>. Like methyllysine, methylated arginine occupies an aromatic cage at the top of the Tudor  $\beta$ -barrel (Fig. 2d). However, this cage is much narrower than the cage for methyllysine and thus favors the planar guanidinium group.

The WD40 repeat of WDR5 prefers H3R2me2s over H3 peptides that are unmodified or methylated at K4 (ref. 85). The first three N-terminal residues of H3 are involved in the majority of direct contacts, which are also conserved in the complexes of WDR5 with unmodified H3 and H3K4me<sup>86–89</sup>. The side chain of R2me2s inserts deeply into the open channel of the  $\beta$ -propeller, with the methylated guanidine moiety being sandwiched between two phenylalanine rings<sup>85</sup> (Fig. 2d). Notably, in the complexes of WDR5 with various H3 peptides, R2 is bound in the same pocket and adopts a similar conformation; however, Rme2s makes tighter hydrophobic interactions, whereas unmethylated R2 forms an additional water-mediated hydrogen bond<sup>85–88</sup>.

**Functional significance of Rme recognition.** Very little is known about the significance of methylarginine readout in histone proteins. One of the few reported examples involves recognition of H3R17me2a

**Figure 3** Molecular mechanisms for the recognition of acetyllysine, phosphoserine or phosphothreonine and unmodified histone H3. (a–c) Structures of the readers bound to histone peptides that are acetylated at lysine residues (a), phosphorylated at serine or threonine residues (b) and unmodified (c). Binding-site residues are colored green (a), pink (b) and wheat (c) with the corresponding peptides in orange, blue and purple, respectively. PDB codes: 1E6I, 2KWJ, 2WPP2, 4AON, 2AZM, 2C1N, 3L41, 3A1B, 2YBA and 2PUY.

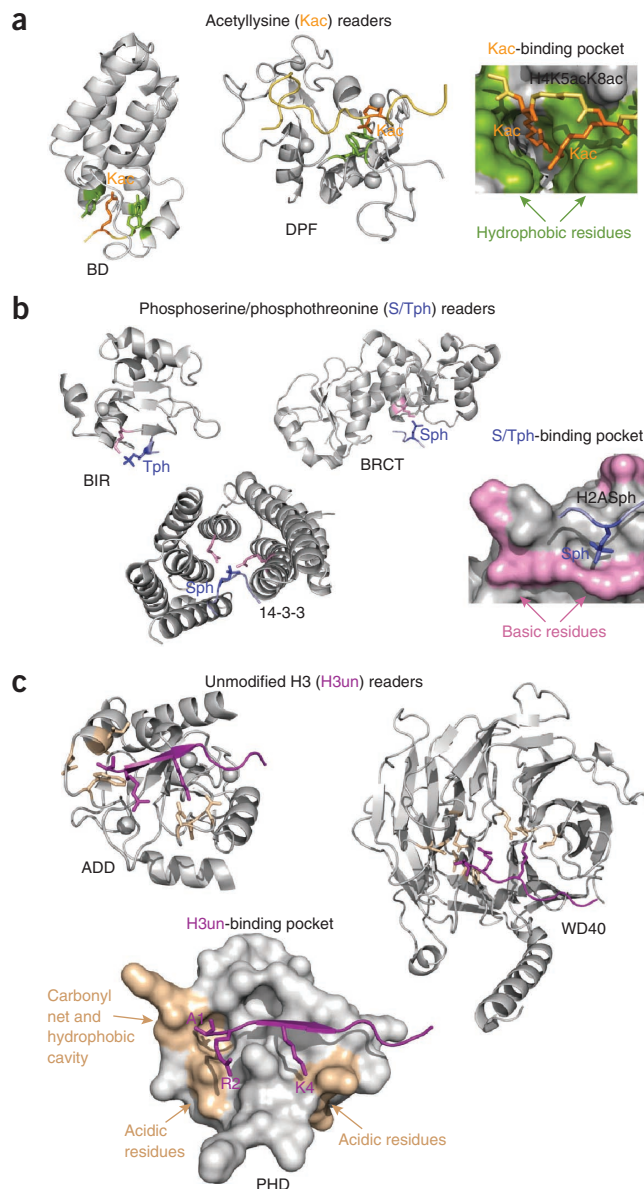
and H4R3me2a by the TDRD3 Tudor domain, which facilitates recruitment of this co-transcriptional activator to gene promoters and is necessary for proper coactivation<sup>90</sup>. The ADD domain of the DNMT3a DNA methyltransferase has been shown to interact with H4R3me2s, promoting silencing of the human  $\beta$ -globin locus<sup>91</sup>. The association of the WDR5 WD40 module with H3R2me2s—which is produced by PRMT5 and PRMT7—is important for euchromatin maintenance<sup>85</sup>. Together, these examples demonstrate that recognition of methylated arginines in histone proteins can influence transcription processes.

### Acetylation readers

Besides being methylated, lysine can also be acetylated at the  $\epsilon$ -amino group. Unlike methylation, this modification changes the electrostatic properties of histone proteins by neutralizing the charge of lysine. At present, lysine acetylation has been found to occur on H3 (K4, K9, K14, K18, K23, K27, K36 and K56), H4 (K5, K8, K12, K16, K20 and K91), H2A (K5 and K9) and H2B (K5, K12, K15, K16, K20 and K120). In general, this PTM leads to a more open chromatin structure, as it weakens the interaction with the negatively charged DNA and is largely associated with a transcriptionally active state. To date, three histone effectors capable of reading acetylated lysine have been identified.

The bromodomain is the most thoroughly characterized acetyllysine reader<sup>15,92</sup>. Despite little sequence similarity between family members, bromodomains fold into a highly conserved four-helix bundle structure (helices  $\alpha$ A,  $\alpha$ B,  $\alpha$ C and  $\alpha$ Z). The inter-helical ZA and BC loops create a deep, primarily hydrophobic cavity into which the acetyllysine inserts. Acetylated lysine makes several contacts with hydrophobic residues, including two often conserved tyrosines, and is stabilized by a hydrogen bond with a highly conserved asparagine. The surface properties of bromodomains vary considerably, accounting for a wide array of ligands<sup>93</sup>. In general, isolated bromodomains bind to acetylated histones weakly, yet at times they display specificity, imparted by interaction with surrounding residues. Simultaneous recognition of multiply acetylated sequences, either by a single bromodomain or by covalently linked bromodomains, can substantially enhance binding, as seen in TAF1 and Brdt<sup>94,95</sup>. Two bromodomains in the double bromodomain (DBD) of TAF1 create a V-shaped structure, with both acetyllysine binding pockets oriented in the same direction and separated by  $\sim 25$  Å, a distance ideal for recognition of a doubly acetylated H4 tail<sup>94</sup>. Consistently, the TAF1 DBD shows a much higher affinity for H4K5acK12ac and H4K8acK16ac peptides than for monoacetylated peptides, suggesting that each bromodomain interacts with one acetyllysine in the same peptide. Brdt utilizes a different mechanism to increase binding. The first bromodomain of Brdt accommodates both acetyllysines of H4K5acK8ac in a wider hydrophobic pocket (Fig. 3a), whereas the second bromodomain binds H3K18ac<sup>95</sup>.

More recently, the double PHD finger (DPF) of Dpf3b has been found to associate with acetylated histone peptides<sup>96,97</sup>. The PHD modules in DPF adopt a typical zinc-finger topology but have a large interface, which results in an overall unique scaffold<sup>97</sup>. The structure



of the DPF–H3K14ac complex reveals that the PHD modules are engaged in coordinated interaction<sup>97</sup>. Whereas the first four N-terminal residues of H3K14ac are bound by the second PHD module, the first PHD module anchors K14ac in the binding pocket composed of hydrophobic and charged residues, including the necessary aspartate-phenylalanine sequence.

The double pleckstrin homology (PH) domain of the histone chaperone Rtt106 has recently been implicated in binding to H3K56ac<sup>98</sup>. Each PH domain consists of a seven-stranded  $\beta$ -barrel, capped by one or two  $\alpha$ -helices at one of the open ends. The opposite end of the barrel is framed by three variable loops, which, in a canonical PH domain, comprise the binding site for a phosphoinositide lipid<sup>99</sup>. Although it is unclear whether Rtt106 binds lipids, chemical-shift perturbation analysis reveals that the H3K56ac-binding pocket is located in the second PH module at the interface of the C-terminal  $\alpha$ -helix and the  $\beta$ -barrel, with an aspartate, tyrosine and arginine likely forming a cleft for acetylated lysine<sup>98</sup>.

**Functional significance of Kac recognition.** For a long time, acetylation of histones was considered to be merely a mechanism

for chromatin opening by way of disrupting the association between histones and DNA; however, a number of effector-harboring proteins have been found to bind acetylated lysine. Many of these interactions are implicated in gene transcription, linking the direct readout of this PTM to transcriptional control. The BRD family of proteins is composed of transcription regulators that possess bromodomains, which are specific for singly or multiply acetylated histone peptides<sup>93</sup>. Activation and regulation of the DPF3b and MOZ target genes requires binding of their DPFs to H3K14ac<sup>97,100</sup>.

Recognition of H3K56ac by the double PH domain of Rtt106 is essential in gene silencing as well as DNA damage response<sup>98</sup>. Likewise, association of the bromodomain of BRG1 (the catalytic subunit of SWI/SNF) with acetylated H3 is necessary for DNA repair<sup>101</sup>. Yeast DBD-containing proteins Bdf1 and Bdf2 target H4Kac and regulate transcription and chromatin dynamics, but are also important for replication-coupled DNA repair and normal mRNA splicing<sup>102–105</sup>. The bromodomain-harboring protein ACF1 is enriched in replicating pericentromeric heterochromatin, and its depletion causes a delay in cell-cycle progression<sup>106</sup>. A closely related protein, BPTF, contains a bromodomain that binds H4K16ac, which is important for the *Hox* gene localization of the BPTF–NURF complex<sup>107</sup>.

### Phosphorylation readers

Similar to lysine acetylation, phosphorylation of serine and threonine alters the electrostatic and topographic properties of histones by adding a bulky, negatively charged group to the modified residue. Recognition of phosphorylated sequences by cytosolic proteins has been characterized relatively well; however, much less is known about histone phosphorylation. Histones can be phosphorylated at T3, T6, S10, T11, S28 and T45 of H3; S1 of H4; S1 and T120 of H2A; S139 of human histone variant H2AX and S14 of H2B. These PTMs are essential in DNA damage response pathways, mitosis and transcriptional regulation.

The tandem BRCT domain has been found to specifically recognize phosphorylated S139 of H2AX (H2AXS139ph, also referred to as  $\gamma$ H2AX)<sup>108</sup>. Each BRCT has an  $\alpha\beta$  sandwich architecture, with a central four-stranded  $\beta$ -sheet surrounded by  $\alpha$ -helices on either side (Fig. 3b). The tandem repeats align in a head-to-tail manner, forming a considerable hydrophobic interface. The H2AXS139ph peptide lies in the groove at the interface of two repeats. The phosphorylated S139 residue is held in a basic site through direct and water-mediated hydrogen bonds and ionic interactions. In all characterized BRCT domains, specificity is defined by a hydrophobic residue at the +3 position, which is bound in a well-formed hydrophobic pocket on the protein surface<sup>108,109</sup>. Additionally, the carboxyl group at the C terminus of  $\gamma$ H2AX (at the +3 position) is stabilized by hydrogen bonds with a conserved arginine, contributing substantially to the strength and specificity of the interaction.

The 14-3-3 $\zeta$  isoform recognizes H3S10ph and H3S28ph<sup>110</sup>. 14-3-3 $\zeta$  belongs to a family of 14-3-3 proteins that generally interact with phosphorylated nonhistone targets. The crystal structure of 14-3-3 $\zeta$  bound to H3S10ph provides insight into the mechanism underlying specificity for the histone ligand. The H3S10ph peptide assumes a partially extended conformation and inserts into a cleft formed by five helices of the U-shaped, nine-helical protein. S10ph is restrained through multiple hydrogen bonding and ionic contacts with two arginine residues and a tyrosine, whereas the G12–G13 sequence adopts a conformation necessary for the C terminus of the peptide to exit the cleft.

The BIR domain of the chromosomal passenger complex subunit Survivin is the newest histone reader that binds H3T3ph<sup>111–113</sup>. BIR

folds into a small  $\alpha\beta$  structure stabilized by the coordination of a single zinc ion. The H3T3ph peptide is bound in an extended conformation across a largely electronegative surface of the BIR domain, with T3ph placed in a small, positively charged patch composed of a lysine and a histidine. The peptide is anchored through an extensive network of hydrogen bonds involving several aspartate and glutamate residues of BIR and the free amino group of A1, the side chain of K4 and the backbone of the peptide.

**Functional significance of Sph and Tph recognition.** One of the best-characterized histone phosphorylation marks is on S139 of mammalian and S129 of yeast histone H2A(X). In response to DNA damage, various kinases involved in the DNA repair pathway (ATM, ATR and DNA-PK) phosphorylate this residue to facilitate the repair process. Apart from the tandem BRCT domains of MDC1, 53BP1 and Crb2 (refs. 108,114,115), other proteins implicated in the DNA repair pathway have also been suggested to bind this PTM<sup>108,116,117</sup>. Likewise, a proapoptotic function has been proposed for the recognition of adjacent phosphorylated Y142 of H2AX by the PTB domain of Fe65 (ref. 118).

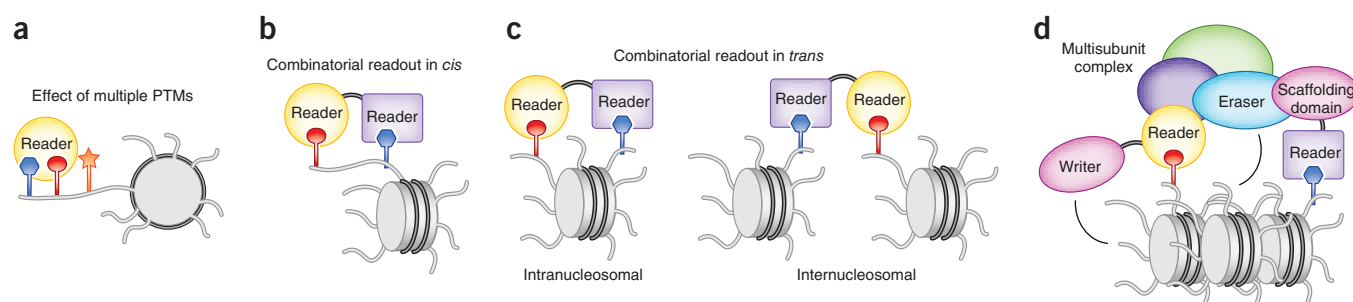
Two other established functions for histone serine or threonine phosphorylation are regulation of mitosis and transcription. The Aurora B kinase phosphorylates H3S10 during mitosis, and this phosphorylation is necessary for chromosome condensation and segregation<sup>119</sup>. This modification acts to release HP1 proteins from chromatin by disrupting the interaction of their chromodomains with H3K9me3 (refs. 119,120). Survivin binds H3T3ph through its BIR domain to allow proper Aurora B activity at inner centromeres<sup>111,121</sup>. The 14-3-3 family of proteins has many isoforms, some of which recognize H3S10ph and H3S28ph<sup>110</sup>. Moreover, this binding is reinforced by acetylation of K14 on the same histone tail<sup>122,123</sup>. The recruitment of 14-3-3 or its yeast homologs Bmh1 and Bmh2 is necessary for transcription activation of *GAL1* and *HDAC1*, among other genes<sup>122,123</sup>.

### Unmodified H3 readers

A large number of PHD fingers, as well as the ADD and WD40 modules, associate with the unmodified H3 tail (Fig. 3c). Some similarity in the molecular mechanisms for the recognition of unmodified H3 and H3K4me3 by the PHD fingers is apparent. Like H3K4me3, the unmodified H3 peptide forms the third antiparallel  $\beta$ -strand in the complex<sup>124</sup>. The side chain of A1 is buried in a small hydrophobic cavity, and the N-amino group of A1 is fixed by hydrogen bonds with backbone carbonyls of the protein. The guanidinium moiety of R2 is commonly restrained through the formation of hydrogen bonding and ionic contacts. Differences between the two binding mechanisms arise from the distinct coordination of K4. The unmodified H3-specific PHD fingers lack the aromatic cage, and instead, unmethylated K4 is bound by a set of hydrogen bonds and salt bridges formed with the acidic residues clustered on the surface of the protein. The majority of PHD fingers interact with a long stretch of the unmodified H3 tail, recognizing up to nine residues of H3 (ref. 44); however, the extended PHD finger of UHRF1 makes contacts with only the first four residues of unmodified H3 (refs. 125–127). The R2 residue of the peptide, rather than K4, is robustly bound by several hydrogen bonds, which results in a decreased sensitivity to PTMs on K4.

The ADD domain consists of a GATA-like C<sub>4</sub> finger followed by a PHD finger, which together coordinate three zinc ions. The ADD domain binds to the unmodified H3 peptide, using a mechanism typical of PHD fingers and recognizing seven residues of the peptide<sup>128</sup>. The WD40 domain of Nurf55 is in direct contact with nine residues of unmodified H3 (ref. 67). The structure of the Nurf55–unmodified H3 complex reveals a hydrophobic cavity accommodating A1 and several





**Figure 4** Combinatorial readout of PTMs. (a–c) Recognition of a target PTM is influenced by adjacent PTMs on the same histone tail (a) and the combined action of multiple readers within the same protein (b,c). (d) Multivalent engagement of readers within individual subunits of the complex. The reader-harboring proteins can also contain the catalytic domains (which act as writers and erasers) or scaffolding domains that bridge their host proteins with other subunits of the complex. Readers can recognize PTMs on a single histone tail (*cis* mechanism) or different histone tails (*trans* mechanism).

hydrogen bonds fixing the amino groups of A1 and K4. However, buried within the propeller fold, R2 is not stabilized through hydrogen bonds and is instead sandwiched between two aromatic residues.

### Combinatorial readout of and cross-talk between PTMs

The list of newly identified histone readers has grown rapidly, yet the finite number of readers and the numerous biological processes they mediate suggest that explicit mechanisms exist to differentiate functions of the effector-containing proteins and elicit distinct biological outcomes. Given the extensive and complex nature of the chromatin landscape, several mechanisms involving the combinatorial readout of epigenetic marks have been uncovered.

The activity of a reader toward a particular modification can be influenced by neighboring PTMs (Fig. 4a). In fact, many readers associate with a substantial stretch of the histone tail, allowing for the sensing of multiple marks. The neighboring PTMs can enhance or impede interaction with the target PTM. The classic example is the inhibition of binding of HP1 chromodomain to H3K9me3 by phosphorylation of S10, which may negatively regulate localization of HP1 at chromatin during mitosis<sup>120</sup>. Similarly, phosphorylation of T3 decreases affinity of CHD1 DCD for H3K4me3 (ref. 46). Methylation of H3K4 abrogates interactions of the unmodified H3-specific PHD fingers<sup>44</sup> and ADD<sup>51</sup>, whereas acetylation of H4K8 increases binding of the Brdt bromodomain to H4K5ac<sup>95</sup>. Methylation of R2 enhances interaction of the Rag2 PHD finger with H3K4me3 (ref. 129) but inhibits the histone binding activities of TAF3 and AIRE PHD fingers<sup>59,130</sup>.

Recruitment of a reader to a specific genomic region can be further modulated through the combinatorial action of multiple effectors (Fig. 4b,c). A number of chromatin-associated proteins contain more than one reader, in the form of several copies of the same reader or a combination of various readers that are often specific for distinct PTMs. The combinatorial readout in *cis*, where effectors bind PTMs within the same histone tail (Fig. 4b), was initially reported for the TAF1 DBD<sup>94</sup>. Each bromodomain of the TAF1 DBD recognizes one acetyllysine on the same diacetylated H4 peptide. Coincidence detection of two histone PTMs by different effectors has been demonstrated for the PHD-bromodomain cassettes of TRIM24 and TRIM33 (refs. 55,131). The PHD finger of TRIM33 binds to K9me3, and the bromodomain recognizes K18ac on the same H3 peptide<sup>55</sup>. Likewise, TRIM24 concomitantly associates with unmodified H3 and K23ac of the H3 tail through its PHD finger and bromodomain, respectively<sup>131</sup>. The PHD finger and TTD of UHRF1, a key factor for maintenance of DNA methylation, synergistically recognize unmodified H3 and K9me3 of the H3K9me3 peptide<sup>132</sup>. Combinatorial reading

of PTMs by an effector and a catalytic histone-binding domain has been shown for the histone demethylases PHF8 and KIAA1718, where binding of the PHD finger to H3K4me3 in the doubly methylated H3K4me3K9me2 and H3K4me3K27me2 peptides directs association of the catalytic domains with K9me2 and K27me2, respectively<sup>60</sup>. BRPF1/2, a member of the trithorax family with a central role during development, contains two PHD fingers, a PWWP domain and a bromodomain. Although it has not yet been determined whether the interactions occur on the same histone tail, the PWWP domain preferentially binds H3K36me3 and the first PHD domain associates with unmodified H3 (refs. 35,36,133,134).

Multiple PTMs can also be recognized on separate tails of a single nucleosome or on adjacent nucleosomes that are directly linked by DNA or are otherwise in close spatial proximity (Fig. 4c). Despite the widespread phenomenon of multivalent engagement, characterization of combinatorial readout in *trans* poses unique challenges, and only a few examples have thus far been described. BPTF, a subunit of the NURF chromatin-remodeling complex, contains an H3K4me3-specific PHD finger and an acetyllysine-binding bromodomain, connected by a rigid  $\alpha$ -helical linker<sup>40,43</sup>. A modeling study has revealed that the PHD-linker-bromodomain assembly fits complementarily on the surface of a single nucleosome with the PHD finger and bromodomain concomitantly interacting with H3K4me3 and H4K16ac, respectively<sup>135</sup>. More recently, the intranucleosomal engagement of the two readers and coexistence of the H3K4me3 and H4K16ac marks on a single nucleosome within human cells have been confirmed experimentally<sup>107</sup>. Another example of the in *trans*-acting readers is the PHD fingers of CHD4, the interdomain organization of which positions these two modules with a duplicate function to concurrently associate with two H3 tails of a nucleosome<sup>136</sup>.

An even more complex combinatorial *trans* readout can be seen in chromatin complexes containing histone readers in multiple subunits (Fig. 4d). The interplay between such effectors generates a multifaceted network of intertwined contacts that can provide a high degree of specificity. The PHD finger-containing subunits of the HBO1 complex work together to regulate its acetyltransferase activity<sup>137,138</sup>. Similarly, the methyltransferase function of the PRC2 complex is modulated by its subunits harboring WD40 repeats, Tudor and PHD fingers<sup>139</sup>. The SAGA complex, which has a major role in transcription regulation, contains bromodomains, chromodomains and TTDs that bridge the complex to acetyllysine- and H3K4me3-enriched chromatin<sup>140,141</sup>. The MYST HATs are also present in complexes harboring multiple histone readers<sup>142</sup>, including the PHD fingers of BRPF1, ING5 and MOZ<sup>138</sup>. The tumor suppressor Tip60 HAT complex contains subunits with the H3K4me3-specific PHD domain, H3K36me3- and

H3K4me1-binding chromo-barrel domains and two bromodomains linked to diacetylated H4 (refs. 41,61,95,143). Furthermore, many nuclear complexes are not static and undergo component swapping, a powerful mechanism for altering their chromatin-targeting capabilities. Beyond mediating specific chromatin anchoring, PTM cross-talk can also trigger a cascading series of writing, erasing and reading events. For instance, the PIM1 kinase phosphorylates H3K9ac, generating H3K9acS10ph<sup>144</sup>. This mark is recognized by 14-3-3, which recruits the acetyltransferase MOF. MOF, in turn, deposits H4K16ac, a mark targeted by the bromodomain of BRD4, which subsequently recruits P-TEFb, necessary for transcription elongation<sup>144</sup>.

### Concluding remarks

Recognition of PTMs by histone readers has become an important paradigm in chromatin biology. Over the past decade, a wealth of information has been gathered on histone PTMs and PTM-reader relationships, shedding light on the incredibly intricate nature of the chromatin landscape and resultant interactions. The biological consequences of effector-PTM interactions are highly context dependent, relying on the combinatorial readout of the spatially and temporally fluctuating local epigenetic environment and leading to a highly fine-tuned targeting of particular genomic sites.

Recent significant advances in high-throughput technologies allow for the exploration of interactions between readers and peptide fragments of modified histones at a comprehensive level (**Box 1**); however, little is known about the activities of these domains in the context of the nucleosome or chromatin fiber. Moreover, the multifaceted contacts between PTMs and readers, the synergistic or antagonistic nature of their interactions and the engagement of other components of the epigenetic machinery are only beginning to be characterized. Future studies will provide significant insight into the epigenetic regulatory mechanisms and may lead to the discoveries of new pharmacological targets and biomarkers. For example, recently developed

small-molecule inhibitors for the bromodomains of the BET protein family show anticancer activity and demonstrate promising results in preclinical studies<sup>145</sup>. Unlike mutations in the genome, epimutations are largely reversible, and thus epigenetic therapy has a high therapeutic potential. Although in-depth characterization of the epigenetic pathways is necessary to better understand and exploit this potential, the prospective benefits are far reaching.

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The authors declare no competing financial interests.

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### BOX 1 Methodologies for the identification of histone readers

Novel techniques are being developed for rapid high-throughput and discovery-based screening of novel PTM readers and characterization of the effect of adjacent PTMs.

**Histone microarrays**<sup>146–149</sup>. Histone peptides carrying various PTMs are synthesized on cellulose support (SPOT and CelluSpot) or attached to streptavidin-coated platforms by a biotin tag. GST- or His-tagged protein is incubated with the screen, then the bound fraction is detected by western blot or fluorescence. Binding to a single PTM or combinations of PTMs can be assayed in a high-throughput manner; however, this technique requires purification of the protein of interest, and initial generation of the screen can be time consuming.

**Bead-based approach**<sup>150</sup>. This approach utilizes a PTM-randomized combinatorial library of histone peptides. Proteins incubated with the library are captured on beads and their target peptides are analyzed by MS. This method is very powerful in detecting synergistic and inhibitory combinations of known and yet-to-be-discovered PTMs, but it has the same limitations as histone microarrays.

**Stable isotope labeling by amino acids in cell culture (SILAC) technology**<sup>151,152</sup>. Nuclear extracts from cells grown in SILAC medium are incubated with PTM-bearing immobilized histone peptides or recombinant nucleosomes. Bound proteins or protein complexes are pulled down and analyzed by quantitative MS. This approach provides an unbiased way for the identification of new PTM readers; however, the particular protein and domain responsible for the targeting may not be easily identified.



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