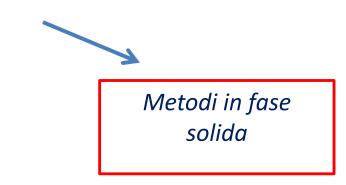
Estrazione Acidi Nucleici

Scelta del protocollo di estrazione dipende da:

- •Tipo di tessuto
- •Resa del metodo
- •Purezza e grandezza degli acidi nucleici isolati
- •Costo
- •Possibilità di automazione



Grandi volumi



Piccoli volumi

Procedimento generale: Fase liquida

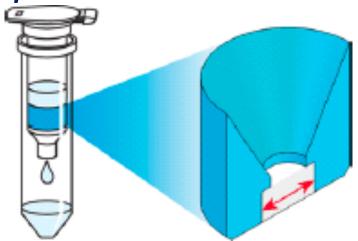


- Lisi cellule: digestione membrane cellulari e nucleari e proteine in generale (Proteasi K)
- Separazione degli acidi nucleici dalle proteine (estrazione con fenolo:cloroformio): fase acquosa/ interfase/fase fenolo:cloroformio
- Precipitazione DNA/RNA (alcoli)
- Quantificazione spettrofotometrica e misura della purezza (A260 nm e A280 nm)

Fase solida

Estrazione con kit che impiegano colonnine con filtri specifici per il legame degli acidi nucleici:

- Digestione con proteinasi
- Lavaggi per eliminazione proteine
- Eluizione DNA



DNA extraction from fresh tissues

§-Tissue homogenization (or cells washing)

- Tissue homogenization or pellet cells and wash it with PBS.

§-Proteinase K digestion

-Put in 10X volume Proteinase K solution with SDS and enzyme. Proteinase K solution: Proteinase K final 1 mg/ml + 10 mM Tris-HCl pH 7.4 + 10 mM EDTA + 150 mM NaCl + 0.4 % SDS (add just before use)

-O.N. incubation at 37-45°C with stirring (minimum 4 hrs).

§-Phenol/Chloroform extraction

-Extraction with phenol-Tris (pH 8)/chloroform (50%/50%) 1:1 in volume.

§-DNA precipitation

-Precipitate aqueous phase w/ 1 volume of isopropanol. Wash w/ 70% ethanol.

§-DNA pellet solubilization

-Recover pellet in H_2O or TE buffer (better) and store at -20° C.

DNA extraction from blood

§- Red cells lysis (blood in citrate, heparin or EDTA) -Add 45 ml lysis buffer to 5 ml blood, mix by inversion and leave at RT for 5'. Centrifuge X 1000 g at 4°C for 6'.

Red cells lysis buffer: 0.32 M saccarose + 10 mM tris-HCl pH 7.5 + 5 mM MgCl₂ + 1% triton X 100. -Wash the pellet with PBS and centrifuge as before.

§- Proteinase K digestion

-Recover pellet in 600 µl of buffer A and add SDS final 0.5% and proteinase K 1 mg/ml. Incubation at 37°C O.N. (at least 4 hrs). Buffer A: 10 mM Tris-HCl pH 8 + 400 mM NaCl + 2 mM EDTA.

§- Protein salting-out

-Add 220 μ I of sature solution of NaCl, vigorous stirring for 15" and centrifuge 15' X 14000 rpm at 4° C.

\S - DNA precipitation

-Recover supernatant and precipitate with 1 volume isopropanol at rt with immediate precipitation. Centrifuge and wash pellet with 1 ml of 70% ethanol.

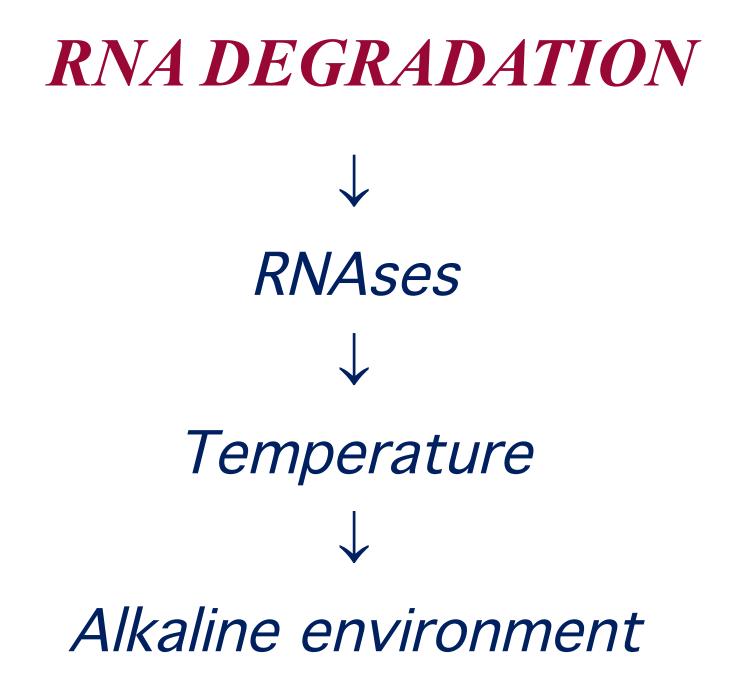
§- DNA pellet solubilization

-DNA is solubilized in 500 μ l of TE or H₂O.

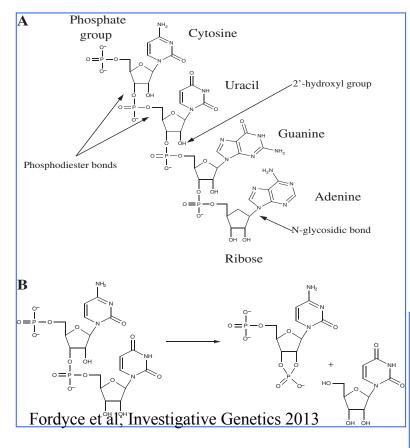
DNA extraction from blood- silica use

- 1. 200 μ L whole blood mixed with 100 μ L lysis buffer^a or 25 μ L mononuclear cells were added to 100 μ L binding solution^b. Mix and incubate for 3 min at room temperature. Centrifuge for 15 s at 65× g and discard supernatant.
- 2. Add 100 μ L lysis buffer and mix. Centrifuge for 15 s at 65× g and discard supernatant. Repeat once.
- 3. Add 100 μ L washing solution^c and mix. Centrifuge for 15 s at 65× *g* and discard supernatant. Repeat once.
- 4. Add 100 μ L absolute ethanol and mix. Centrifuge for 15 s at 65× g. Discard supernatant and vacuum-dry the pellet.
- 5. Add 50 μ L elution buffer^d, resuspend the pellet, and incubate for 3 min at 65°C. Centrifuge for 1 min at 65× *g*, and transfer supernatant in a clean tube.

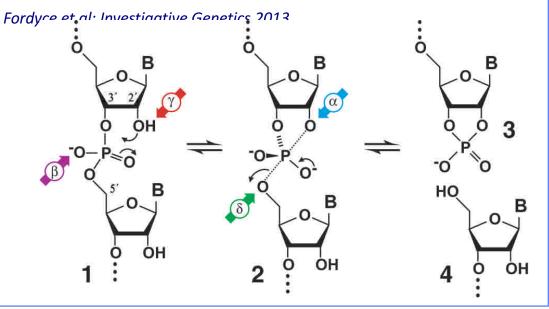
^a3 M guanidine thiocyanate, 20 mM EDTA, 10 mM Tris-HCl, pH 6.8, 40 mg/mL Triton[®] X-100, 10 mg/mL DTT
^b40 mg/mL silica (Sigma) directly suspended in the lysis buffer
^c25% absolute ethanol, 25% isopropanol, 100 mM NaCl, 10 mM Tris-HCl, pH 8.0
^d10 mM Tris-HCl, pH 8.0, 1 mM EDTA
All solutions were prepared in double-distilled water and sterilized by filtration and remain stable for more than six months.



RNA degrades differently from DNA



- The 2'-OH allows the RNA molecule to be more easily degraded via hydrolysis than DNA.
- The phosphodiester bond in RNA can be broken during hydrolysis.
- *The N-glycosidic bond is stronger in RNA than DNA*
- The chemical process of hydrolysis, where the 2'-hydroxyl group has attacked the adjacent phosphodiester bond, cleaving the backbone of the RNA.



RNAse A increases the rate of RNA cleavage by internal phosphoester transfer

3-RNA extraction from fresh tissues

§-Tissue homogenization (or cells washing)

 $\operatorname{\mathsf{\S-Cell}}\mathsf{lysis}$

§-Phenol/Chloroform extraction

$\ensuremath{\S}\xspace$ -RNA precipitation

$\ensuremath{\S}\xspace$ -RNA pellet solubilization

* Chomczynski P; Sacchi N. Single-step method of RNA isolation by acid guanidinium thiocyanate-phenol-chloroform extraction. Anal Biochem, 162: 156-159; 1987

3-RNA extraction from fresh tissues

§-Tissue homogenization (or cells washing)

-For tissues homogenization in liquid nitrogen / wash the cells with PBS (keep always in ice and use ice cold solution).

§-Cell lysis

-Put it in D solution* (1 ml for 20 million cells) with ß-mercapto-ethanol (0.36 ml for 5 ml solution) (1 ml per 2.0 10⁷ cells) for 5' in ice.

*D solution: 250 g guanidinium thiocyanate + 293 ml of DEPC-H₂O + 17.6 ml of sodium citrate 0.75 M pH 7.0 + 26.4 ml sarcosyl 10% at 65°C.

§-Phenol/Chloroform extraction

-Add 1/10 volume of Sodium Acetate 2M with pH 4.0-4.5. (acid pH is necessary to obtain a good separation at the interface) and extract w/ 1 volume of phenol/H₂O and chloroform (7:3 ratio). Keep in ice for 20' and centrifuge for 20'.

§-RNA precipitation

-Add 1 volume of isopropanol, ON at -20°C -centrifuge -Wash w/ chilled 75% EtOH.

§-RNA pellet solubilization

-Resuspend in DEPC H₂O.

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