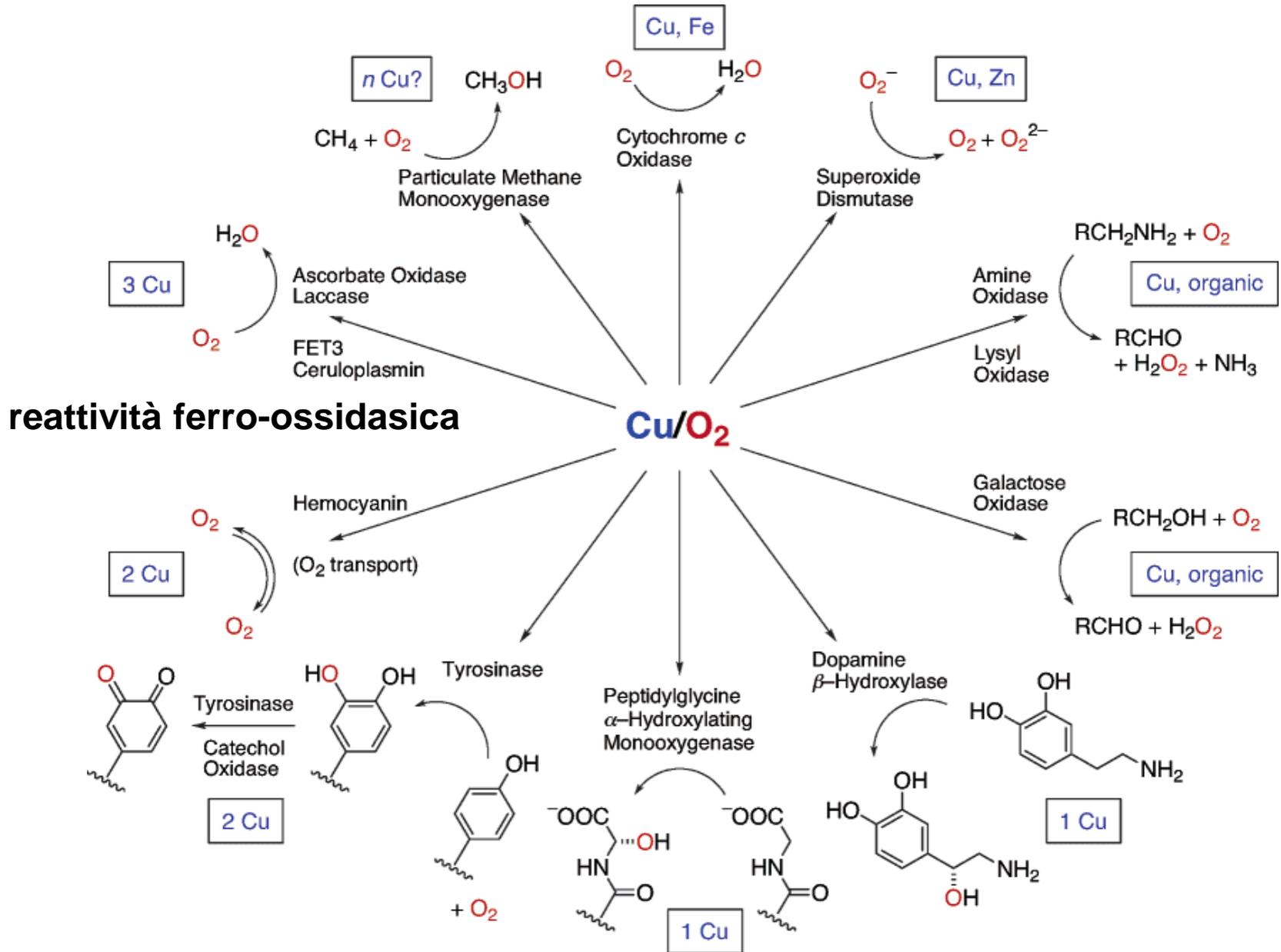
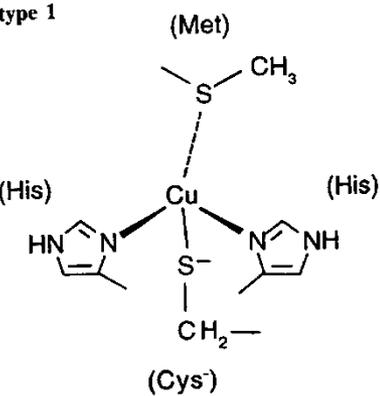
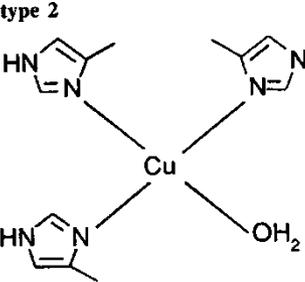
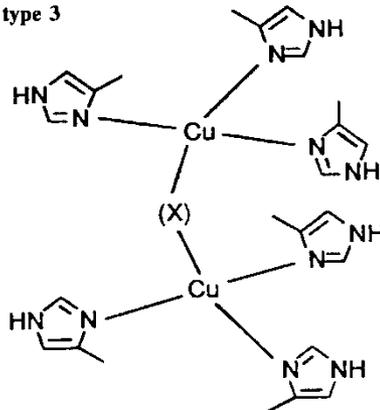


# Proteine al rame



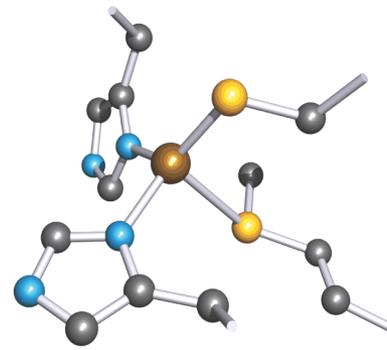
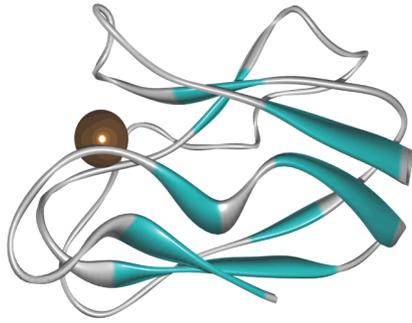
**Table 10.2** Characteristics of 'classical' copper centers in protein

generalized coordination geometry	function, structure, characteristics
<p><b>type 1</b></p> 	<p><b>type 1:</b> 'blue' copper centers  function: reversible electron transfer  <math>\text{Cu}^{\text{II}} + e^- \rightleftharpoons \text{Cu}^{\text{I}}</math>  structure: strongly distorted, (3+1) coordination  absorption of the copper(II) form at about 600 nm, molar extinction coefficient <math>\epsilon &gt; 2000 \text{ M}^{-1}\text{cm}^{-1}</math>; LMCT transition <math>\text{S}(\text{Cys}^-) \rightarrow \text{Cu}^{\text{II}}</math>  EPR/ENDOR of the oxidized form: small <math>^{63,65}\text{Cu}</math> hyperfine coupling and g anisotropy, interaction of the electron spin with <math>-\text{S}-\text{CH}_2^-</math>; <math>\text{Cu}^{\text{II}} \rightarrow \text{S}(\text{Cys})</math> spin delocalization</p>
<p><b>type 2</b></p> 	<p><b>type 2:</b> normal, 'non-blue' copper  function: <math>\text{O}_2</math> activation from the <math>\text{Cu}^{\text{I}}</math> state in cooperation with organic coenzymes  structure: essentially planar with weak additional coordination (Jahn-Teller effect for <math>\text{Cu}^{\text{II}}</math>)  typically weak absorptions of <math>\text{Cu}^{\text{II}}</math>, <math>\epsilon &lt; 1000 \text{ M}^{-1} \text{ cm}^{-1}</math>; ligand-field transitions (<math>d \rightarrow d</math>)  normal <math>\text{Cu}^{\text{II}}</math> EPR</p>
<p><b>type 3</b></p> 	<p><b>type 3:</b> copper dimers  function: <math>\text{O}_2</math> uptake from the <math>\text{Cu}^{\text{I}}\text{-Cu}^{\text{I}}</math> state  structure: (bridged) dimer, Cu-Cu distance about 360 pm after <math>\text{O}_2</math> uptake intense absorptions around 350 and 600 nm, <math>\epsilon \approx 20000</math> and <math>1000 \text{ M}^{-1}\text{cm}^{-1}</math>; LMCT transitions <math>\text{O}_2^{2-} \rightarrow \text{Cu}^{\text{II}}</math>  EPR-inactive <math>\text{Cu}^{\text{II}}</math> form (antiferromagnetically coupled <math>d^9</math> centers)</p>

## Plastocianina

(da spinaci)

10.5 kDa,  
ca. 100 a.a.



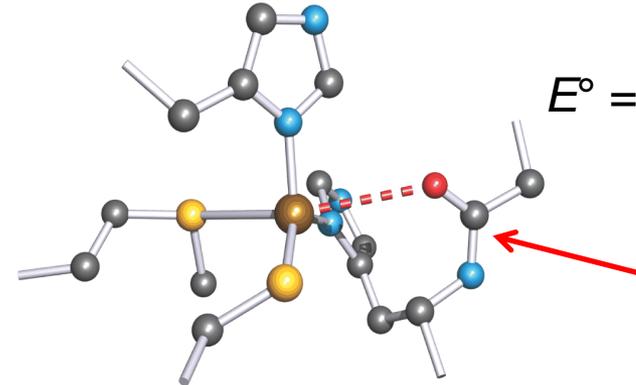
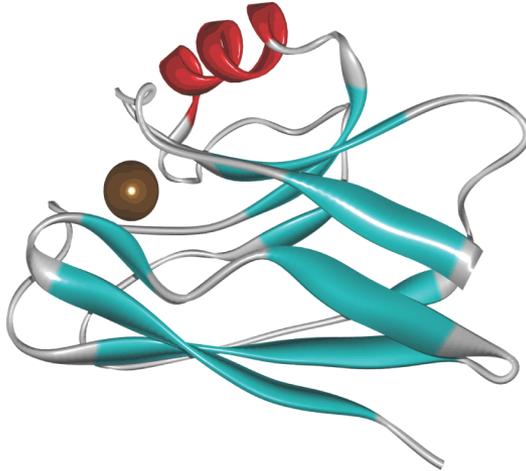
$$E^{\circ} = + 370 \text{ mV}$$

Coordinazione 3 + 1

## Azurina

(da batteri)

14.5 kDa,  
ca. 130 a.a.



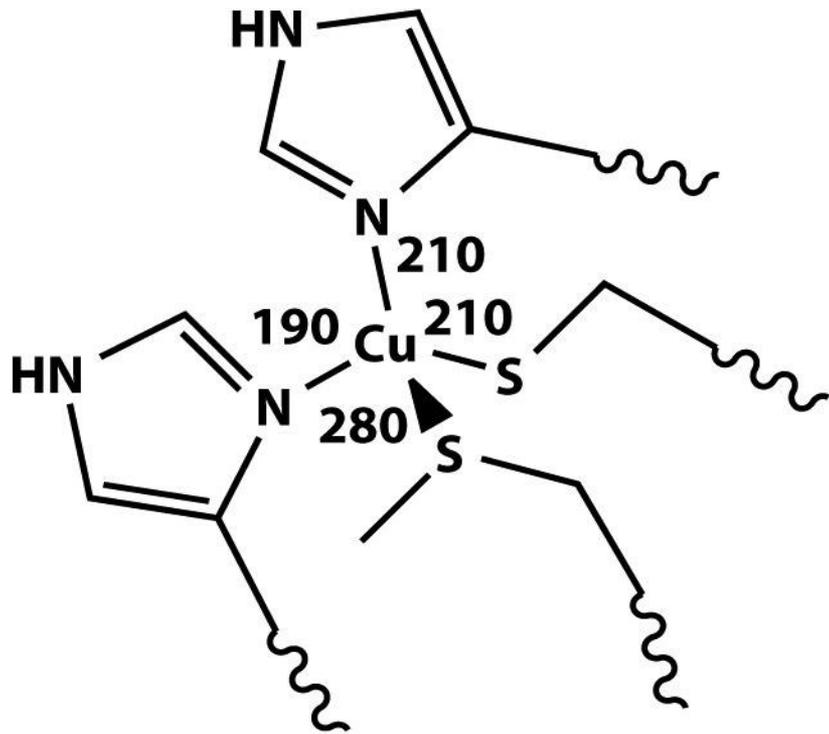
$$E^{\circ} = + 308 \text{ mV}$$

glicina

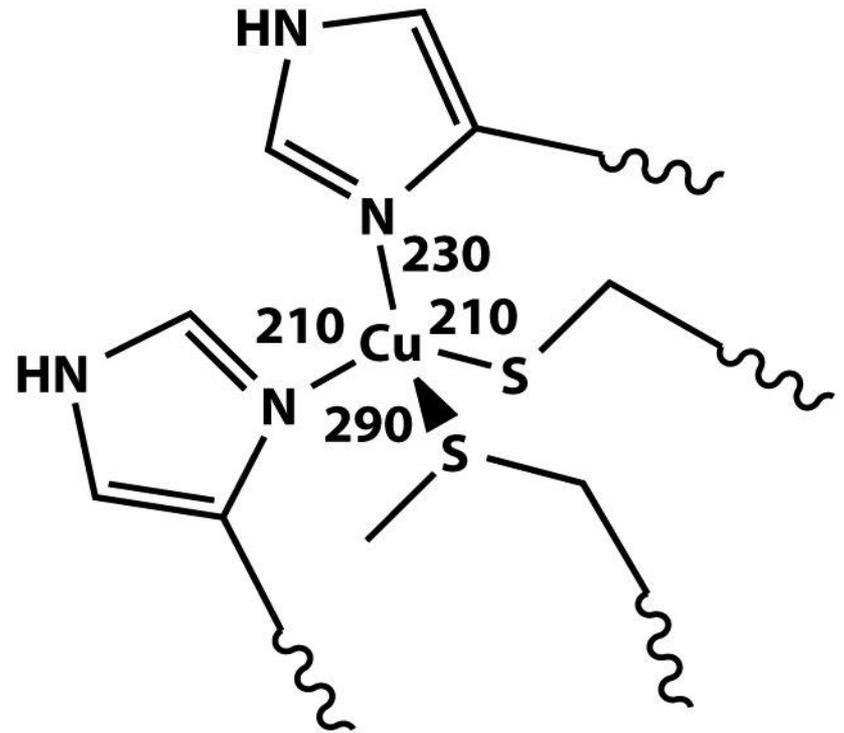
Coordinazione 3 + 1 + 1

*Blue copper proteins*

# Esempio di stato entatico



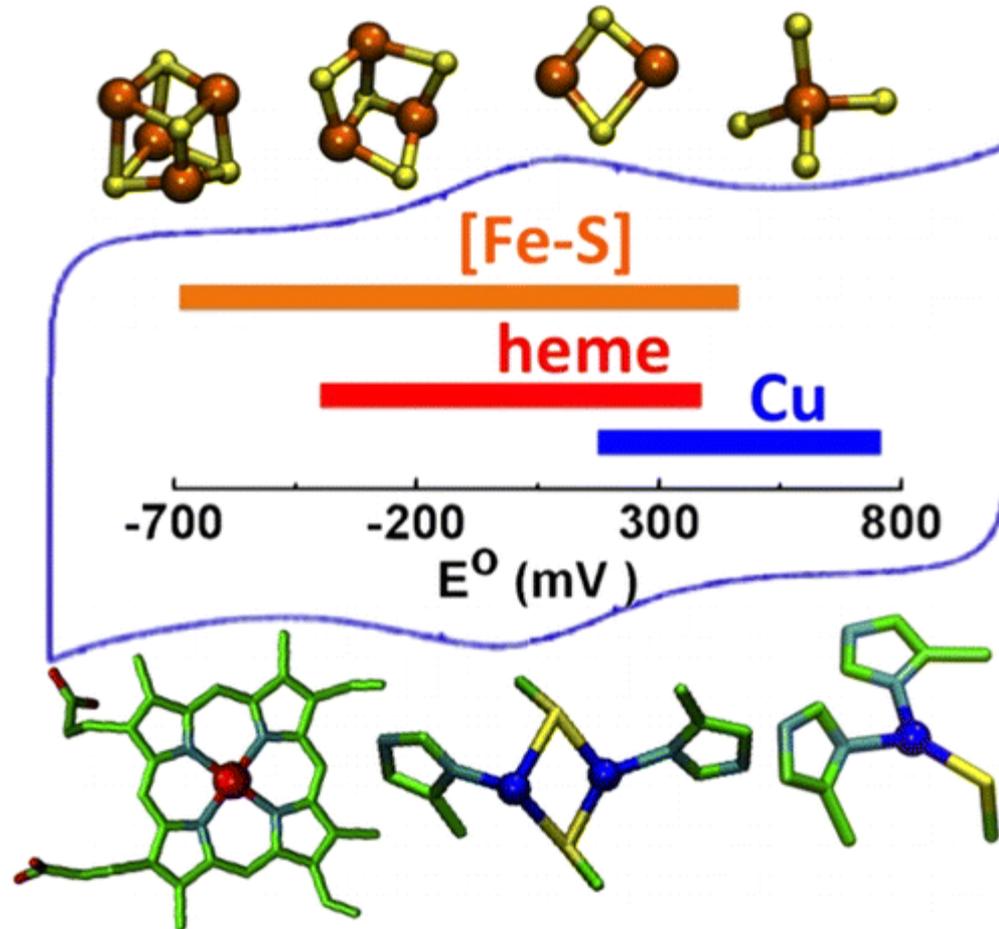
**Oxidized plastocyanin**



**Reduced plastocyanin**

*velocità di trasferimento elettronico dell'ordine  $10^3$ – $10^7$   $M^{-1} s^{-1}$   
(rispetto a  $5 \times 10^{-7} M^{-1} s^{-1}$  per la coppia  $Cu(II)/Cu(I)$  acquosa)*

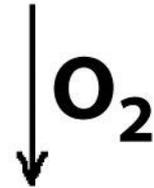
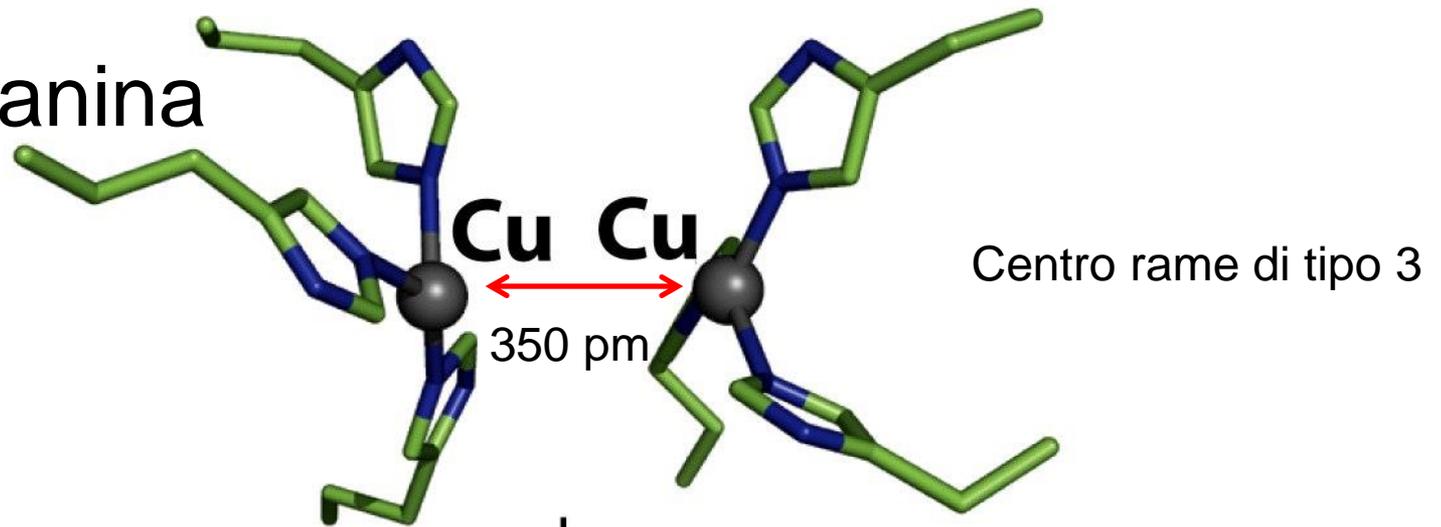
# The electron-transfer systems



# Emocianina

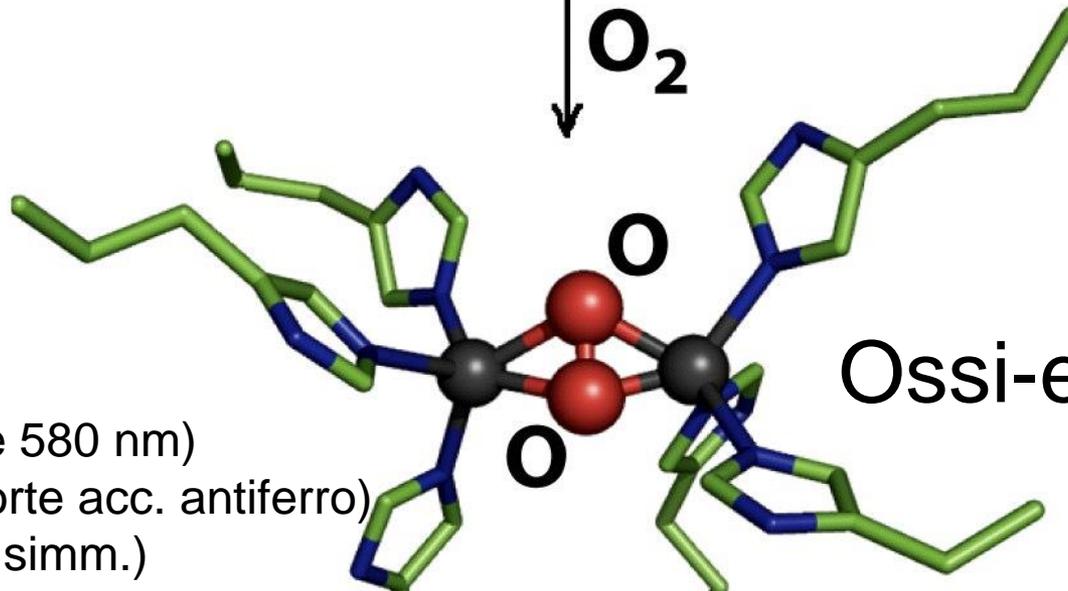
fino a 1500 kDa,  
ogni unità 75 kDa

Incolore  
2 Cu(I) ( $S = 0$ )



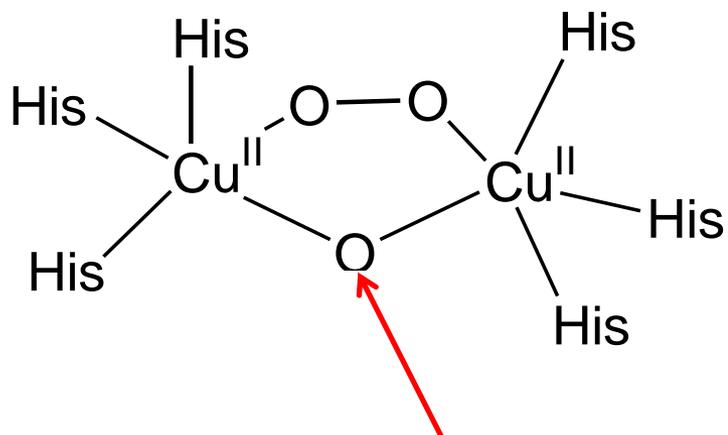
# Ossi-emocianina

Viola (LMCT 350 e 580 nm)  
2 Cu(II) (diamg., forte acc. antiferro)  
IR:  $755\text{ cm}^{-1}$  ( $O_2^{2-}$ , simm.)

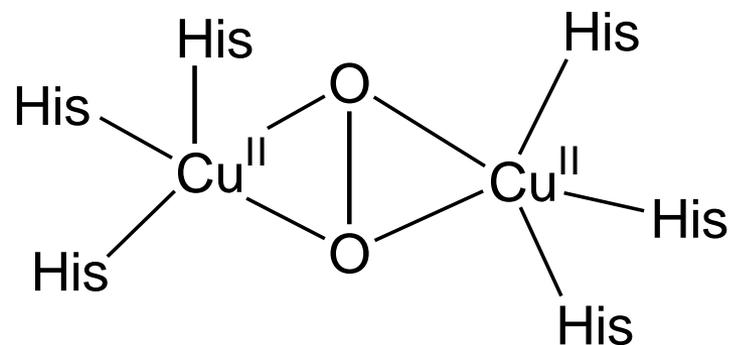


trasporto di  $O_2$  di molluschi (lumache, calamari) e artropodi  
(granchi, aragoste, gamberi, scorpioni)

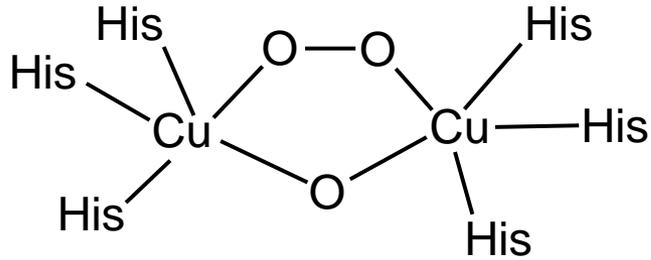
# Possibili coordinazioni simmetriche dello ione perossido



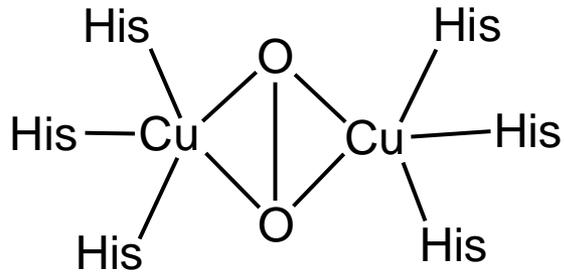
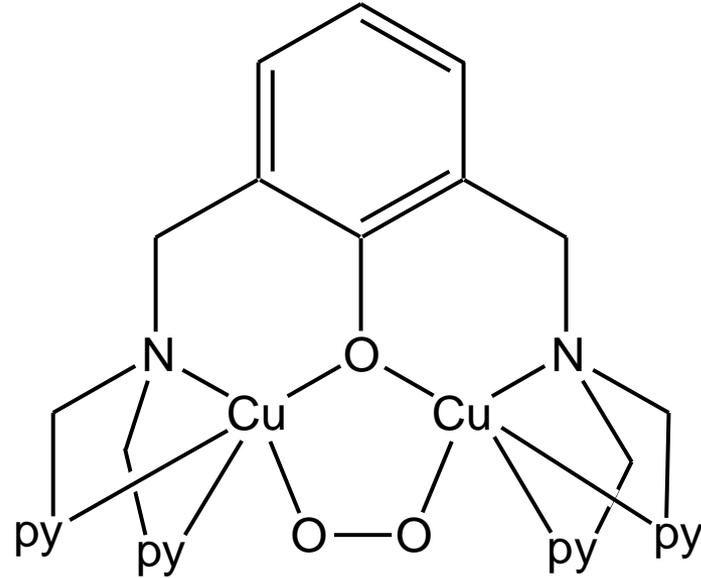
tirosinato?



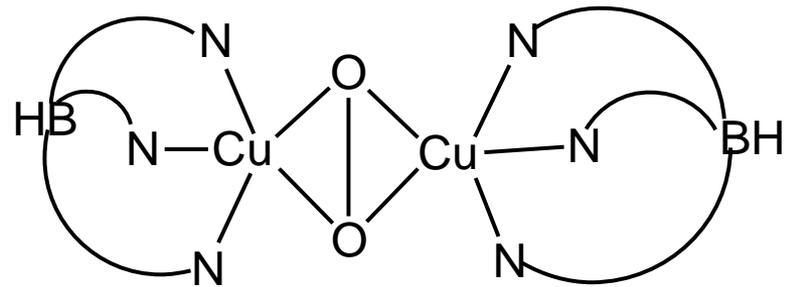
# Modelli per la coordinazione di $O_2$ alla emocianina



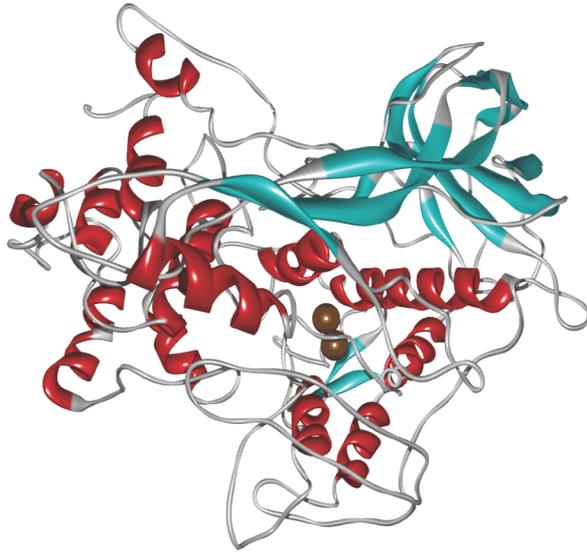
ipotesi sbagliata



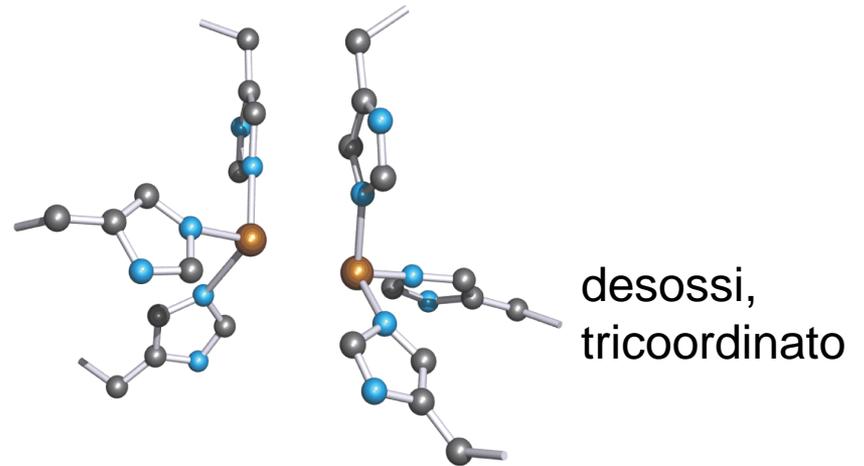
ipotesi corretta



# Emocianina

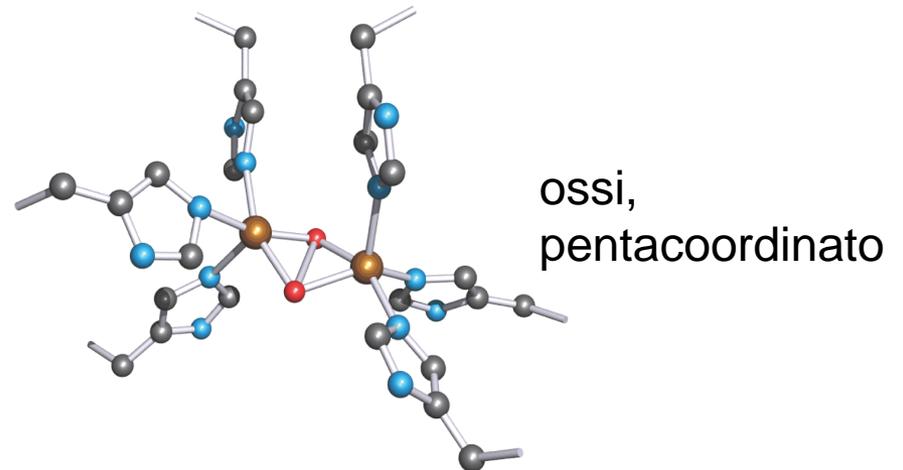


(a)



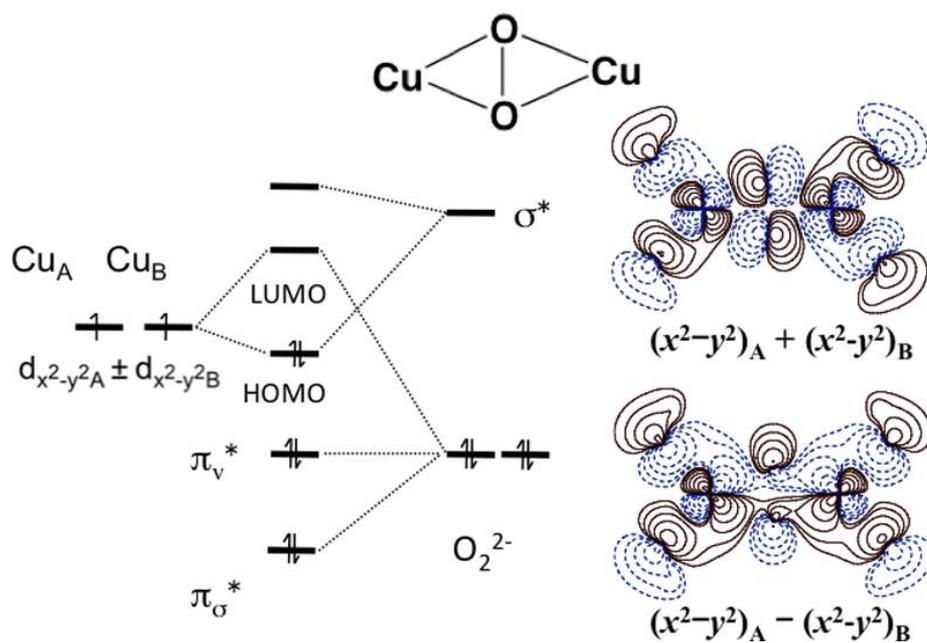
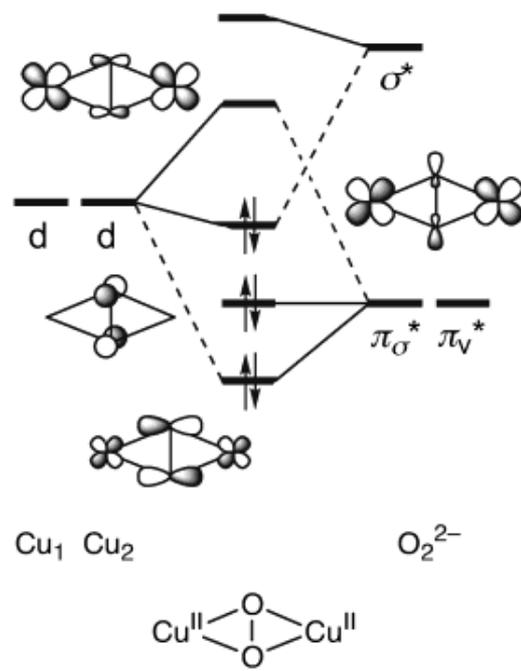
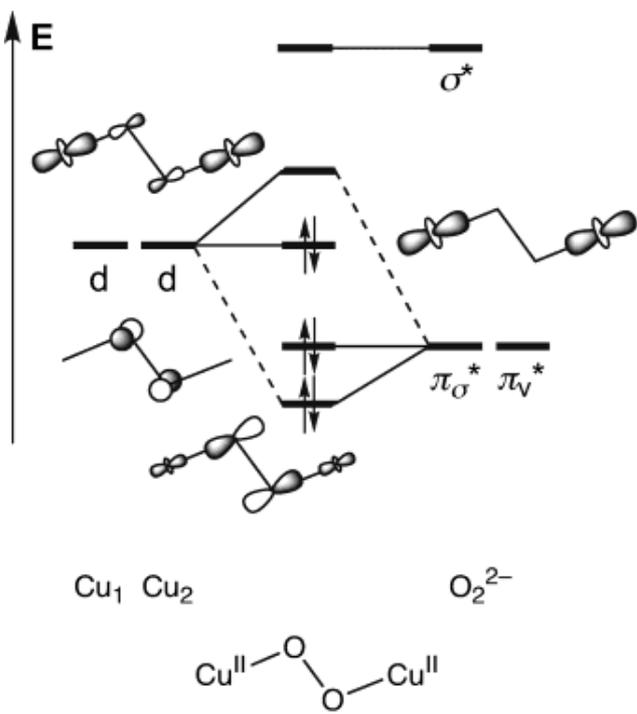
desossi,  
tricoordinato

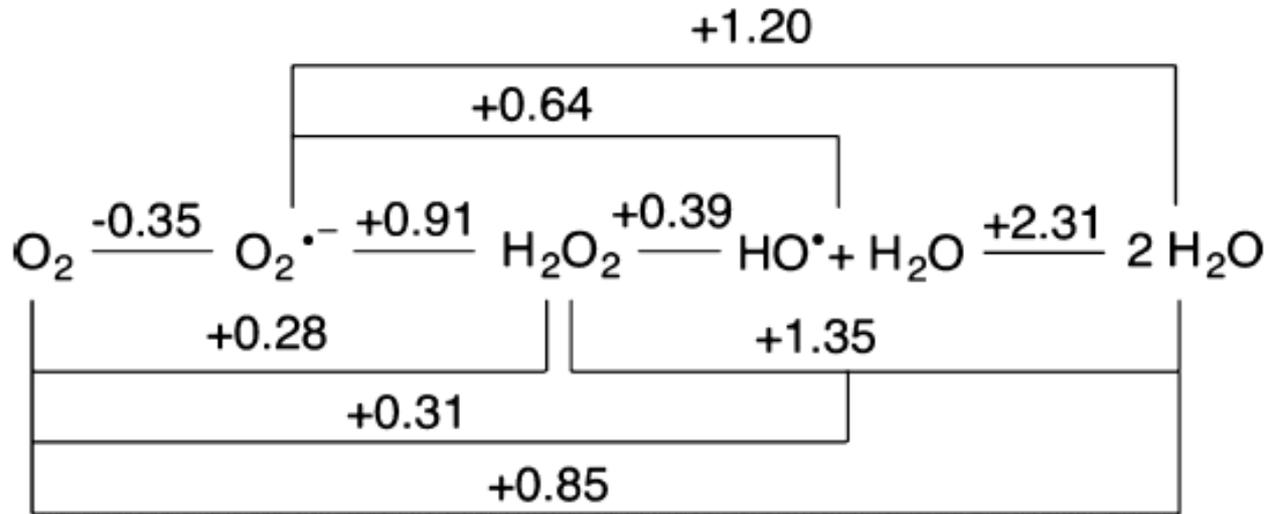
(b)



ossi,  
pentacoordinato

(c)



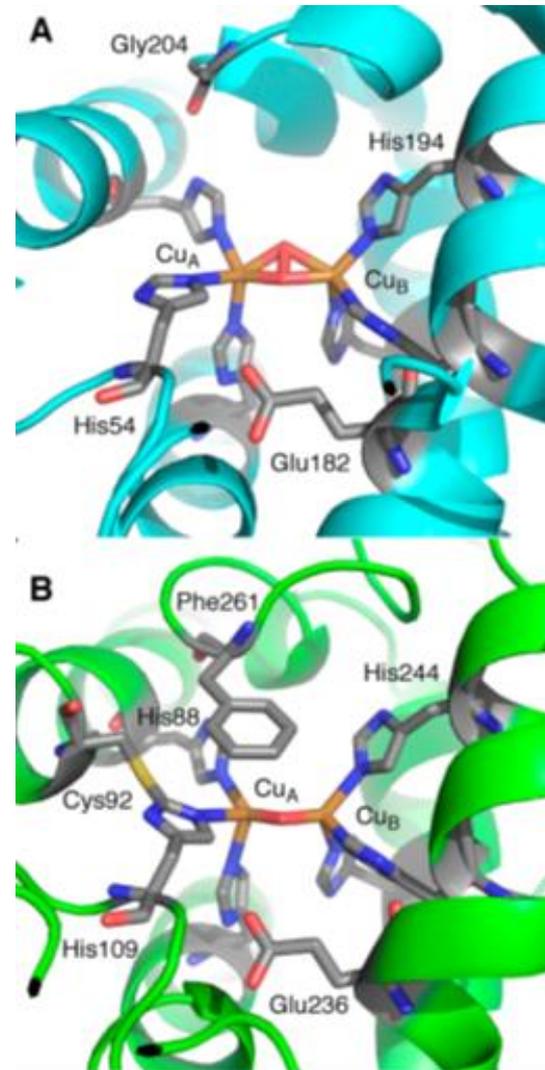
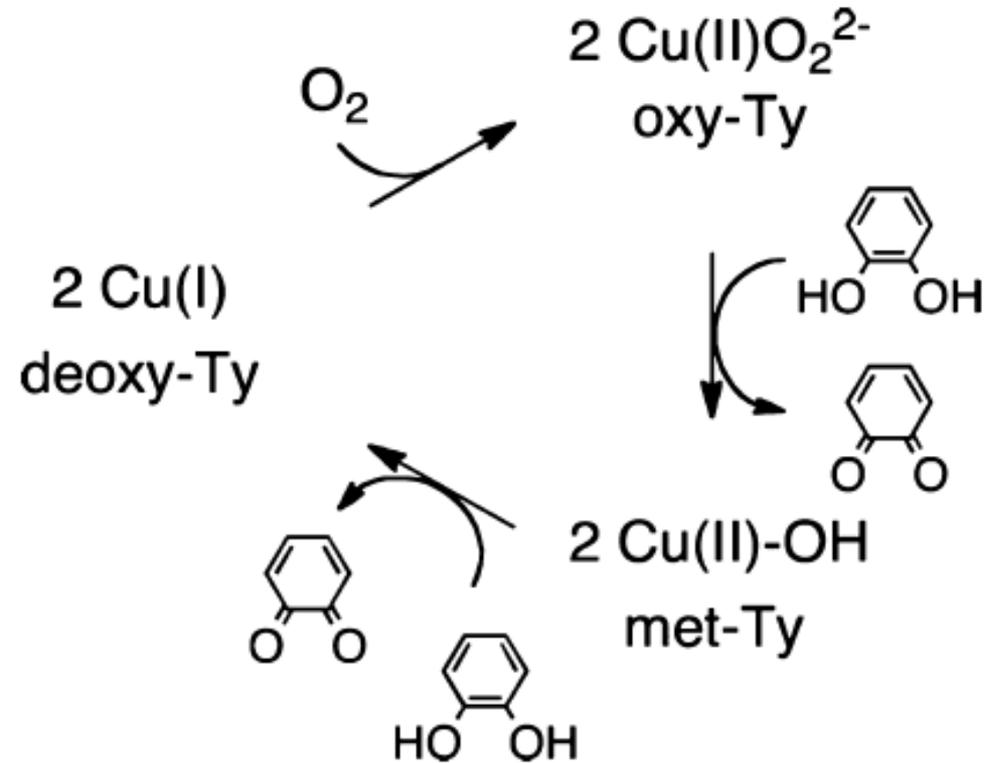
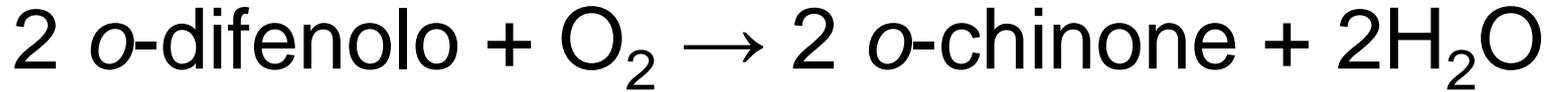


*...la riduzione mono-elettronica di  $O_2$  a superossido è termodinamicamente sfavorita*

*fornire due elettroni richiede o la presenza di più ioni Cu oppure di un Cu e di un cofattore organico redox-attivo*

# Polifenolo ossidasi

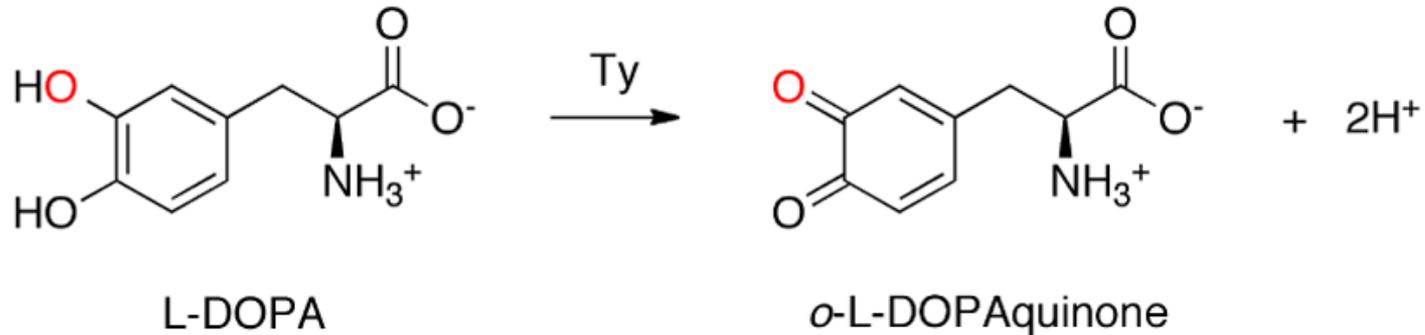
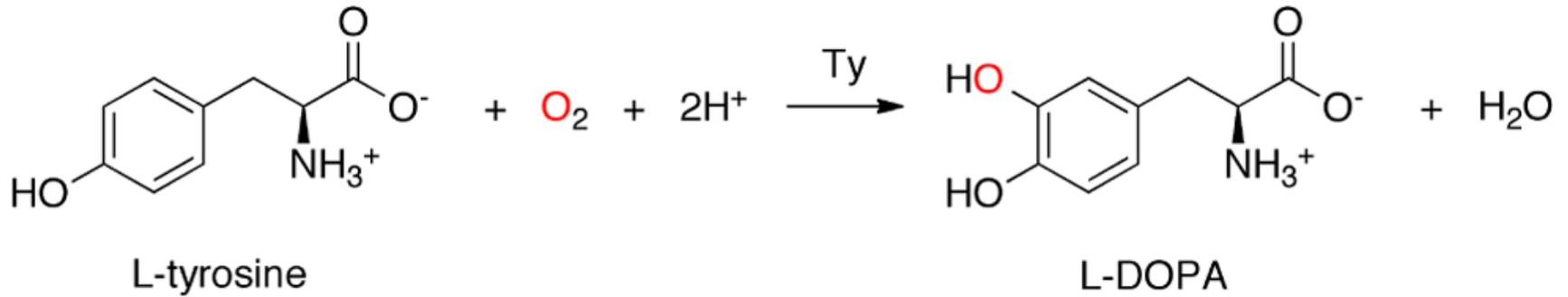
*Tirosinasi, Catecolo-ossidasi*

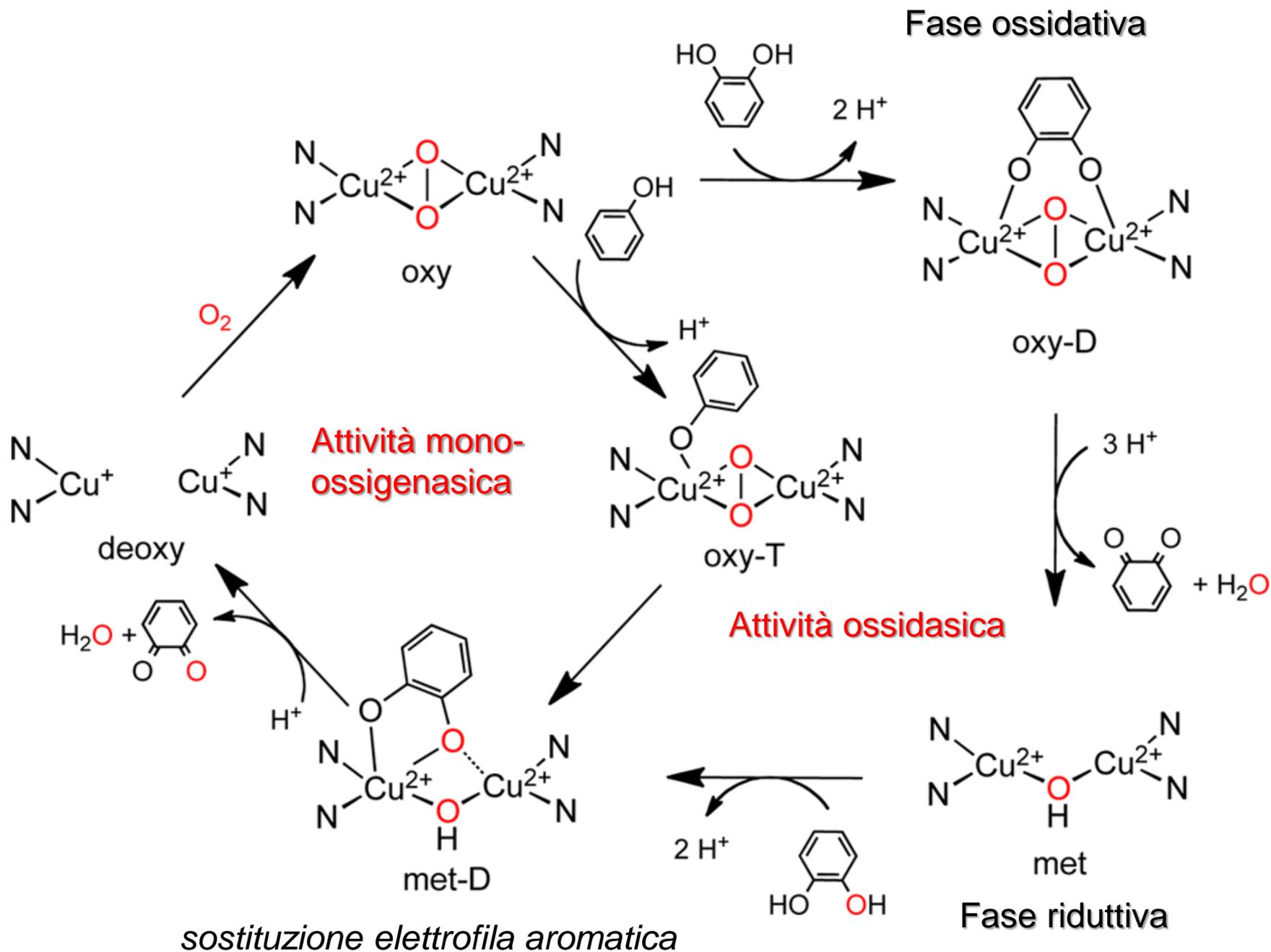


oxy-Ty

met-Ty

# Tirosinasi come mono-ossigenasi

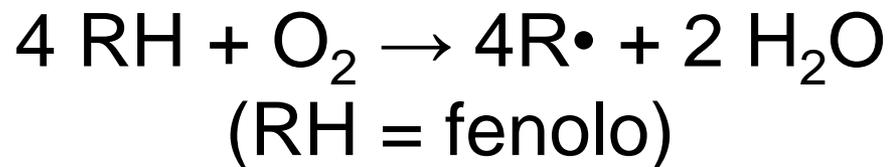


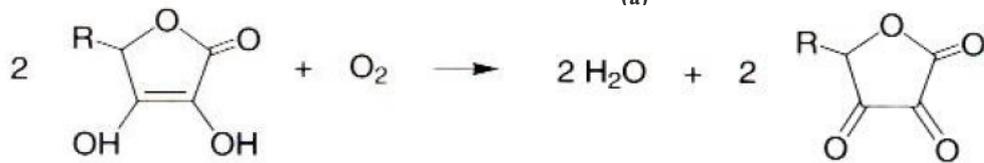


*Multicopper oxidases, MCOs*  
(ossidasi azzurre)

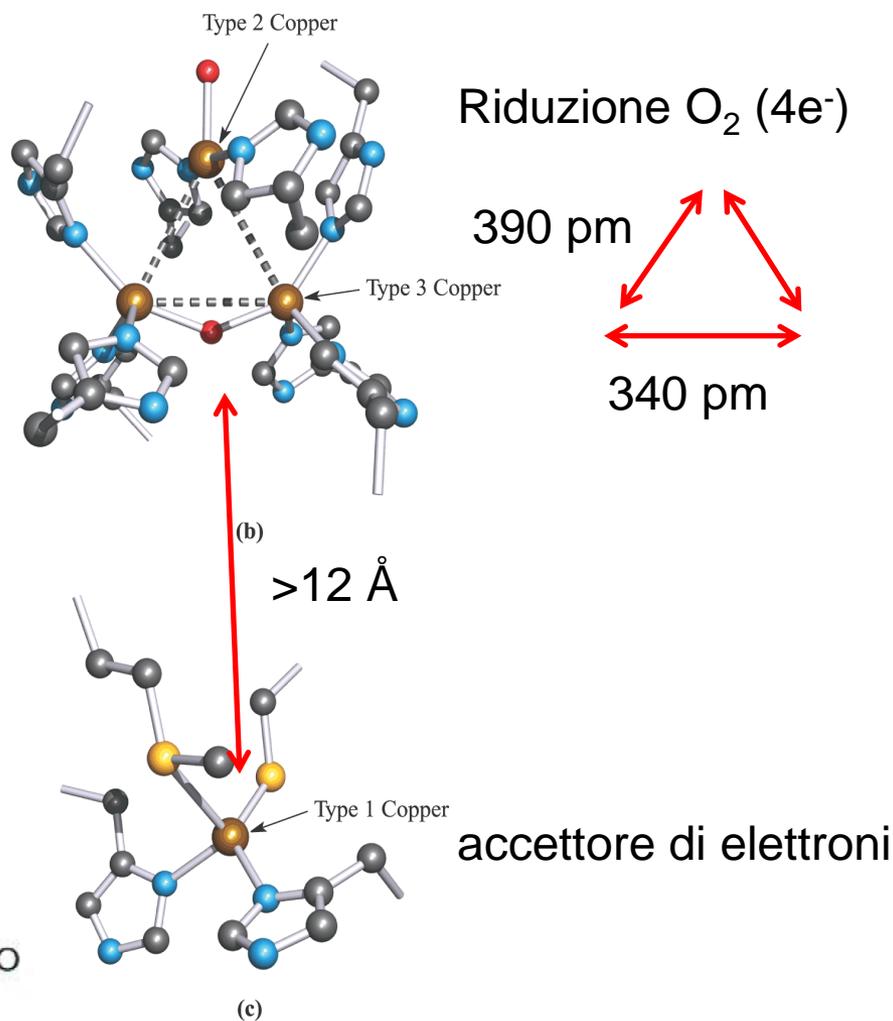
Ascorbato ossidasi  
Laccasi

Ceruloplasmina  
Epeptina  
Fet3p  
Rame-ossidasi (CueO)

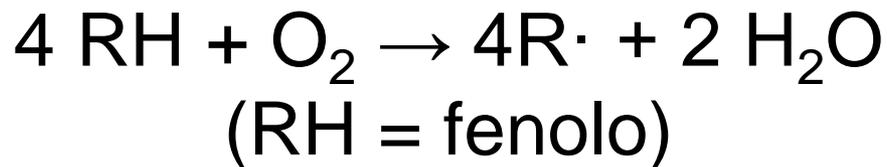


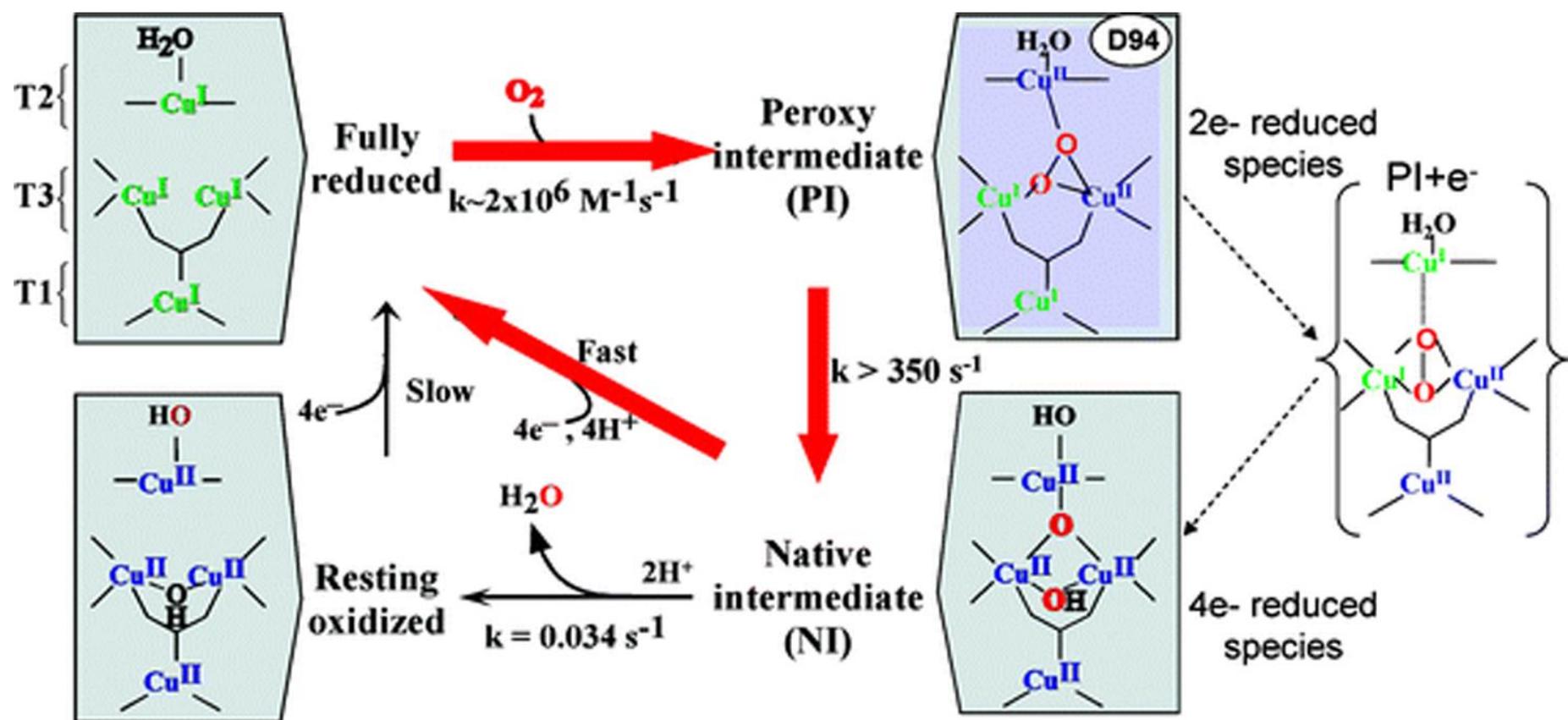


(a)

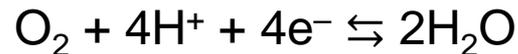
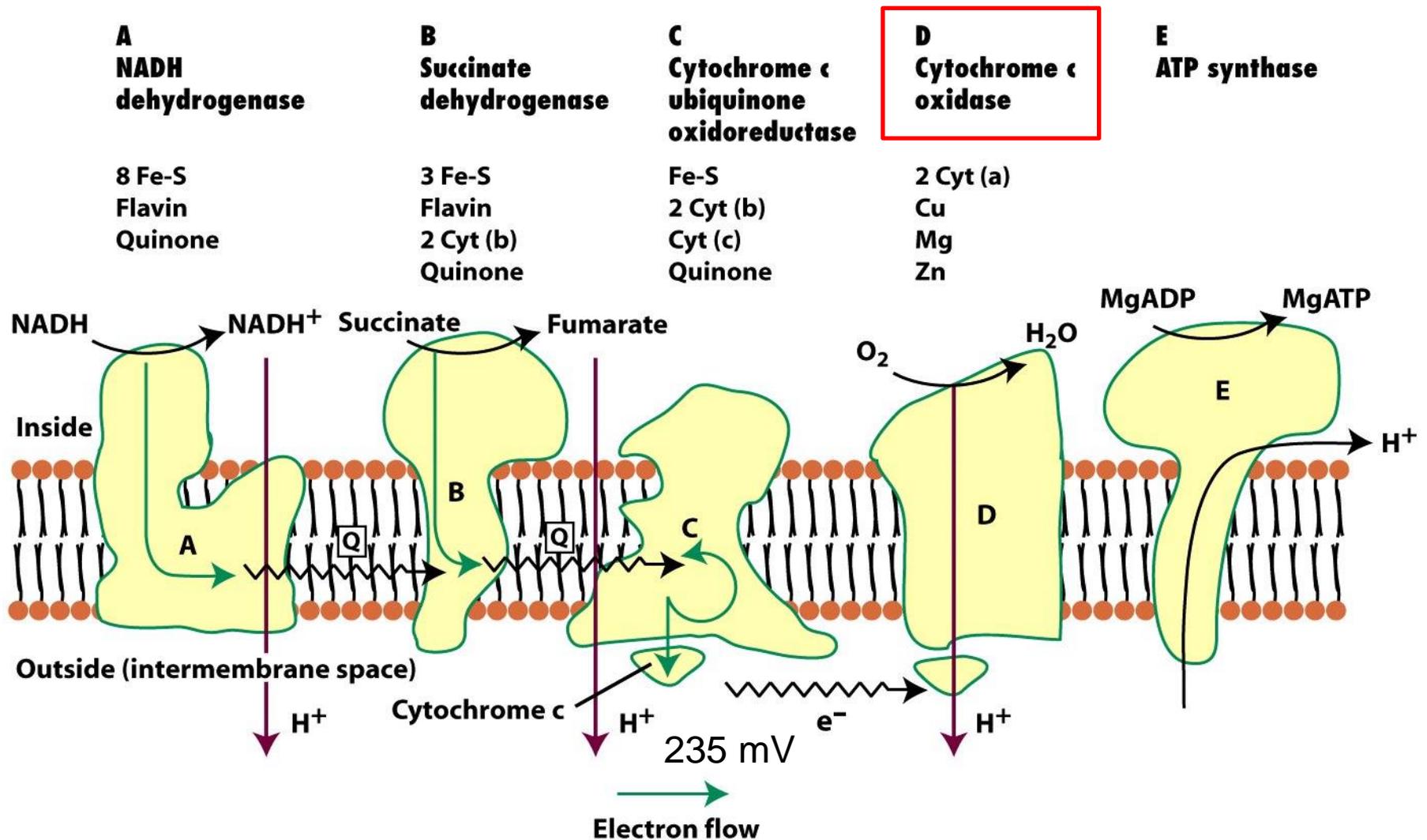


## Ascorbato ossidasi (da zucchini)



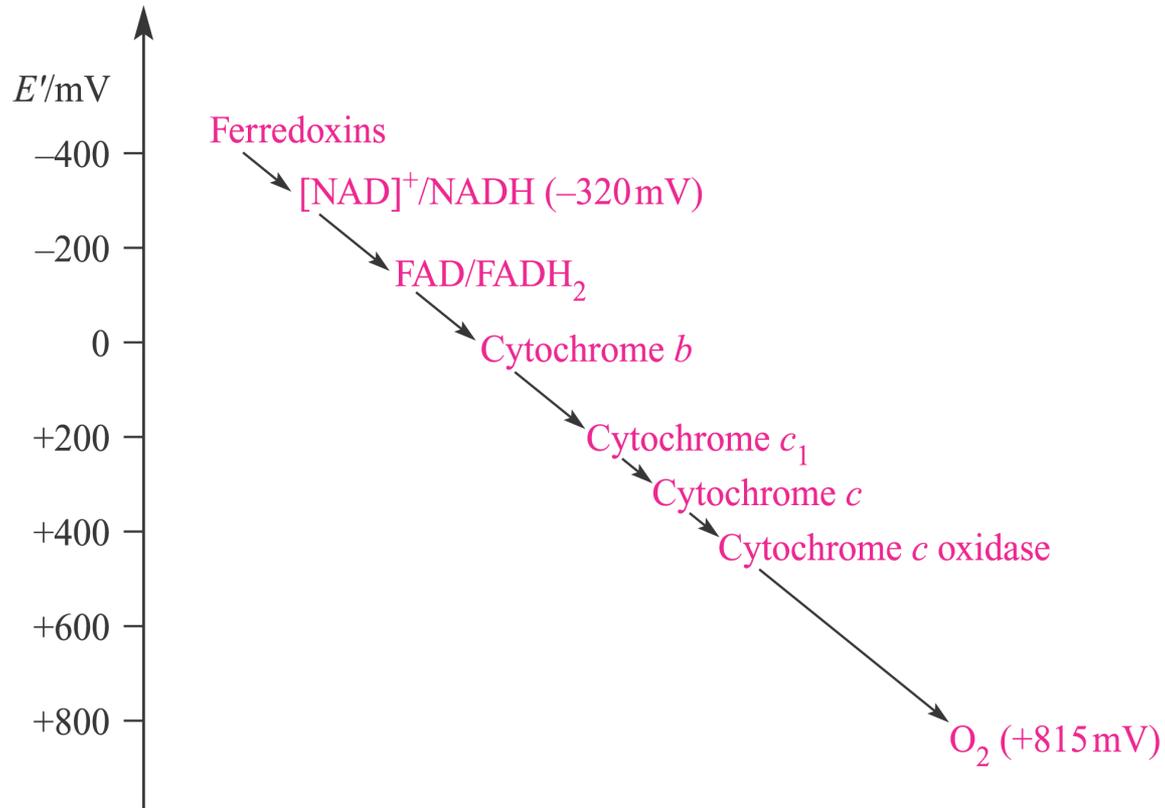


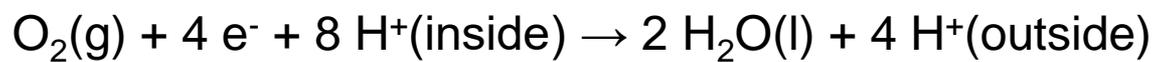
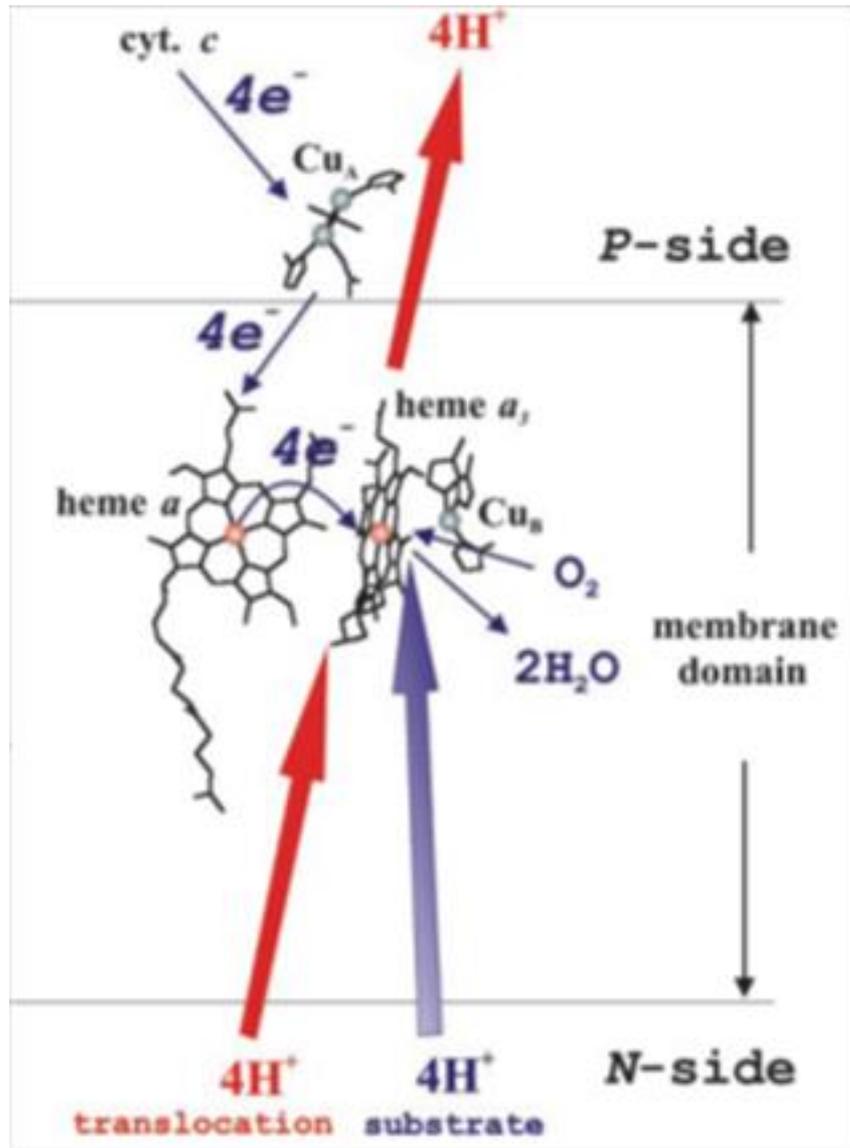
# Catena respiratoria (fosforilazione ossidativa)



$$\Delta E^\circ (\text{pH } 7) = 815 \text{ mV}; \Delta G^\circ = -80 \text{ kcal/mol}$$

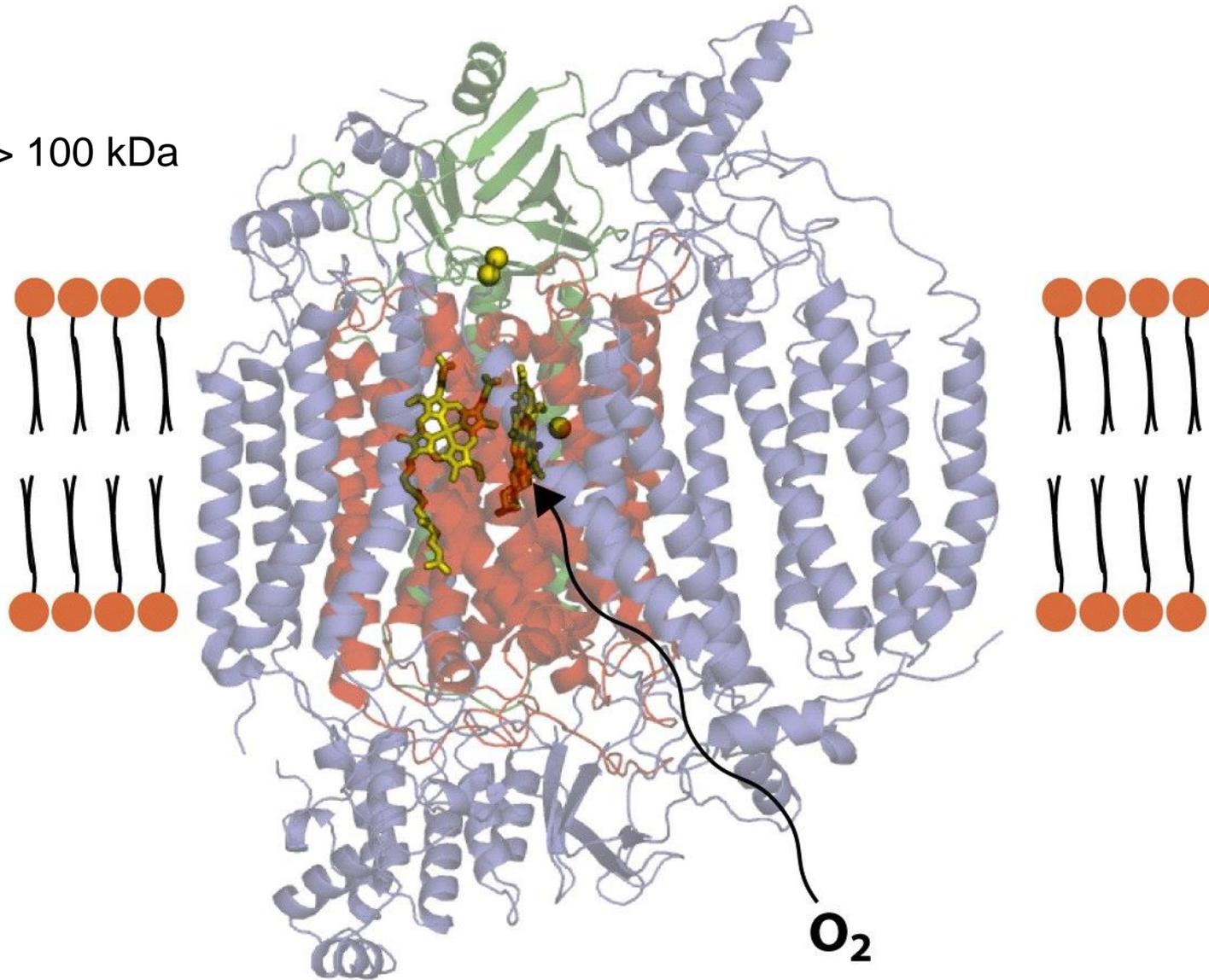
# La sequenza di trasferimenti elettronici nei mitocondri



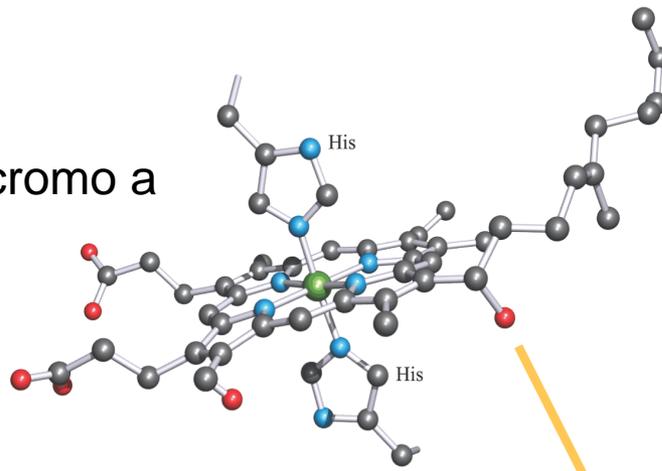


# Reacts with cytochrome C

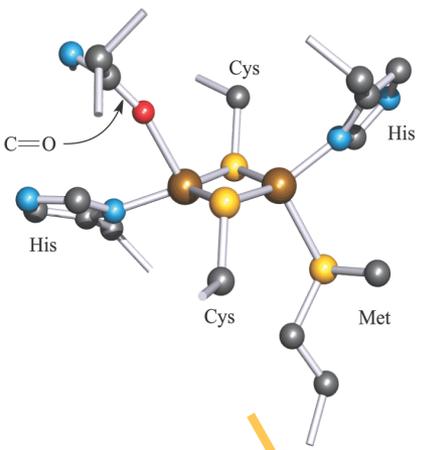
> 100 kDa



citocromo a

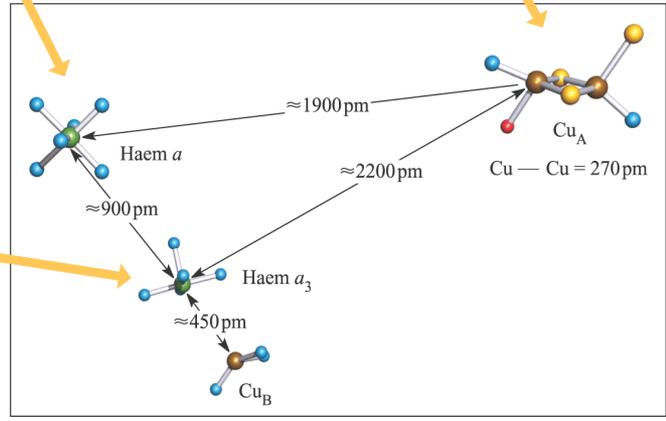
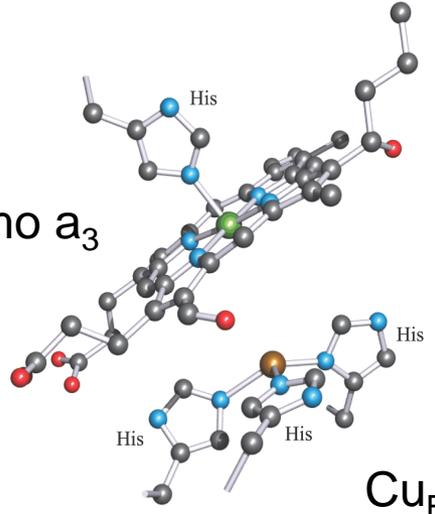


Peptide C=O  
of Gly

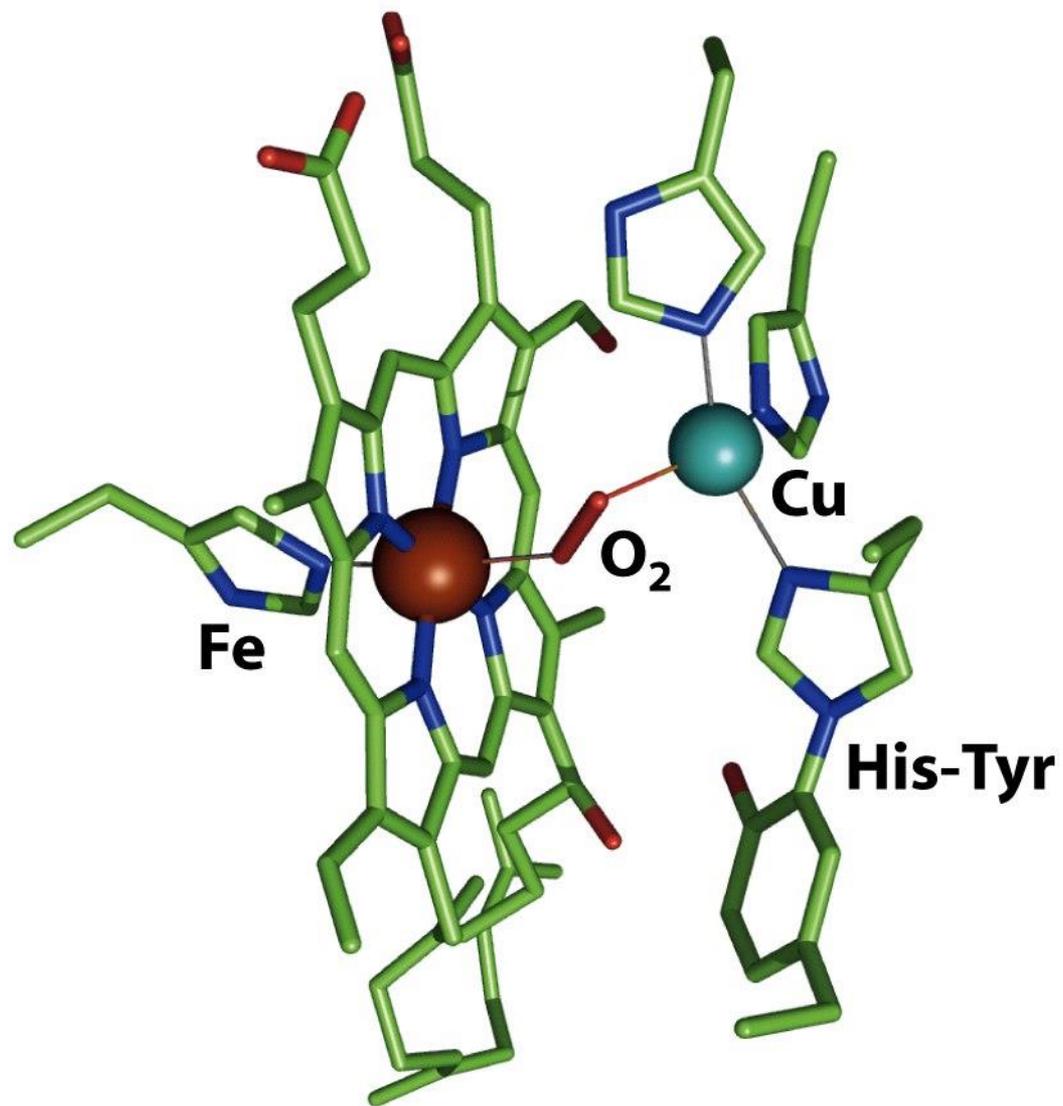


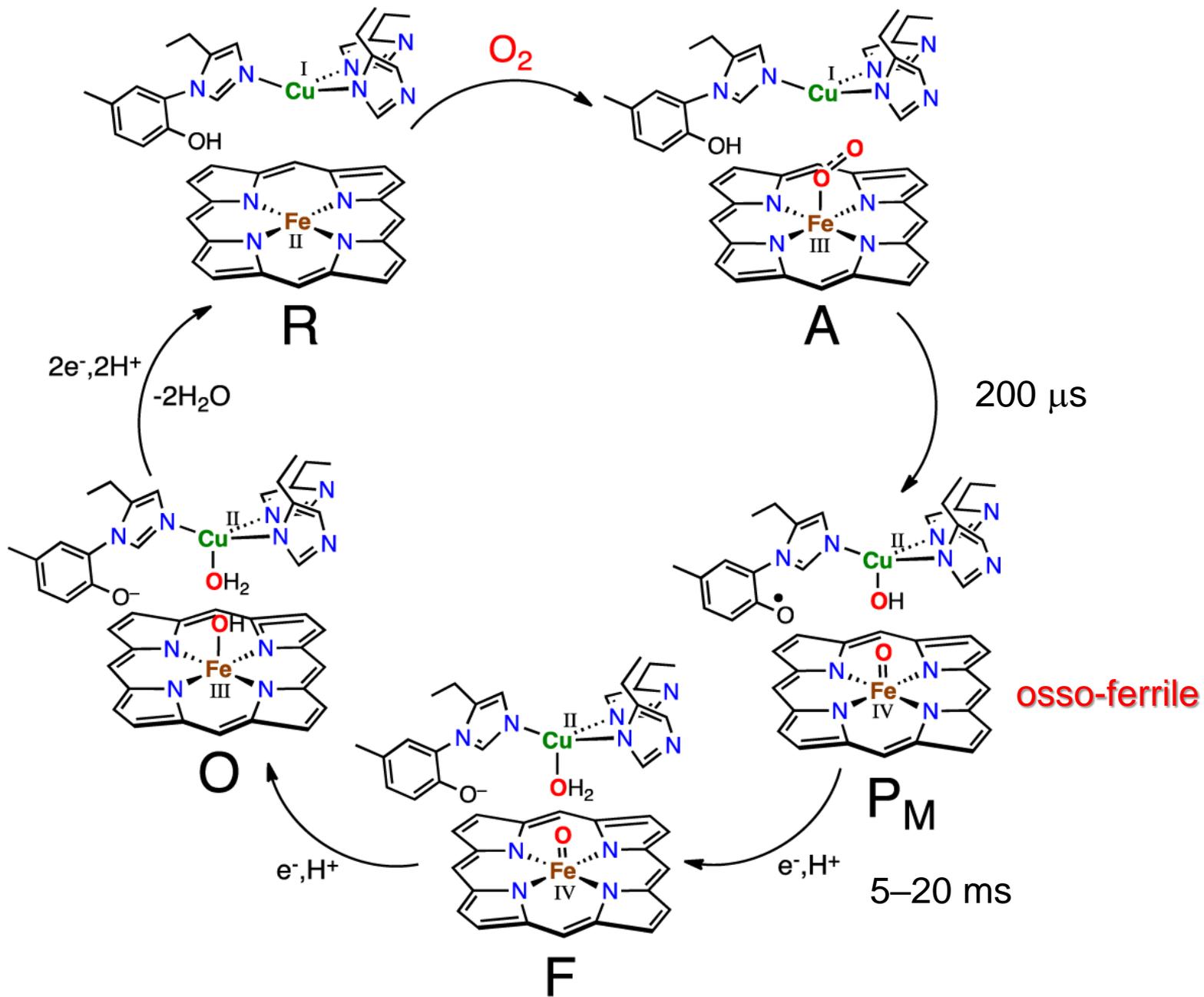
Cu<sub>A</sub>

citocromo a<sub>3</sub>



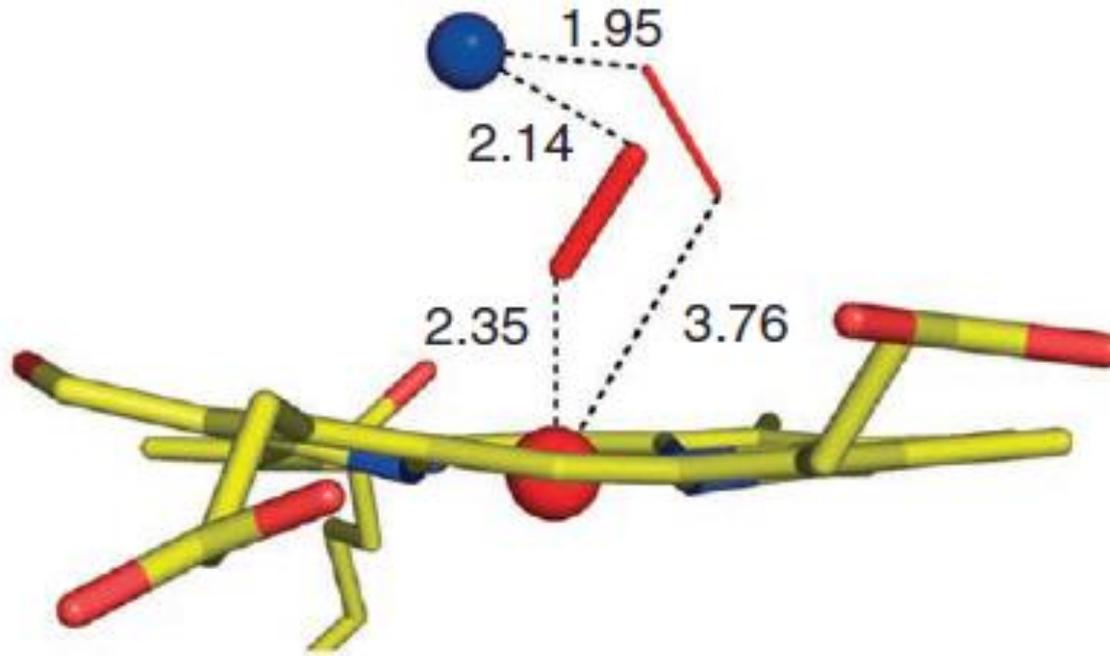
Cu<sub>B</sub> (rame tipo 2)



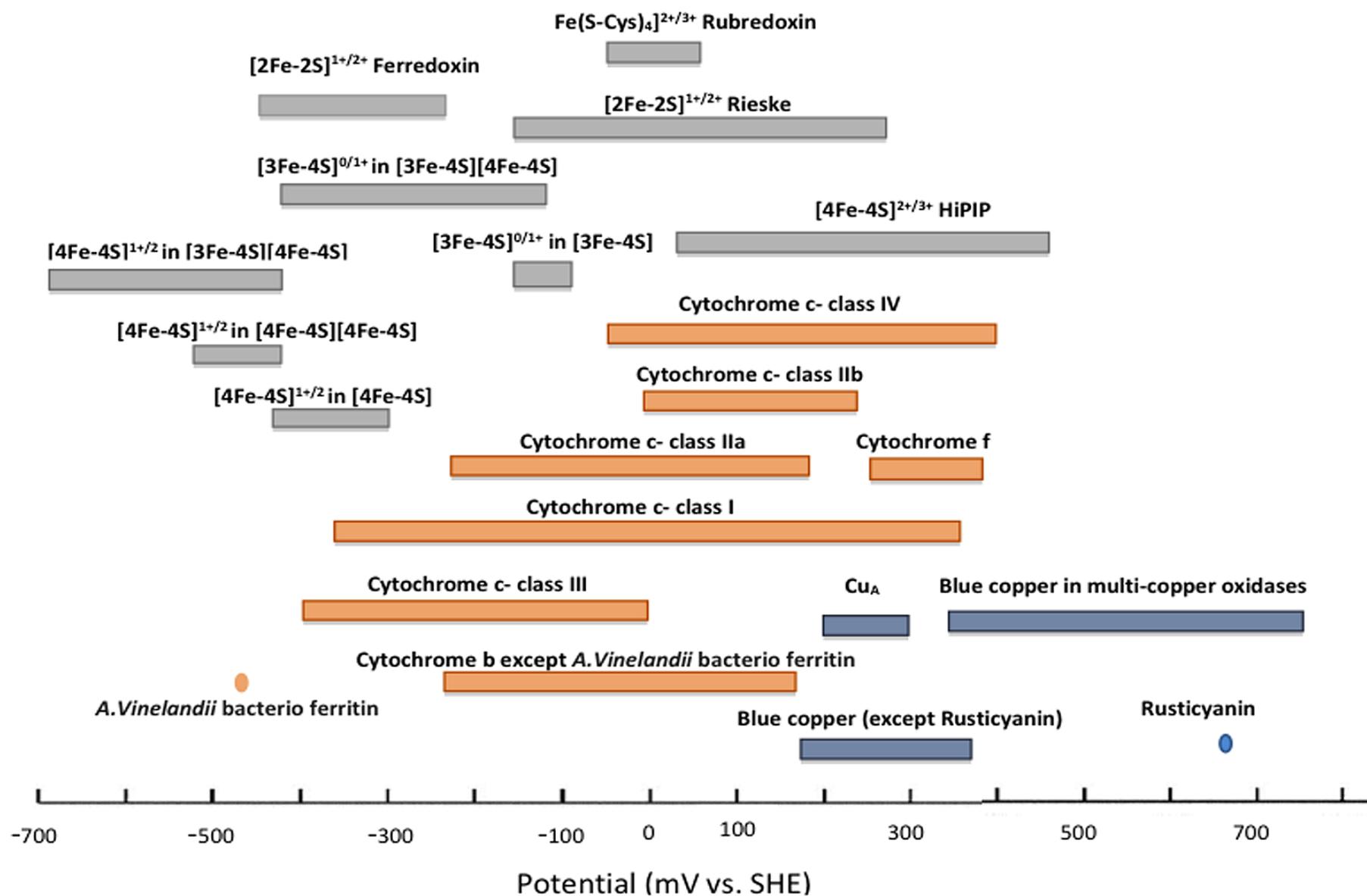


# Intermedio perossidico

*X-ray free-electron laser (XFEL)*

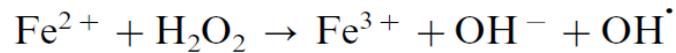
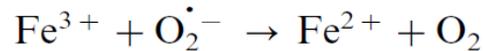
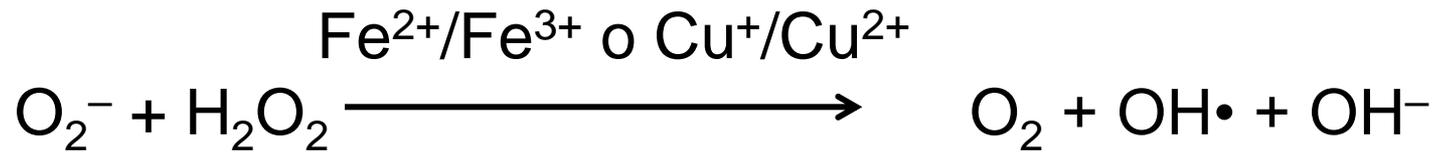


*Nature*, 2014



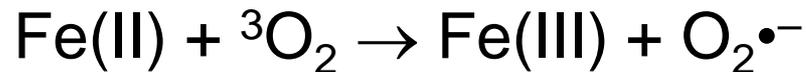
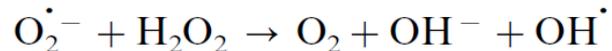
# Reactive Oxygen Species (ROS)

## Reazione di Haber-Weiss

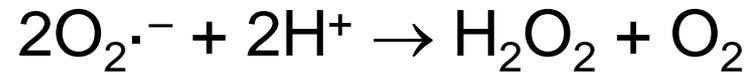


## Reazione di Fenton

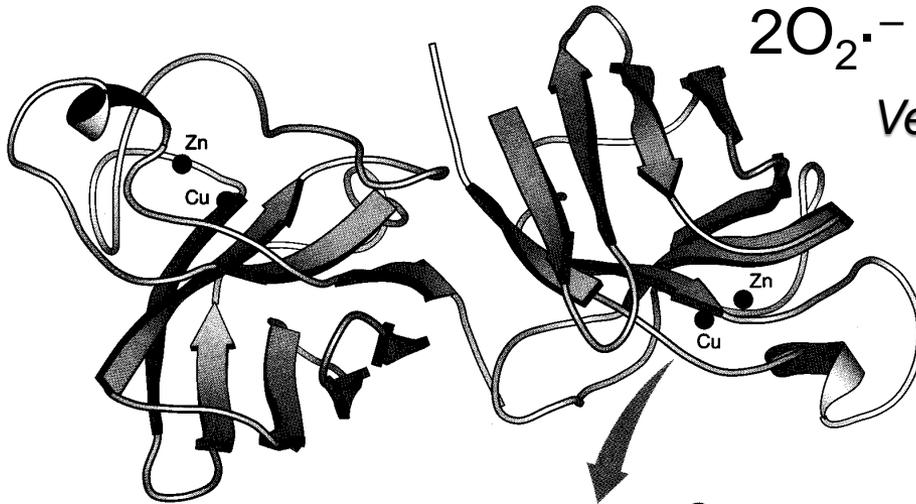
The net reaction:



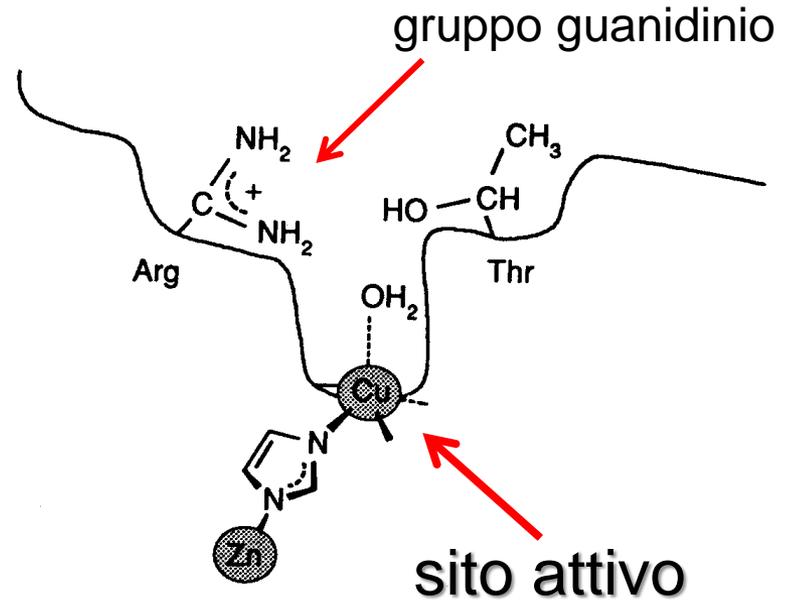
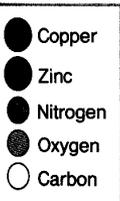
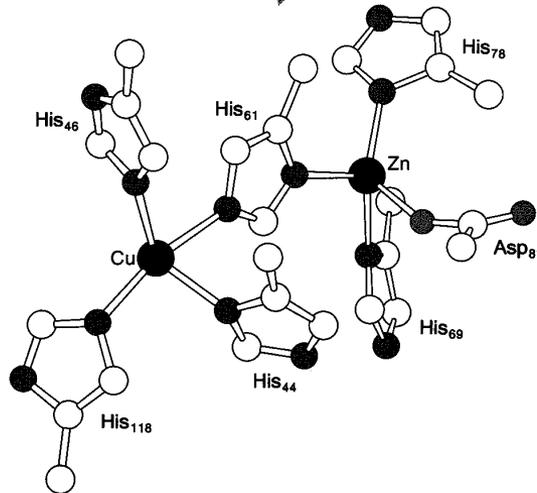
# Cu-Zn superossido dismutasi



*Velocità quasi-diffusiva*



16 kDa



# Ciclo catalitico della superossido dismutasi

