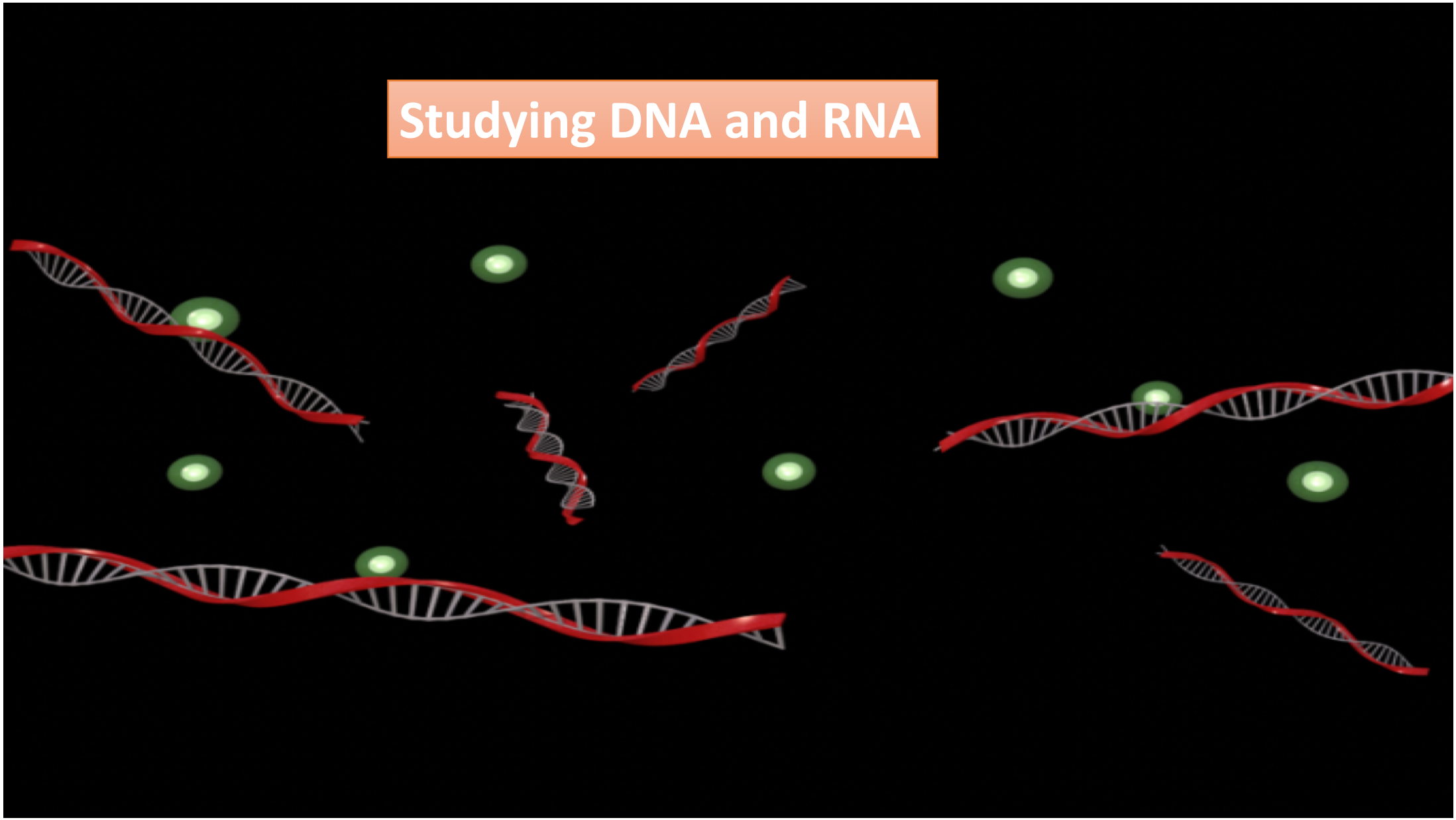
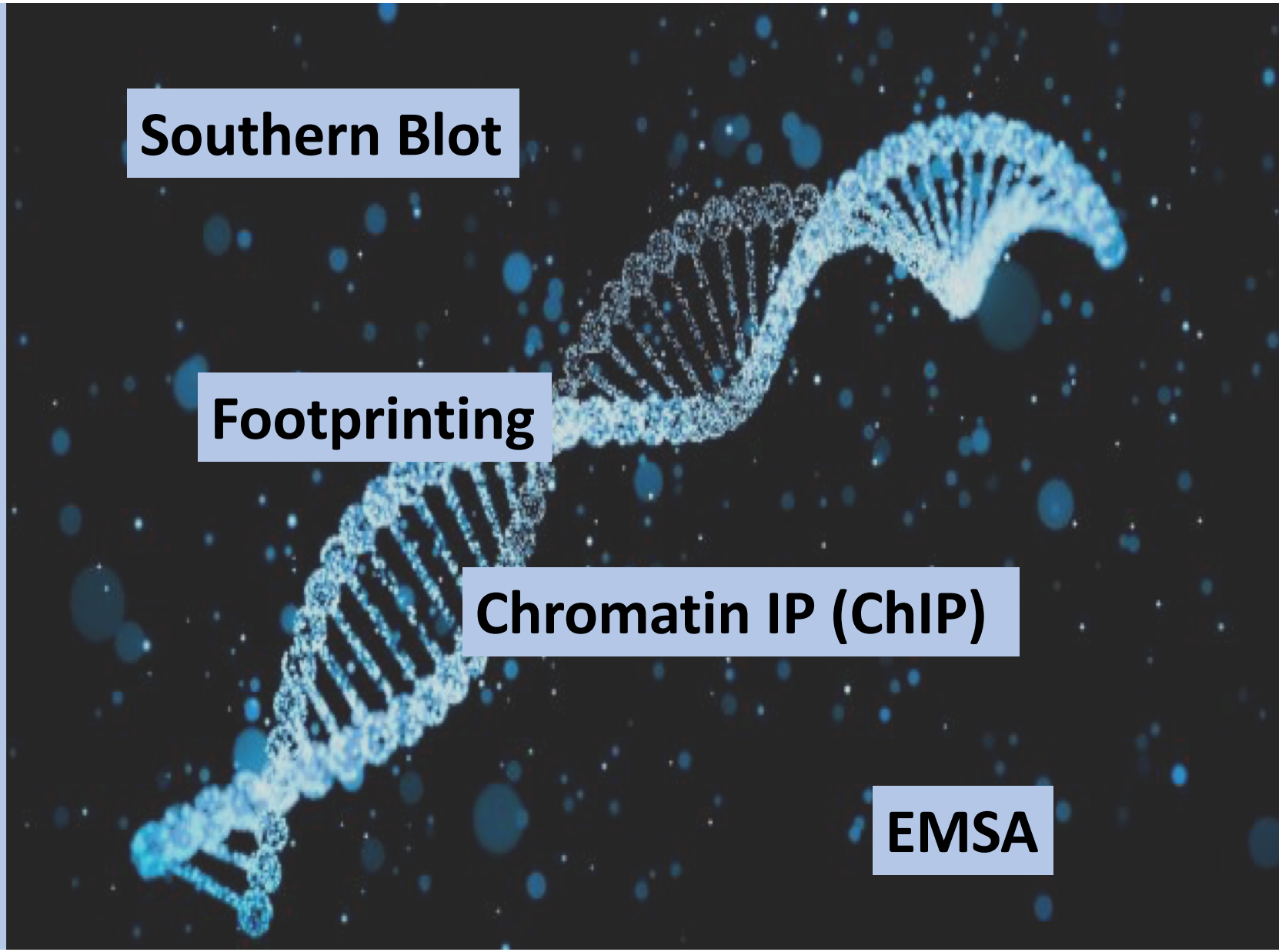
Studying DNA and RNA



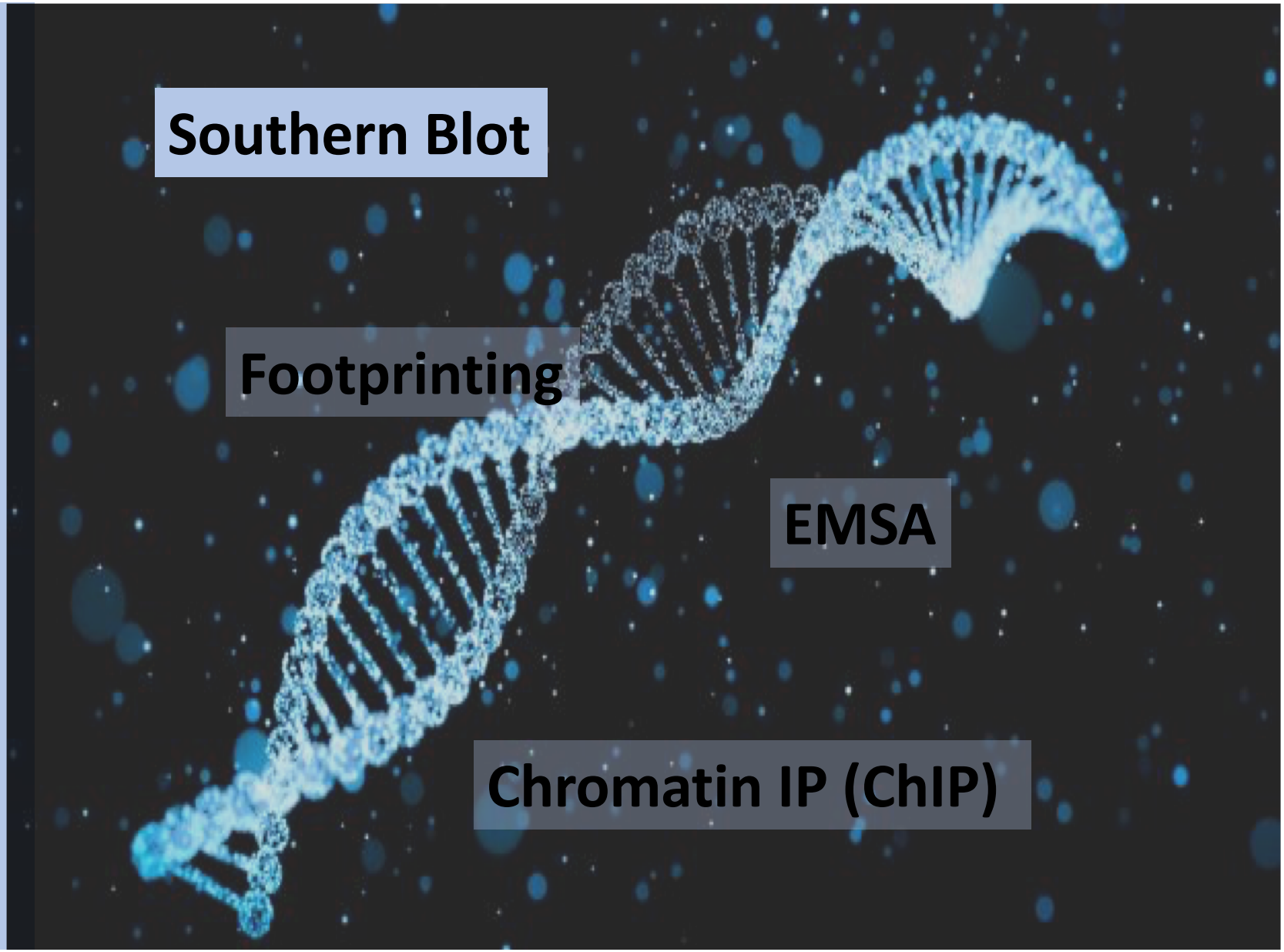
Focus on

DNA



Focus on

DNA



Southern blot - Introduction

Blots are techniques for transferring **DNA** ,**RNA** and **proteins** onto a carrier so they can be separated, and often follows the use of a gel electrophoresis.

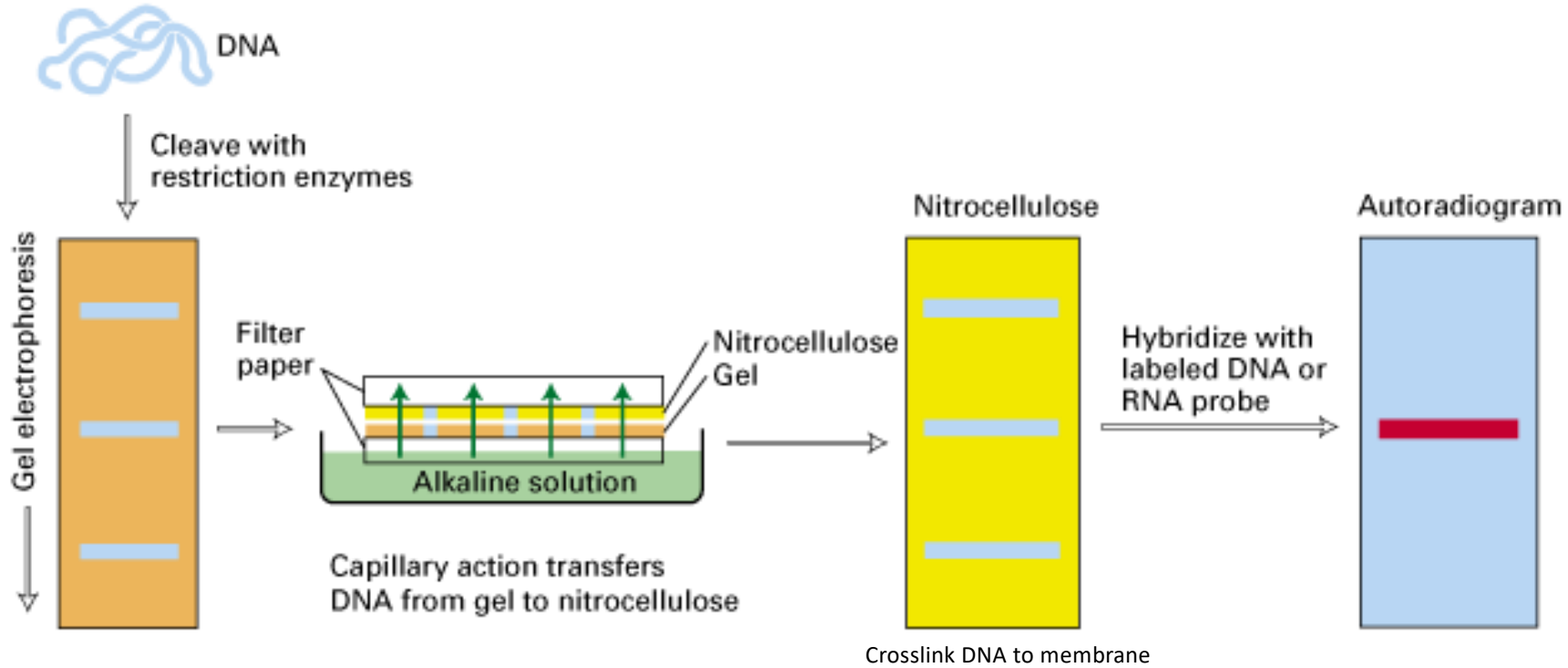
The **Southern blot** is used for transferring DNA, the **Northern blot** for RNA and the western blot for PROTEIN.

Southern blotting

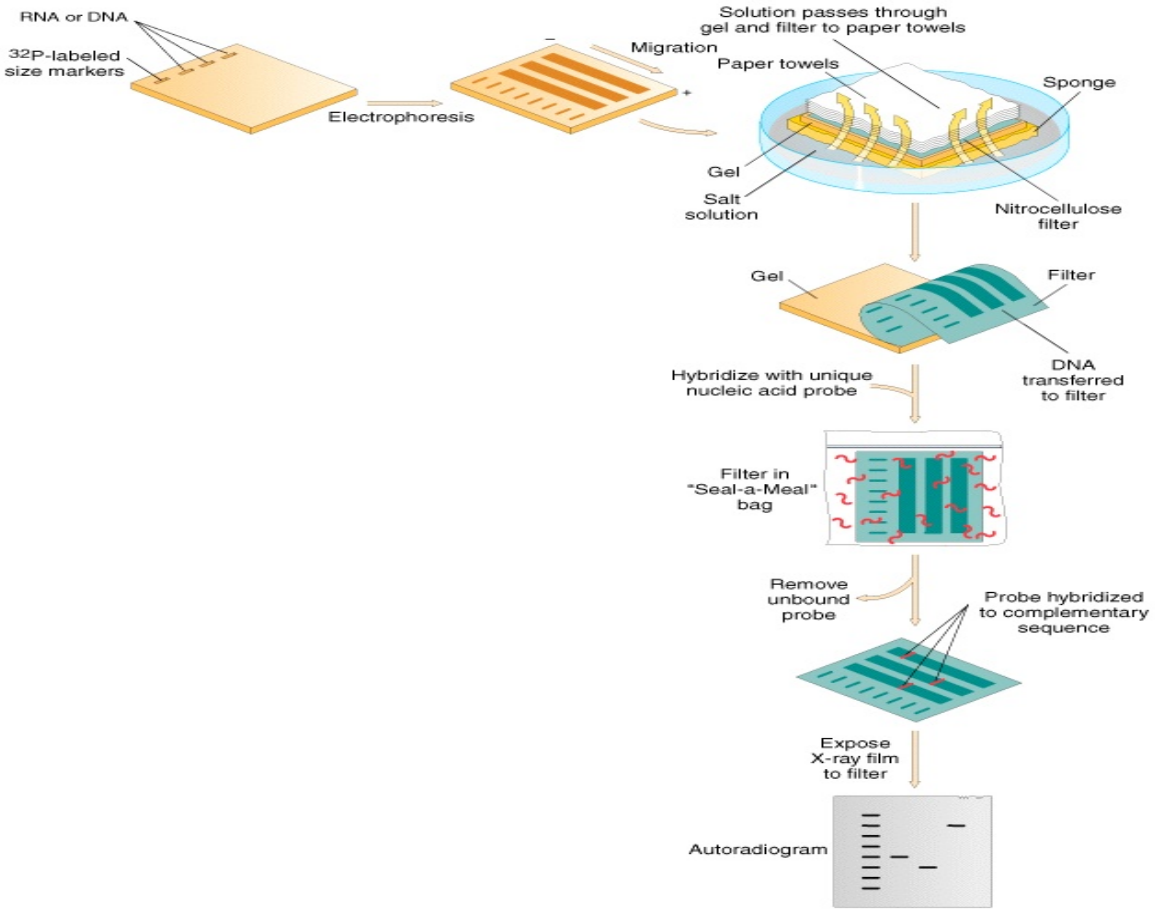
In the 1970s **Edwin Southern** of Oxford University invented a revolutionary DNA blotting technique.

The Southern Blot allows the visualization of one DNA fragment from a whole genome DNA extract.

Southern Blotting



Southern Blotting – general scheme



Southern Blotting – Steps

Step 1: DNA purification

Isolate the DNA in question from the rest of the cellular material in the nucleus.

Incubate specimen with detergent to promote cell lysis.

Cell Lysis frees cellular proteins and DNA.

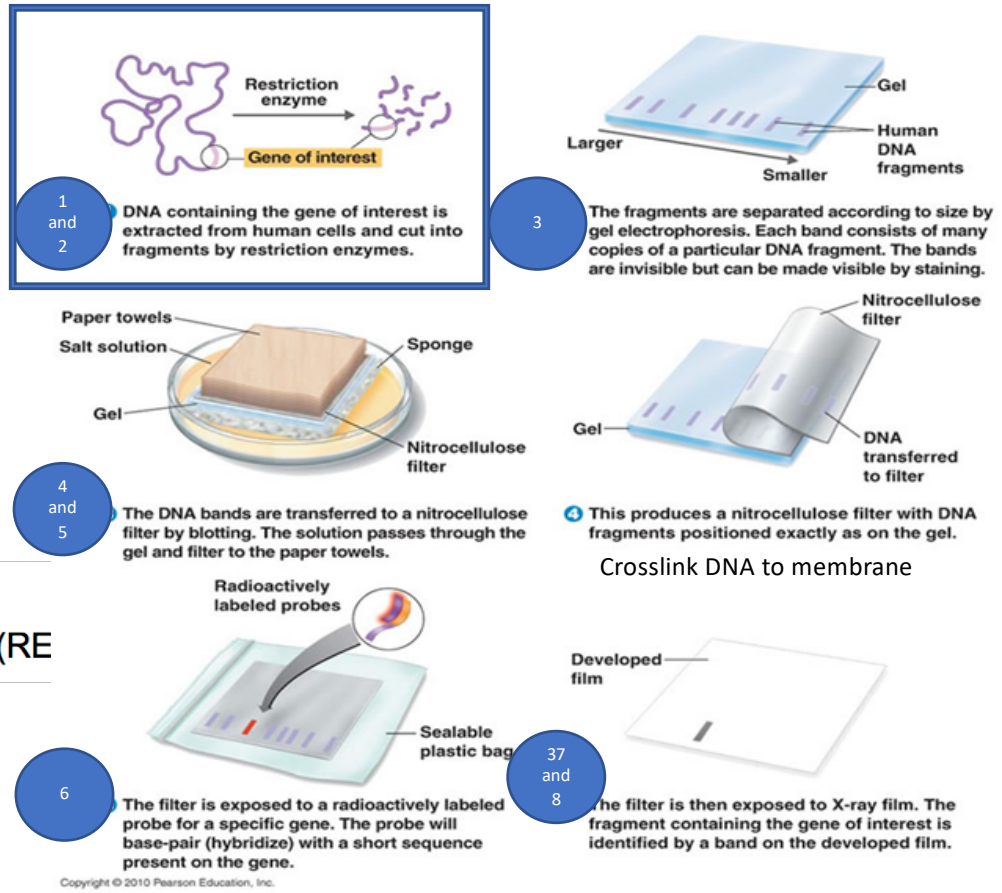
- Proteins are enzymatically degraded by incubation with proteinase.
- DNA is purified from solution by alcohol precipitation.
- Visible DNA fibers are removed and suspended in buffer.

Step 2 : Restriction digestion

Cut the DNA into different sized fragments using restriction endonucleases (RE

By purifying DNA use

- **DNAse free** solutions and reagents
- **RNAse** to avoid RNA contamination
- **ice**



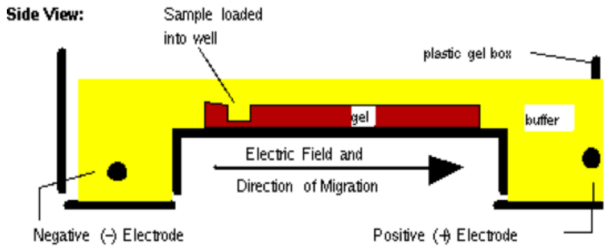
Crosslink DNA to membrane

Southern Blotting – Steps

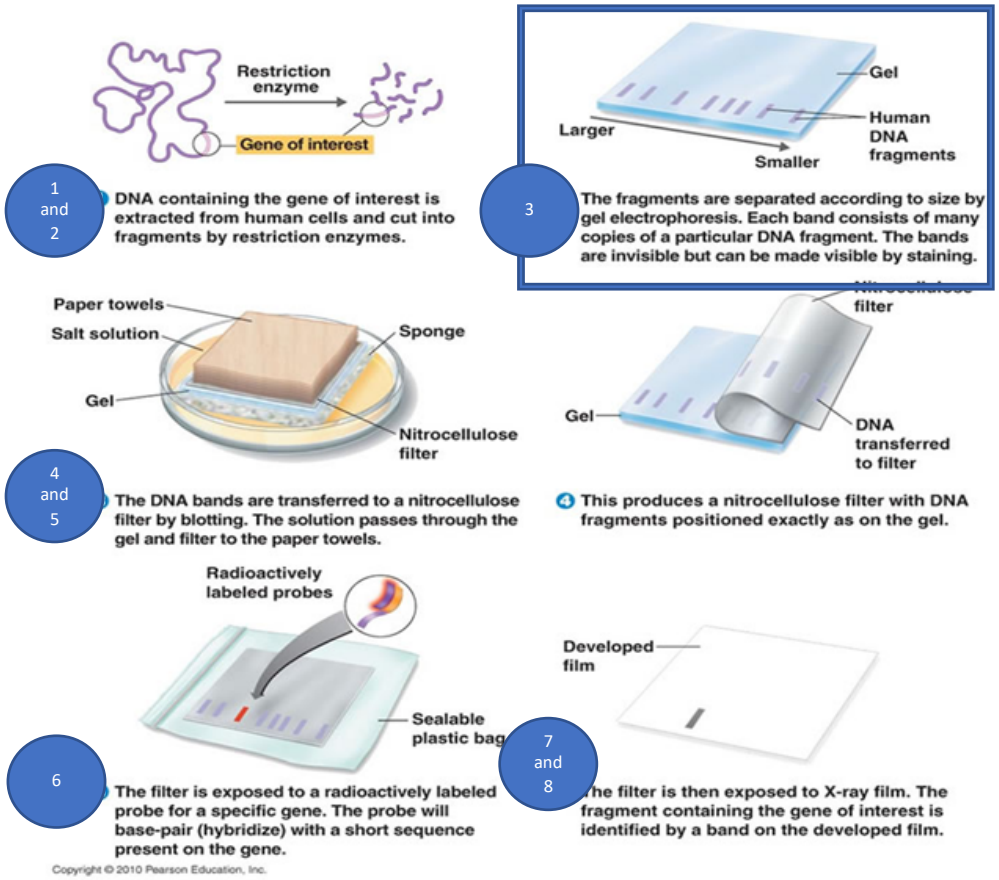
Step 3 : Gel Electrophoresis

Nucleic acids have a net negative charge and will move from the left to the right. The larger molecules are held up while the smaller ones move faster. This results in a separation by size.
Gels are Agarose or polyacrylamide with microscopic pores

Standards should also be run



Gels can be stained with ethidium bromide (EtBr). This causes DNA to fluoresce under UV light which permits photography of the gel.
This will be help us to know the exact migration of DNA standards and the quality of the RE digestion of the test DNA.



Southern Blotting – Steps

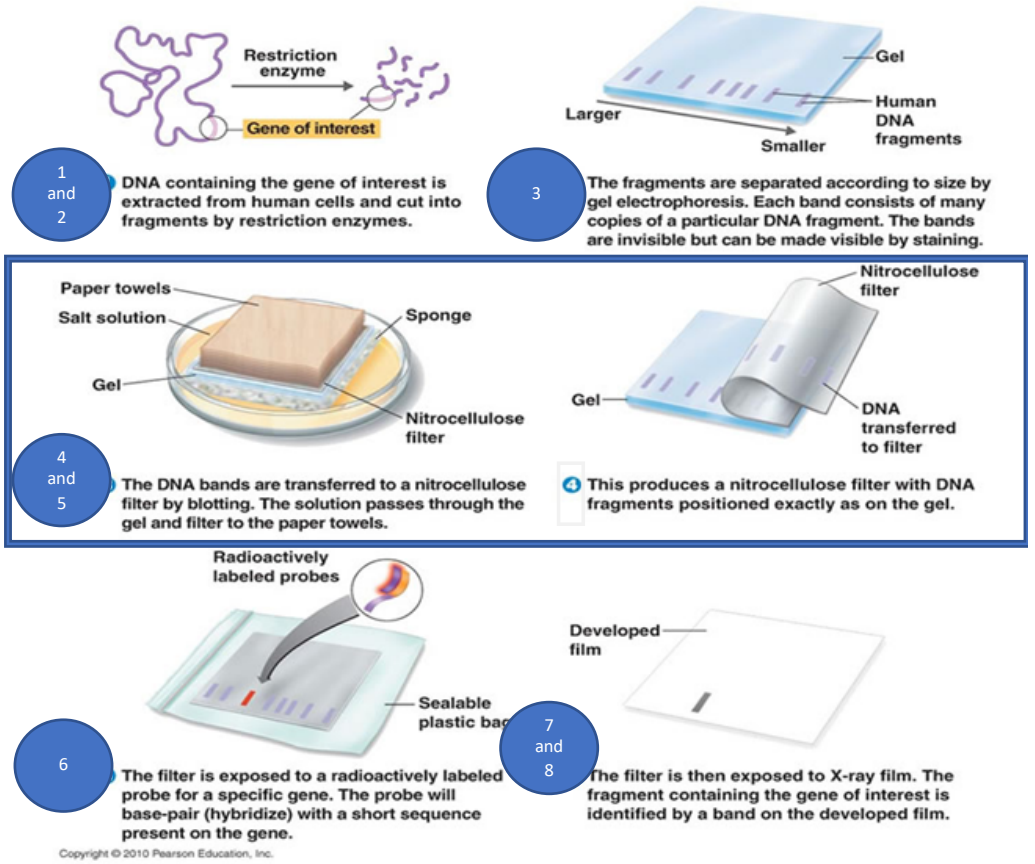
Step 4 & 5 : Denaturation & blotting

DNA is then denatured with an alkaline solution such as NAOH. This causes the double stranded t become single-stranded.

The process of transferring the DNA from the gel to a membrane is called as blotting. The blot is usually done on a sheet of nitrocellulose paper or nylon. DNA is then neutralized with NaCl to prevent re-hybridization before adding the probe. Transferred by either electroblotting or capillary blotting.

The blot is made permanent either by:

- Drying at ~80°C
- Exposing to UV irradiation



Southern Blotting – Steps

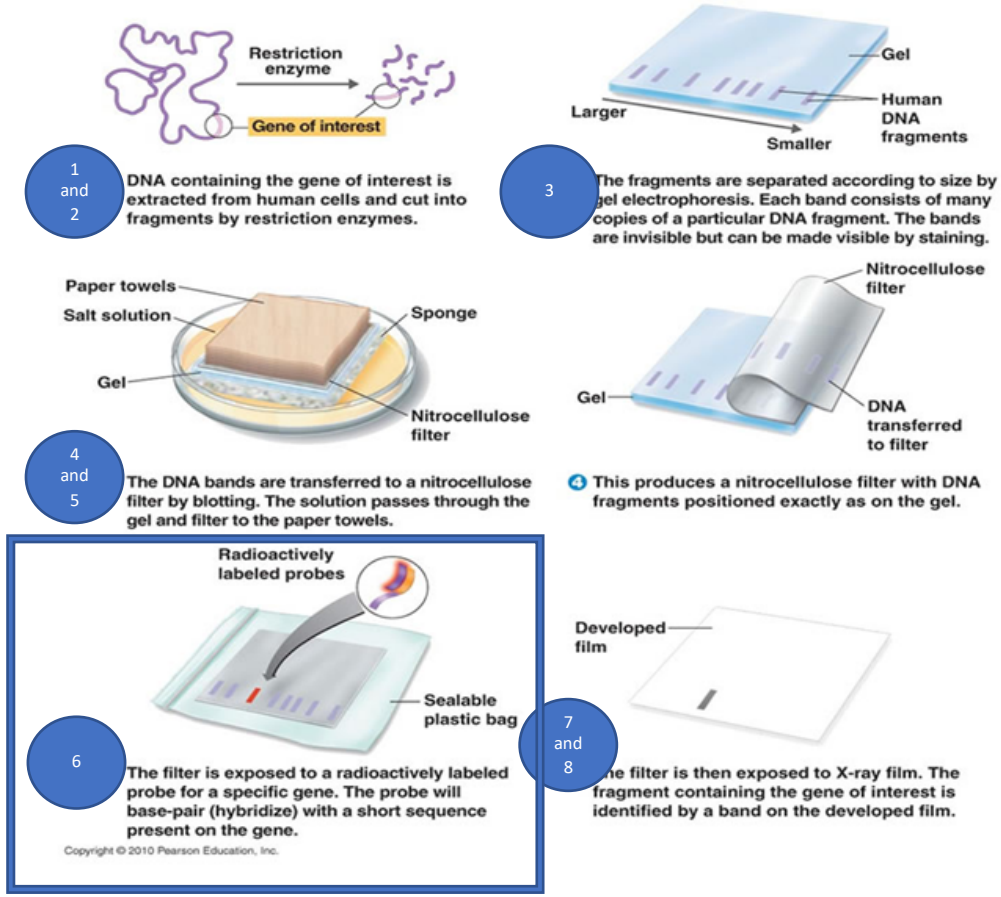
Step 6 : Hybridization

The labelled probe is added to the membrane in buffer and incubated for several hours to allow th probe molecules to find their targets.

Preparing the probe

Small piece of labelled DNA used to find another piece of DNA usually prepared by making a radioactive copy of a DNA fragment.

Probe: fragment of ideally 200 – 1000 bp containing the sequence of interest (present in plasmid; RE; gel-eletrophoresis; purification from gel; alpha-32P-dCTP labelling using Klenow and random oligos)



Southern Blotting – Steps

Step 7 & 8 : Wash and autoradiography

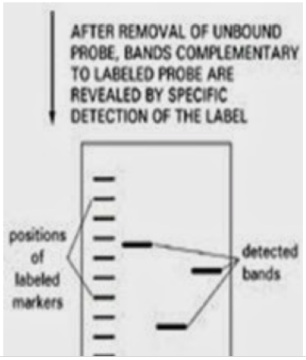
Wash excess probe that are bound non-specifically to the membrane

Blot is incubated with wash buffers containing NaCl and detergent to wash away excess probe ar reduce background.

Detection: Radioactive probes enable autoradiographic detection.

If the probe is radioactive, the particles it emits will expose X-ray film.

By pressing the filter and film, the film will become exposed wherever probe is bound to the filter.



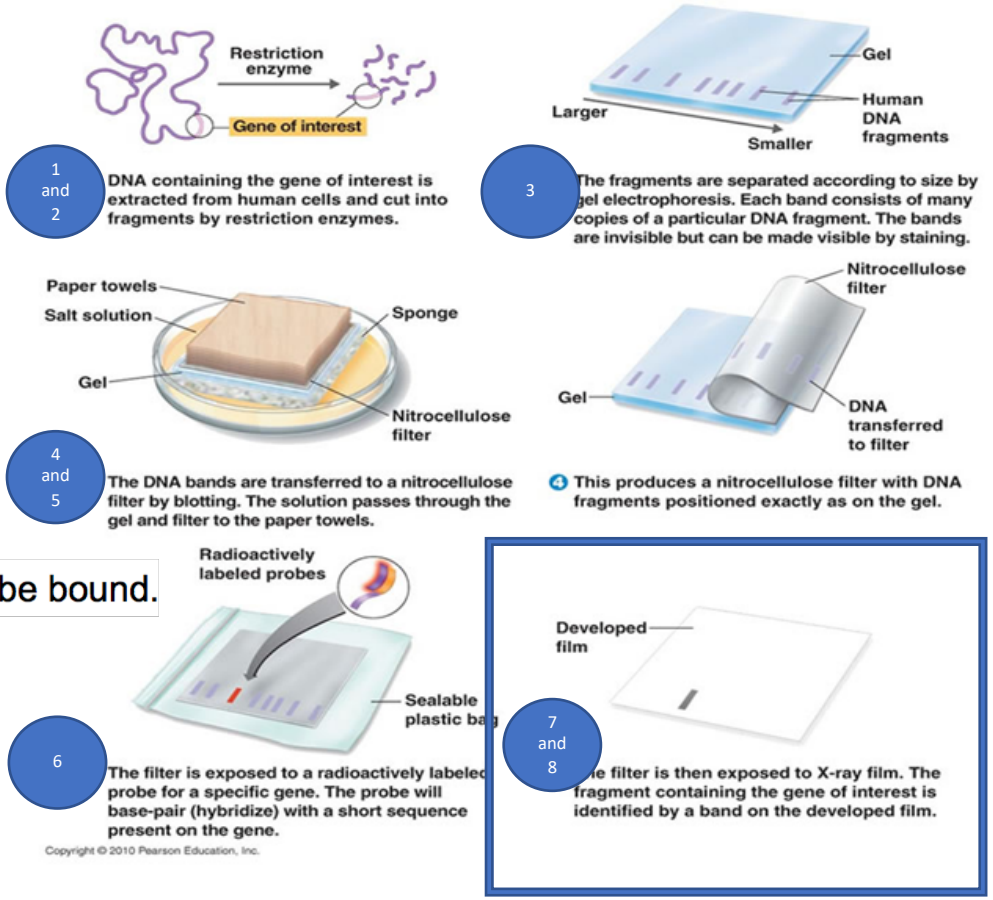
After development, there will be dark spots on the film wherever the probe bound.

Making the probe

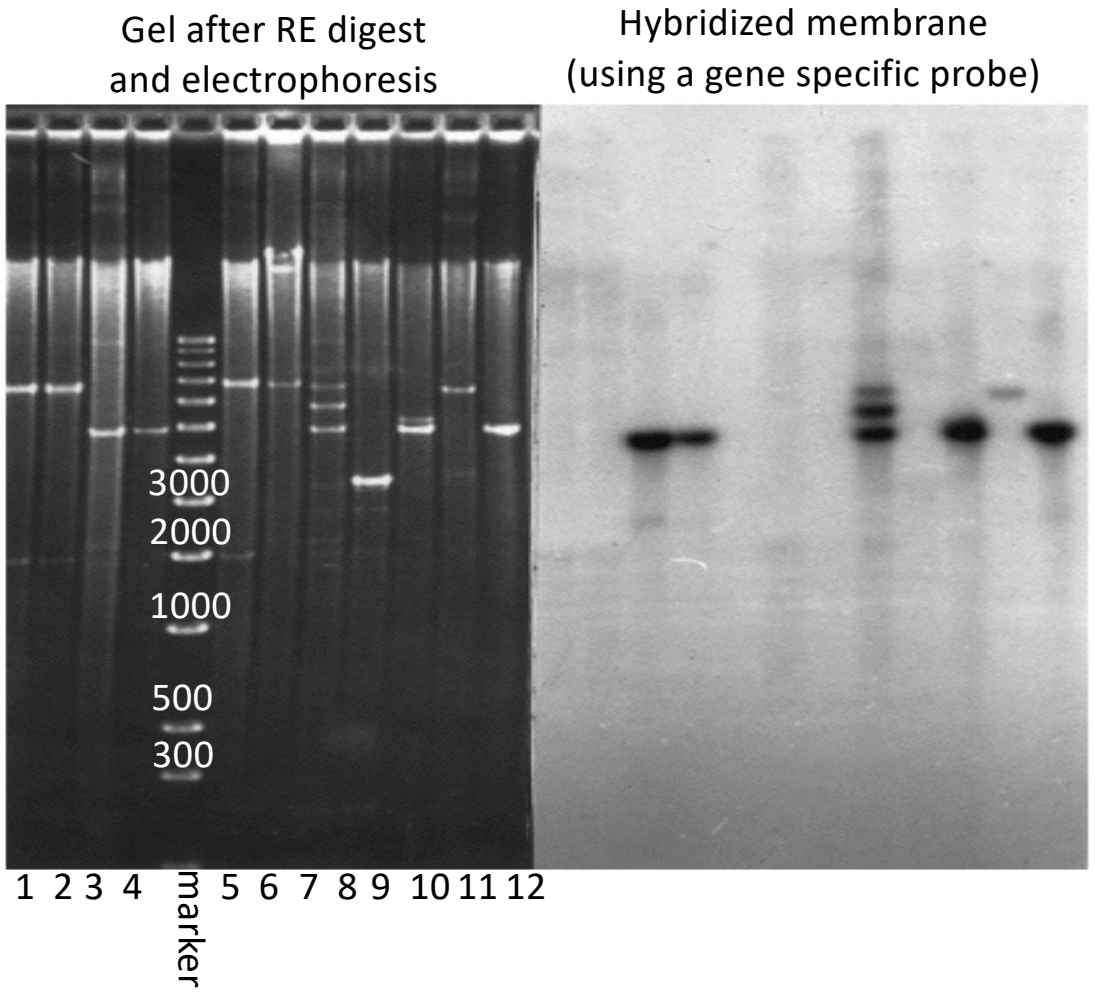
Label the probe to be hybridized using radioactive or non-radioactive methods

Non-radioactive methods A) Colorimetric

B) Chemiluminescent

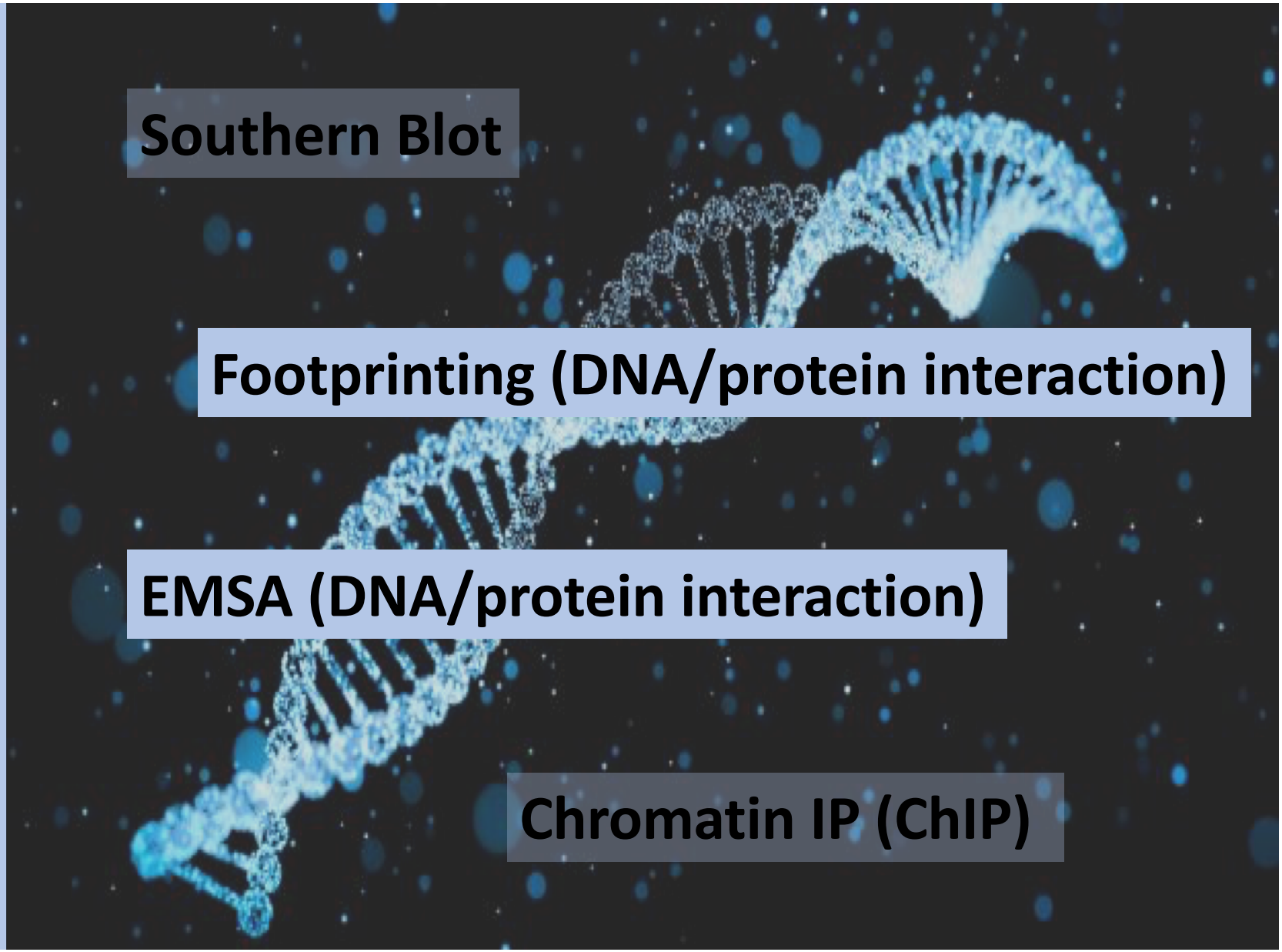


Southern Blotting – Results



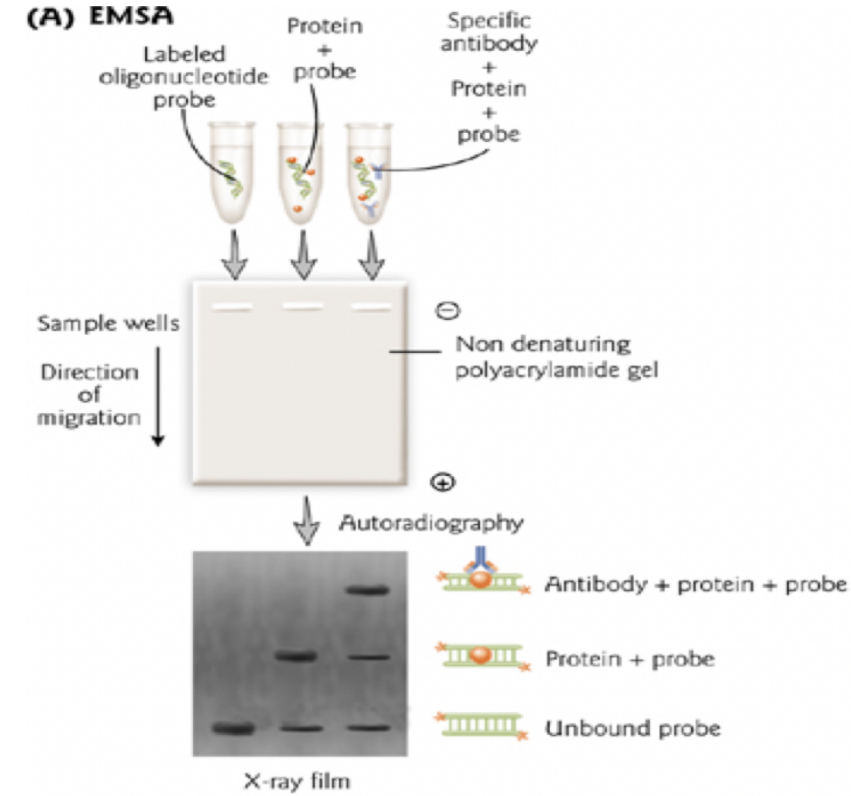
Focus on

DNA

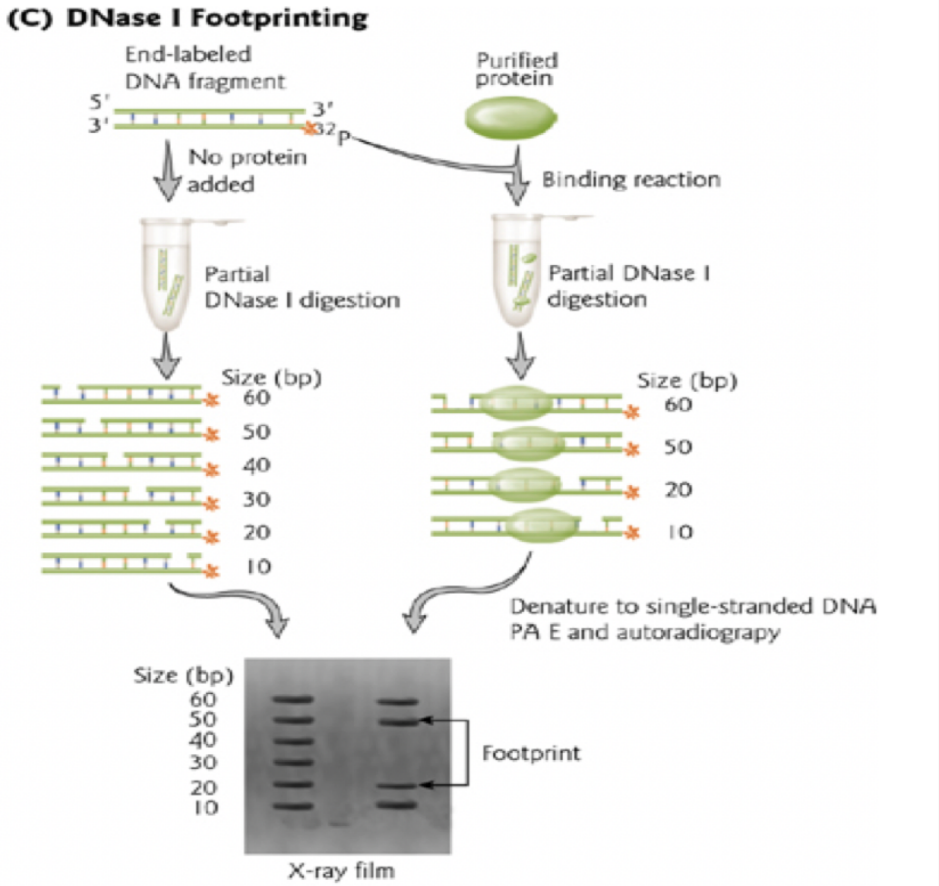

Southern Blot
Footprinting (DNA/protein interaction)
EMSA (DNA/protein interaction)
Chromatin IP (ChIP)

DNA-protein interaction: Footprinting and EMSA

Detecting interactions between DNA and Proteins (for example transcription factor (TF) binding sites)



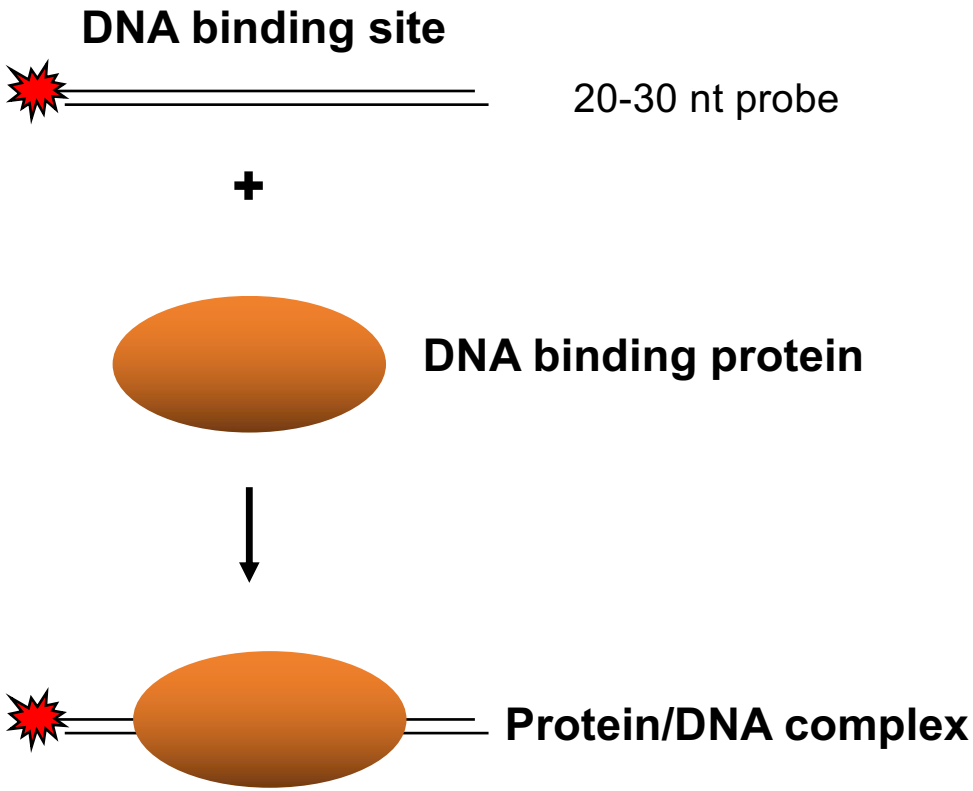
Electophoretic mobility shift assay



DNA: short seqeunce containing putative binding site for TF
Protein: TF, recombinant protein

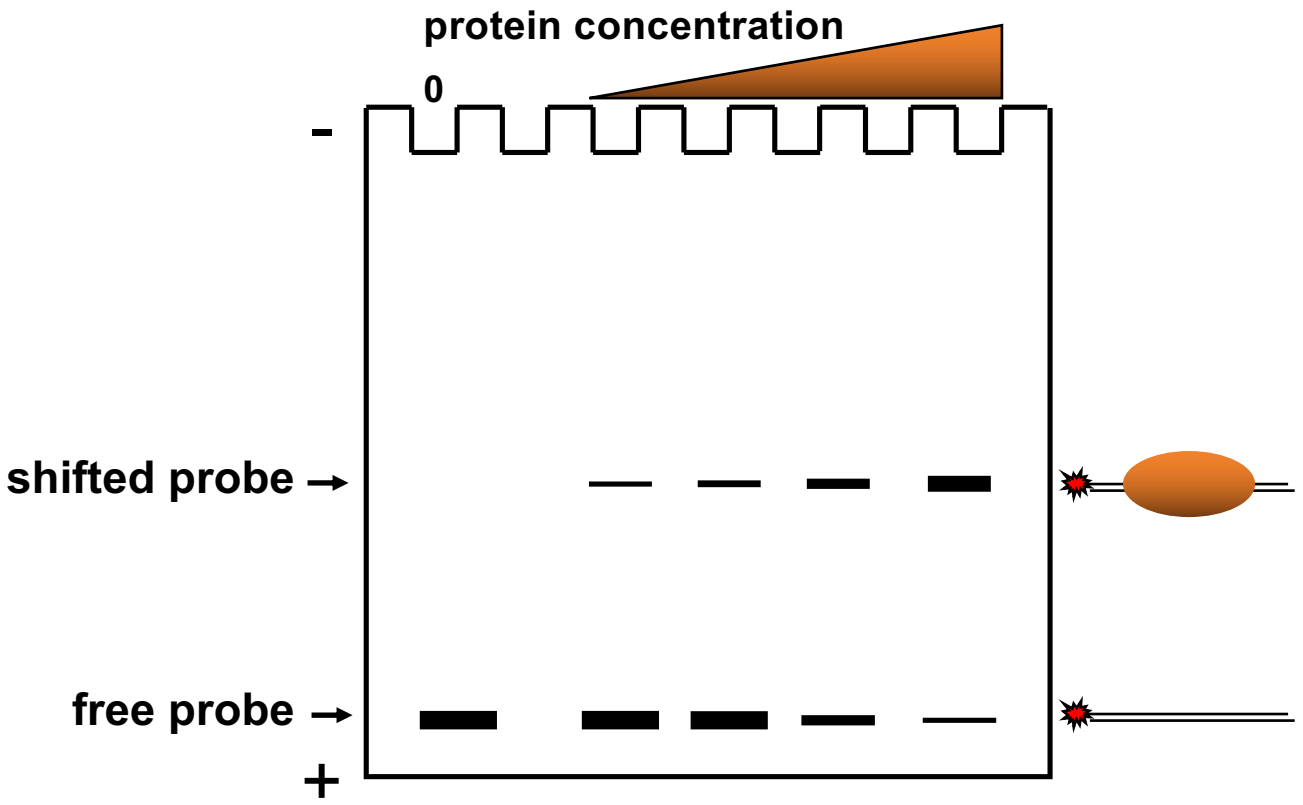
DNA: short seqeunce containing putative binding site for TF
Protein: TF, recombinant protein

Electrophoretic-Mobility Shift Assay (EMSA)



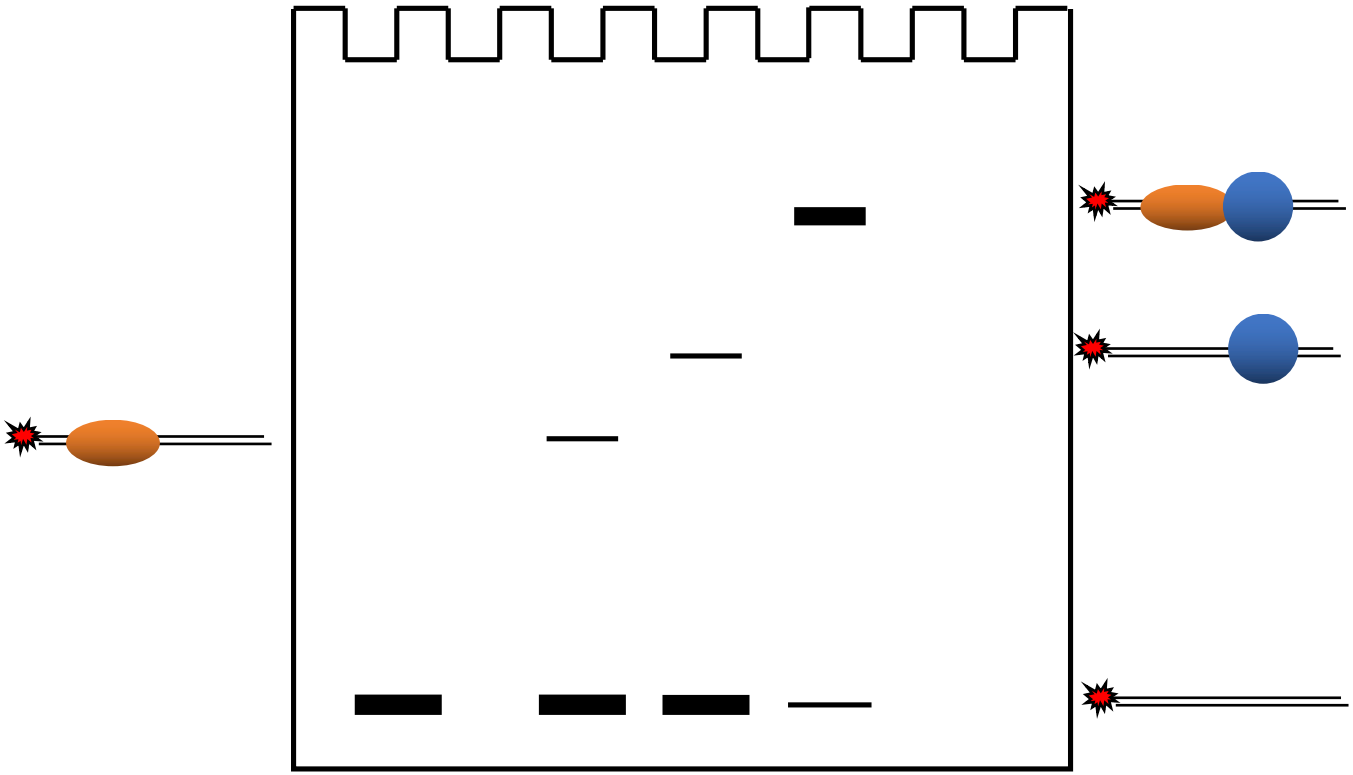
Electrophoretic-Mobility Shift Assay (EMSA)

Incubate protein and DNA probe
Load onto non-denaturing PAGE
Resolve complexes & free probe

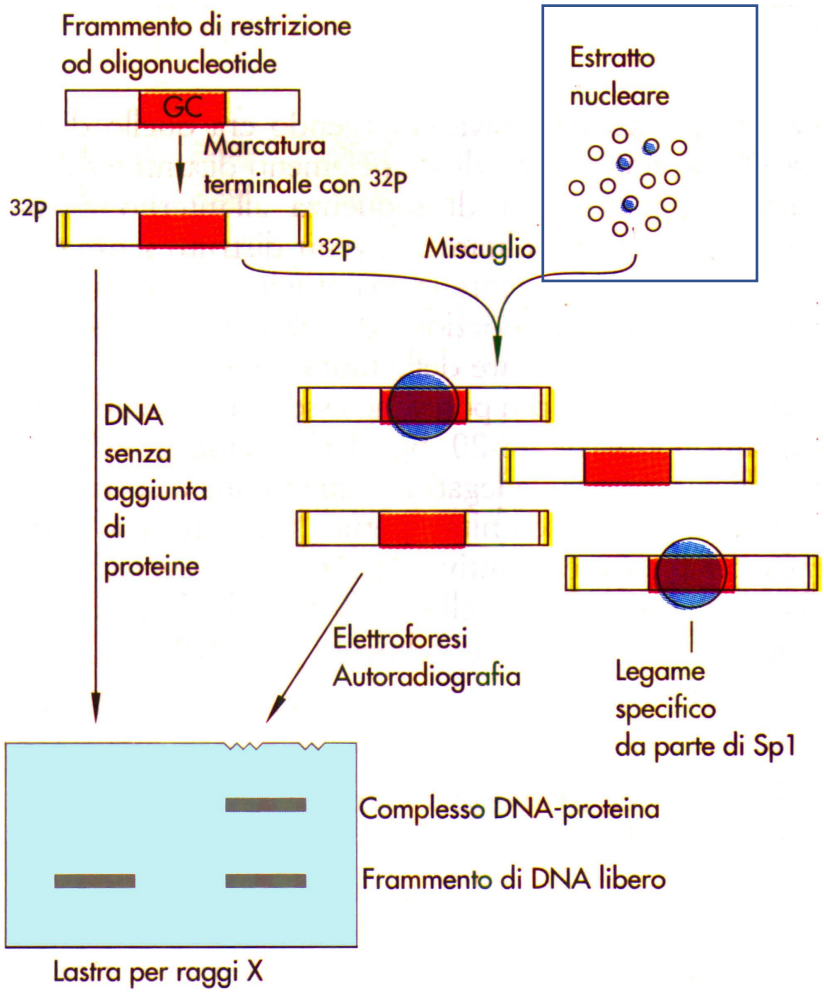


Electrophoretic-Mobility Shift Assay (EMSA)

Resolution of multiple protein/DNA complexes: Proteins sharing a DNA binding element

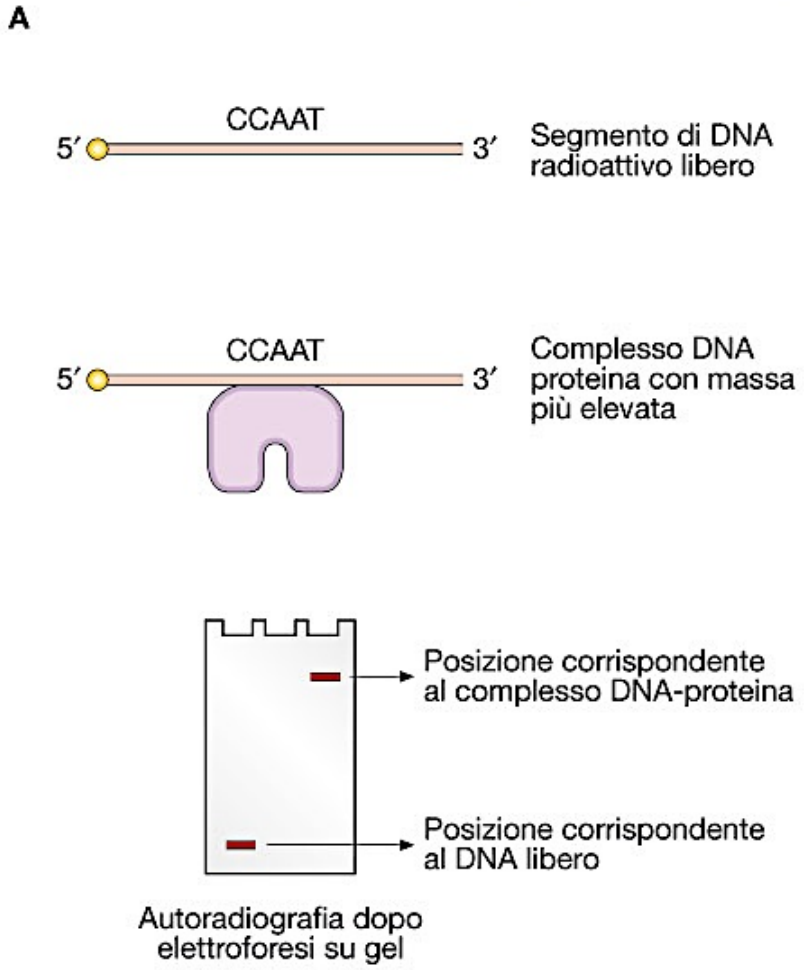


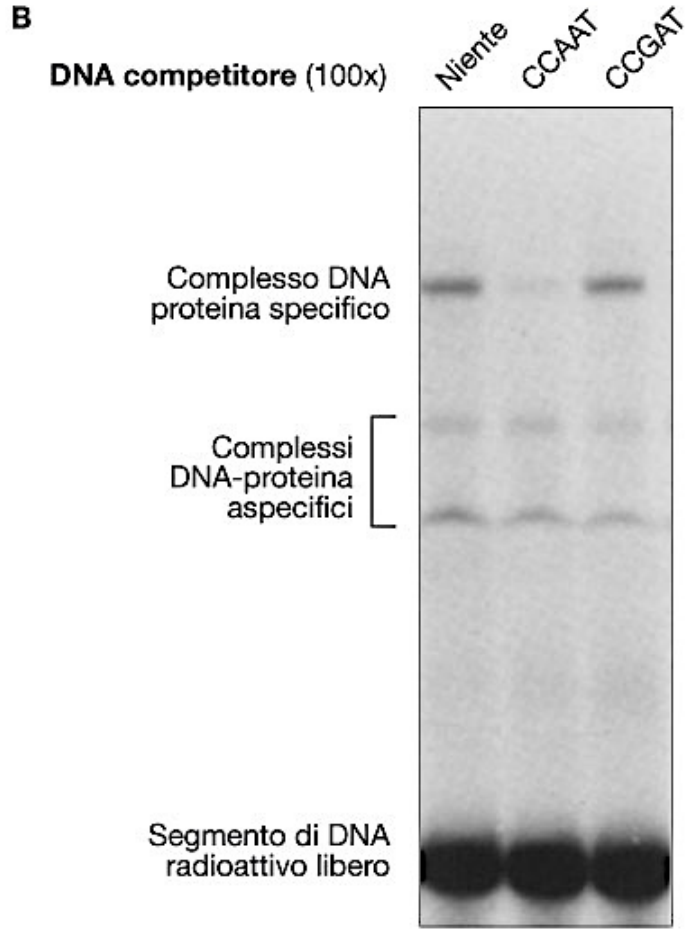
Electrophoretic-Mobility Shift Assay (EMSA)



Electrophoretic-Mobility Shift Assay (EMSA)

Verifying the specificty of DNA-protein interaction





DNAseI- Footprinting

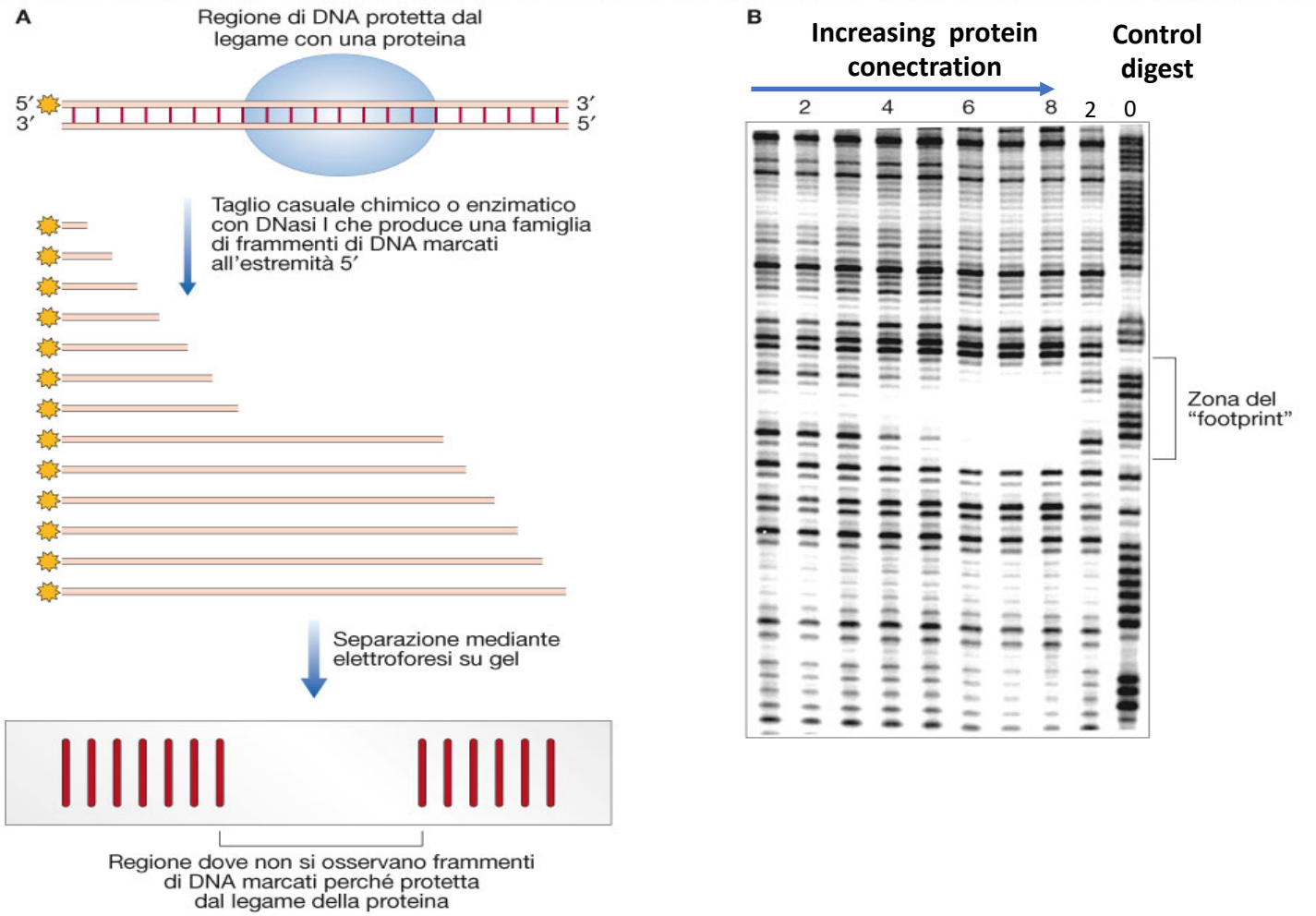
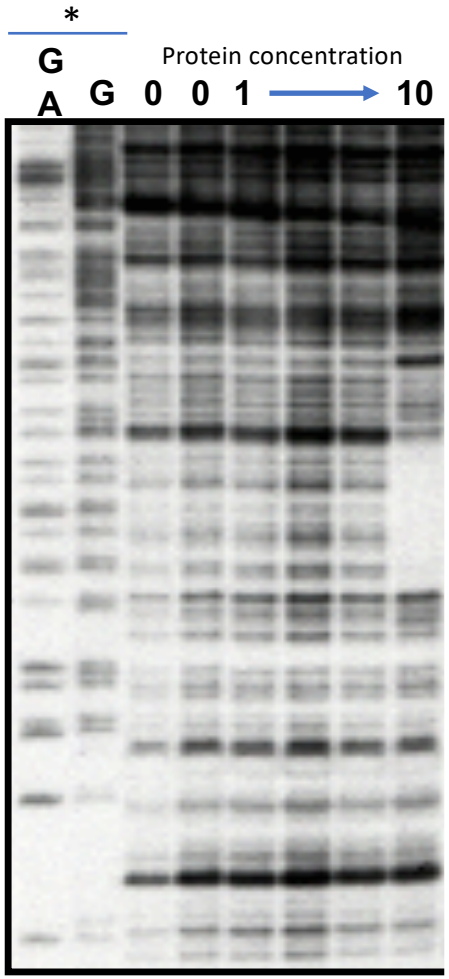
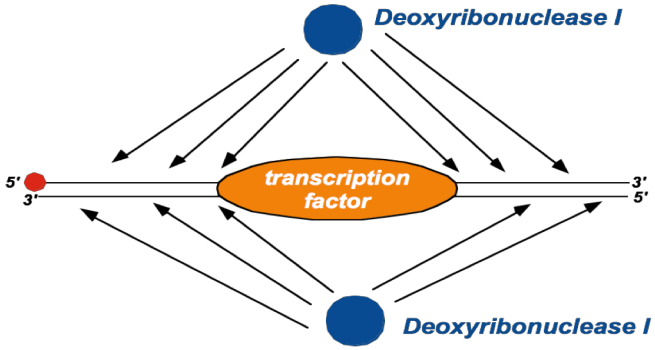


Figura 21.51 Saggio di DNA "footprinting".
(**A**) Schema riassuntivo della procedura. L'asterisco all'estremità 5′ di uno dei due filamenti del frammento di DNA indica un atomo di fosforo radioattivo. Le bande radioattive differiscono una dall'altra per un singolo nucleotide.
(**B**) Esempio di un esperimento di DNA footprinting in cui diverse frazioni di una colonna cromatografica per la purificazione della proteina che si lega al DNA sono state analizzate nelle diverse corsie. La proteina che lega il DNA si trova nelle frazioni 6-8. La posizione dei nucleotidi corrispondenti ai confini della zona di "footprinting" protetta dal legame con la proteina dal taglio con DNasi I può essere definità con estrema precisione.

DNAseI- Footprinting

* Chemical degradation of DNA Maxam-Gilbert sequencing (no protein)





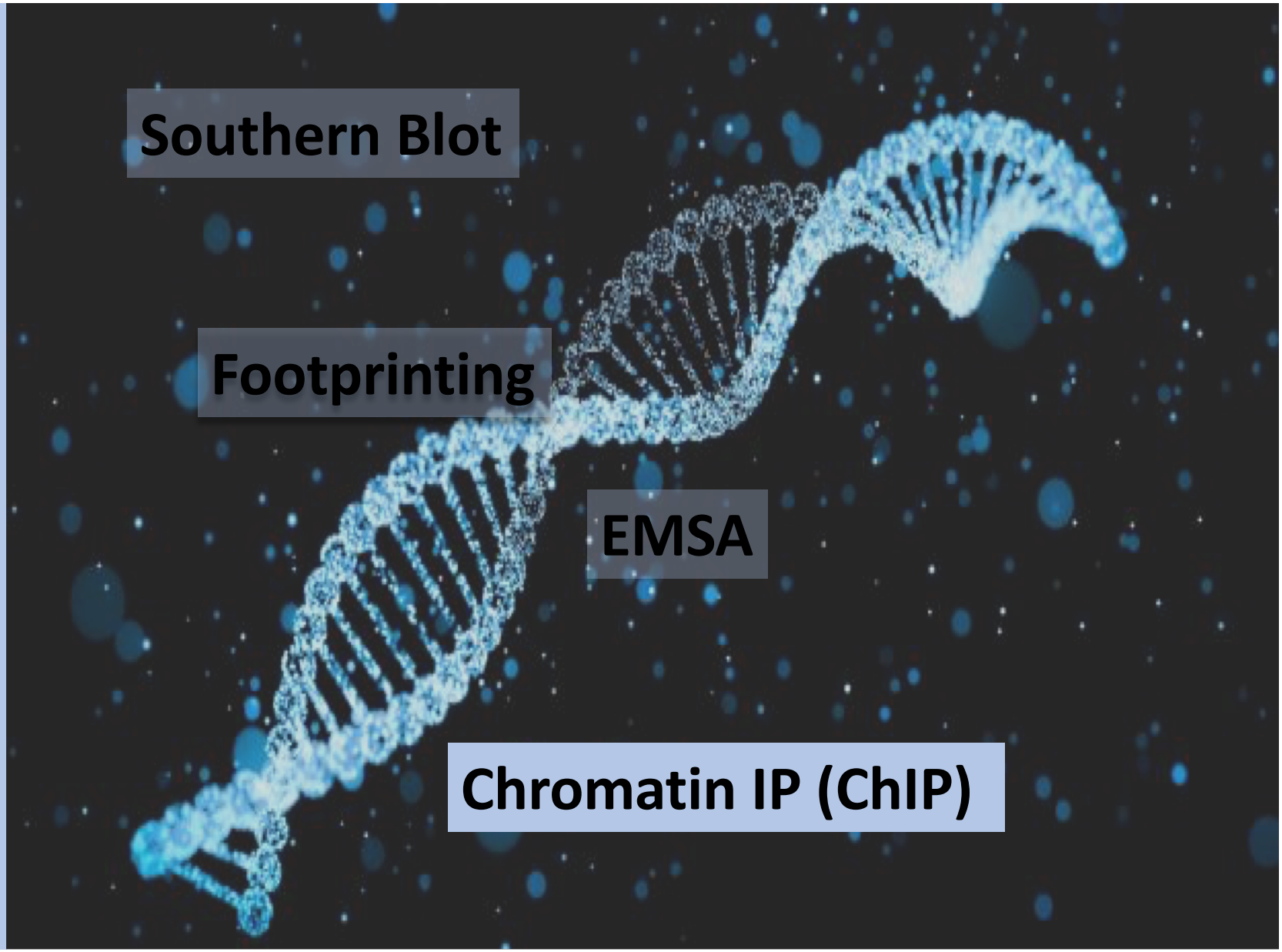
100-300 bp DNA fragment with unique end-label

Incubate with test protein(s)
Digest with DNase I

Run digested DNA on PAGE

Focus on

DNA



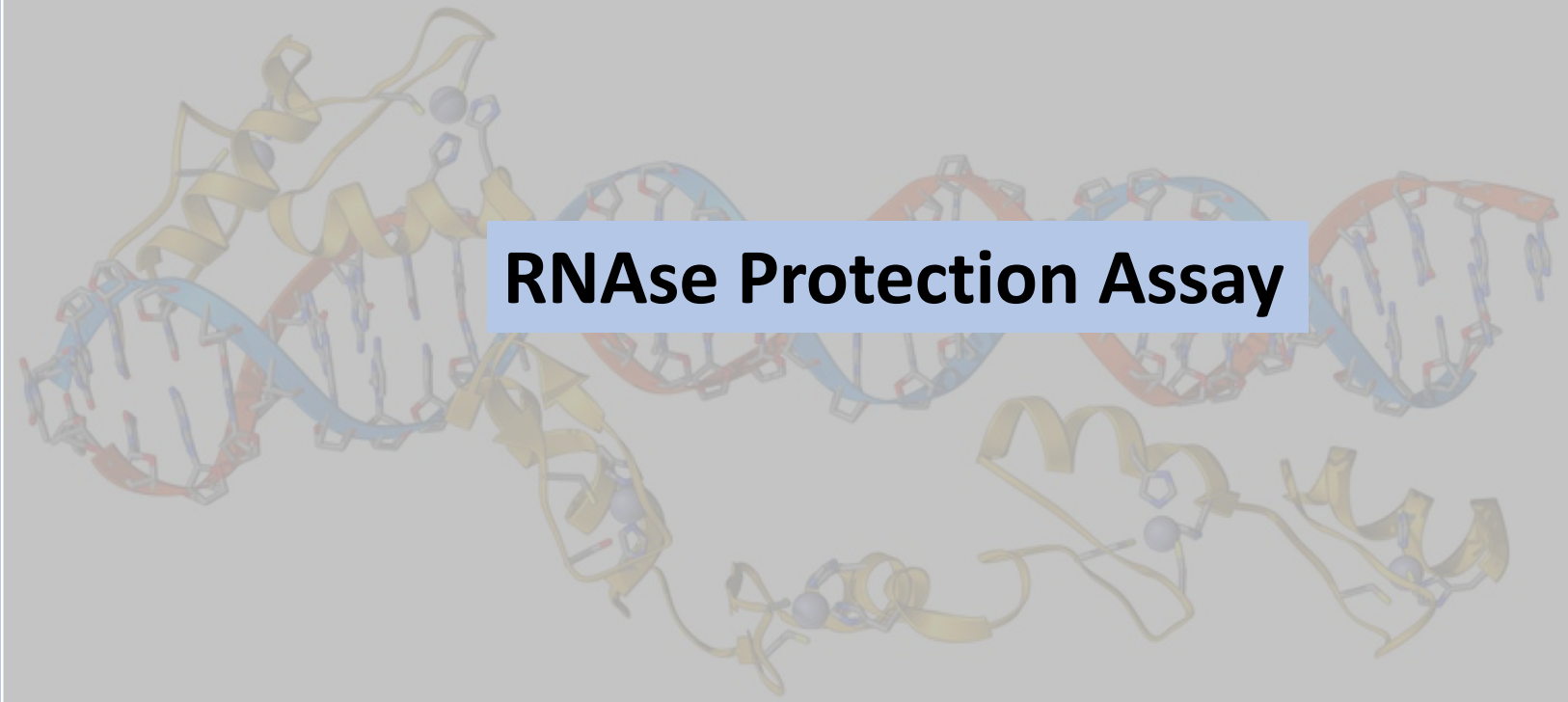
FROM

DNA

TO

RNA

Northern Blotting



RNAse Protection Assay

EMSA for RNA (same as DNA)

FROM

DNA

TO

RNA

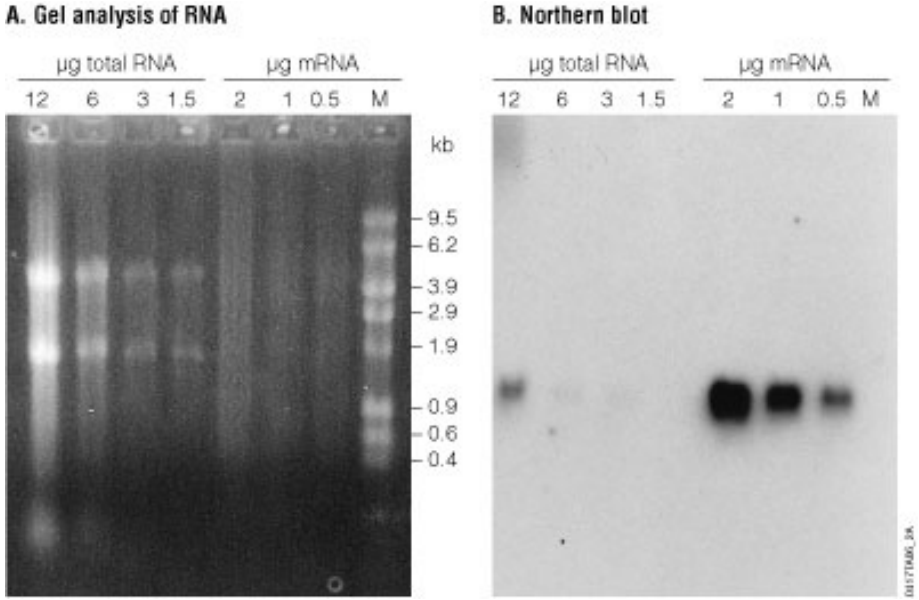
Northern Blotting

RNAse Protection Assay

EMSA for RNA (same as DNA)

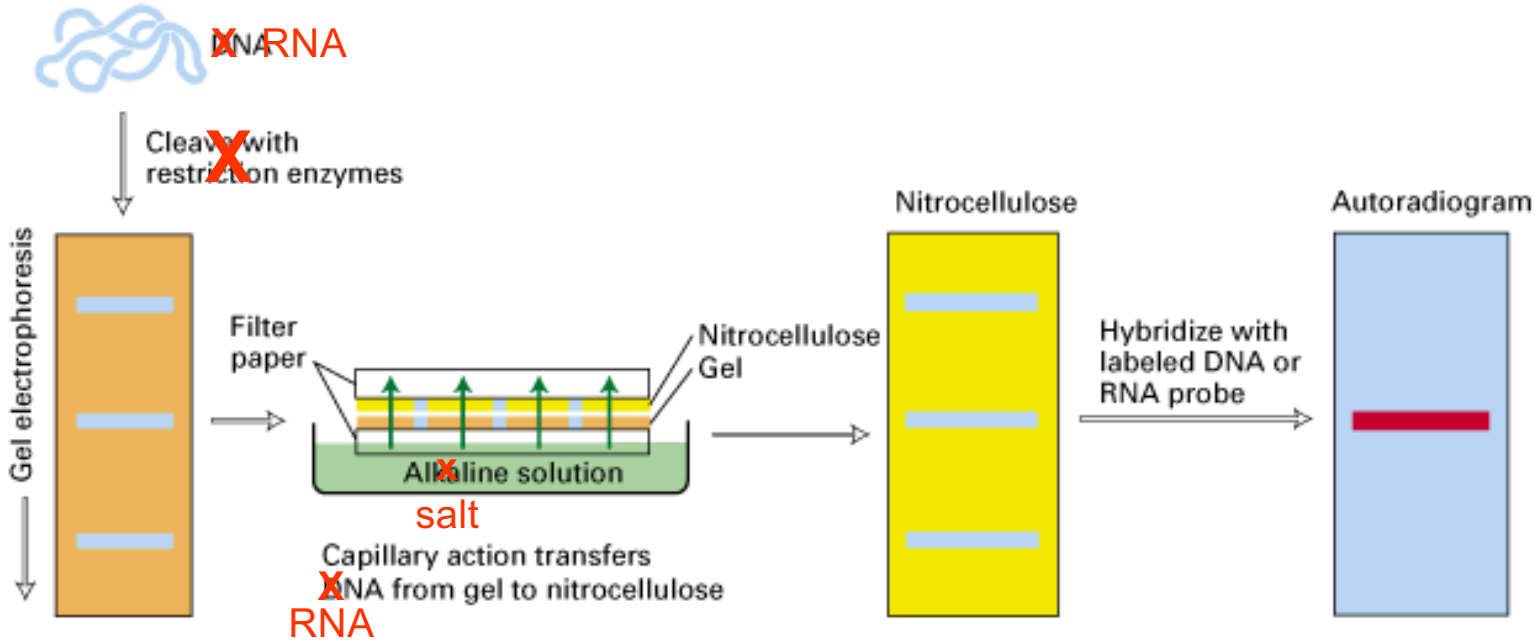
Northern blotting

Northern analysis remains a standard method for **detection and quantitation of mRNA levels** despite the advent of powerful techniques, such as RT-PCR, gene array analysis and nuclease protection assays.
Northern analysis provides a direct relative comparison of message abundance between samples on a single membrane.
It is the preferred method for determining transcript size and for detecting alternatively spliced transcripts

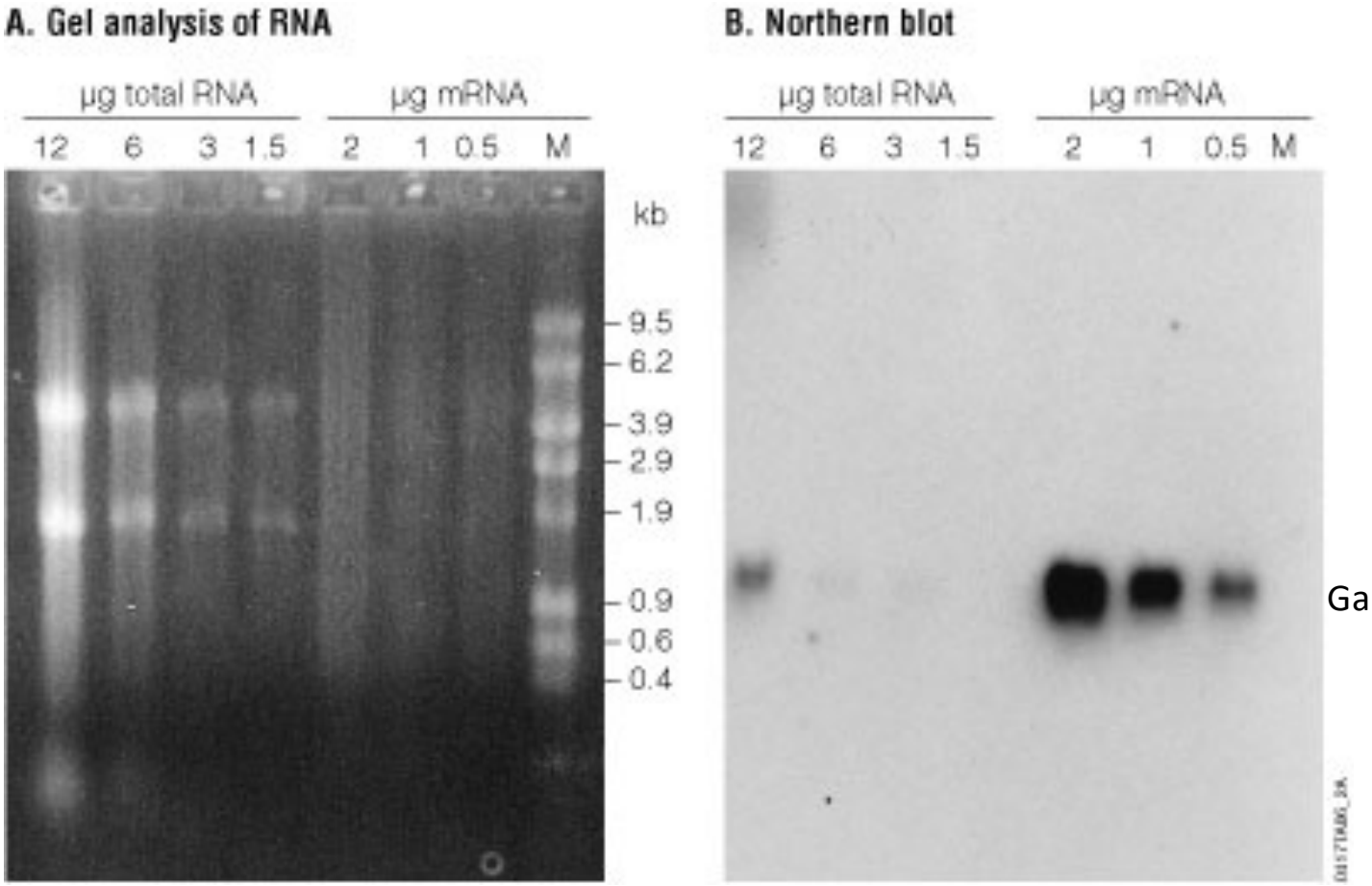


The steps involved in Northern analysis include:
- RNA isolation (total or poly(A) RNA)
- Probe generation
- Denaturing agarose gel electrophoresis
- Transfer to solid support and immobilization
- Prehybridization and hybridization with probe
- Washing
- Detection
- Stripping and reprobing (optional)

Northern blotting vs Southern blotting



-transfer of RNA from a gel to a membrane (e.g., nitrocellulose, nylon)
-reveals mRNA size (and approximate protein size), tissue- and organ-specific expression, and kinetic patterns of expression



Ribosomal subuntis

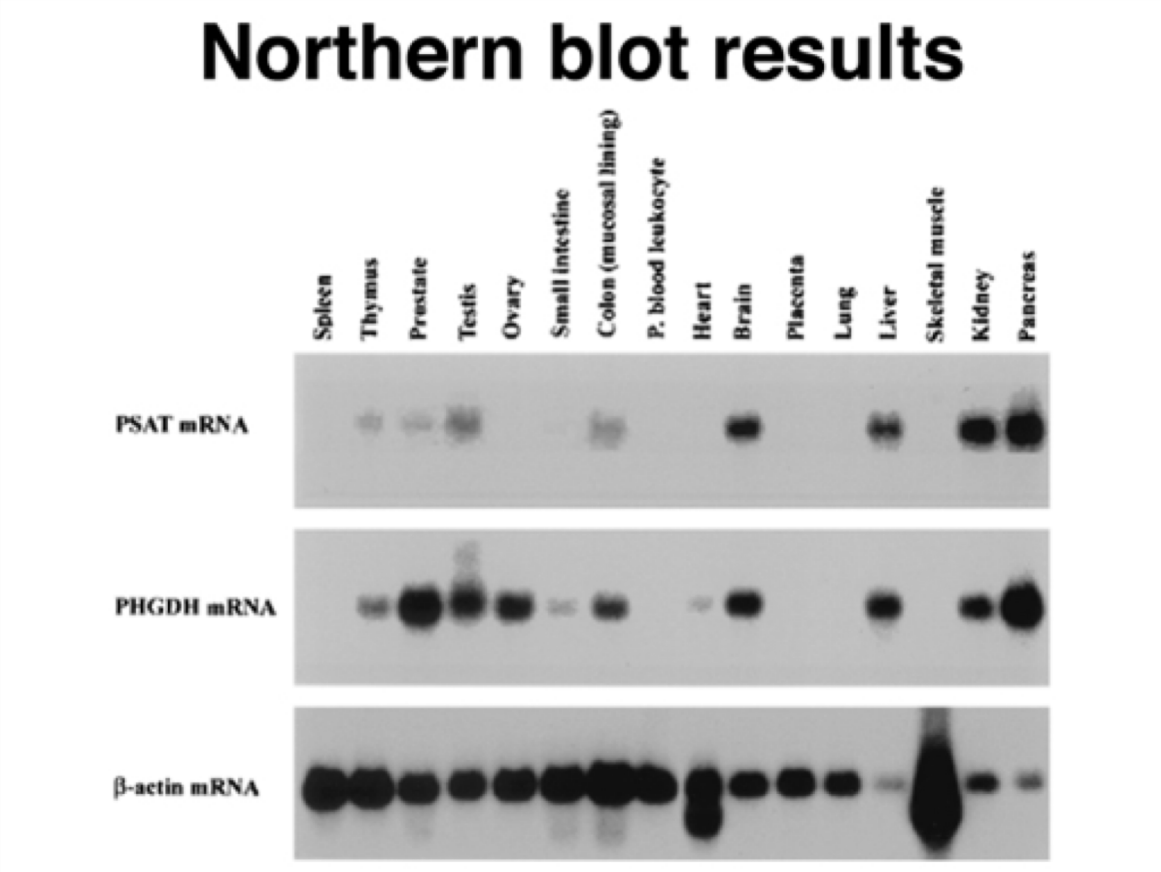
60S

40S

Gapdh mRNA (processed)

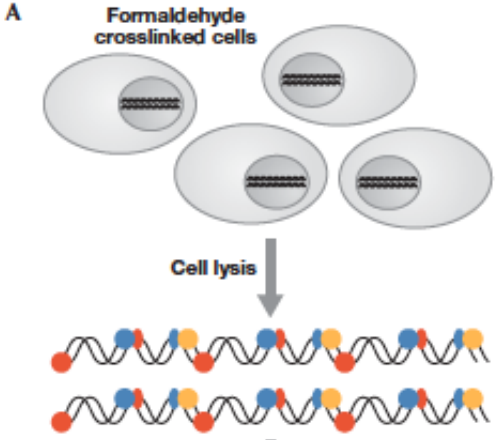
Transfer, hybridisation with radioactive gapdh probe

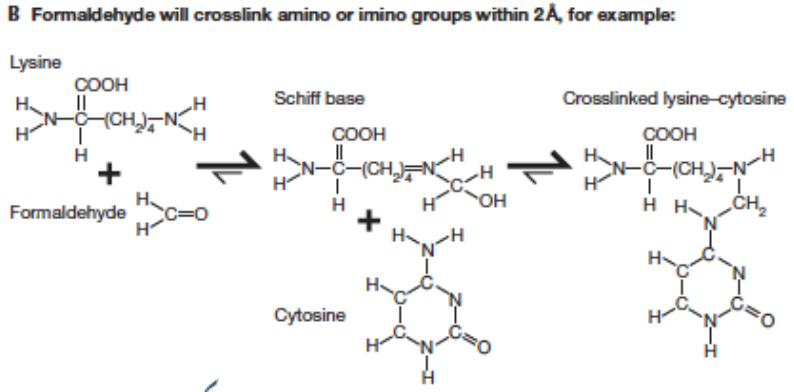
The figure below shows Northern blot analysis of different human tissues. Three probes were used: PSAT, PHGDH and Actin. Actin was used as a loading control, meaning transcription of actin is consistent between human tissues. Which tissue showed the highest level of transcription of the PSAT gene?



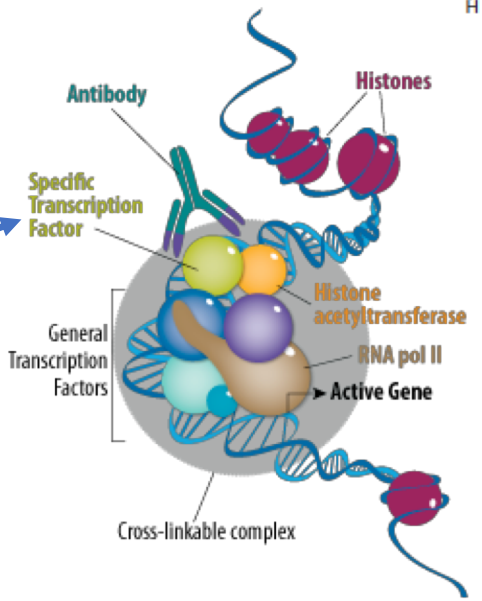
CHROMATIN IMMUNOPRECIPITATION

1. Cross linking with FA



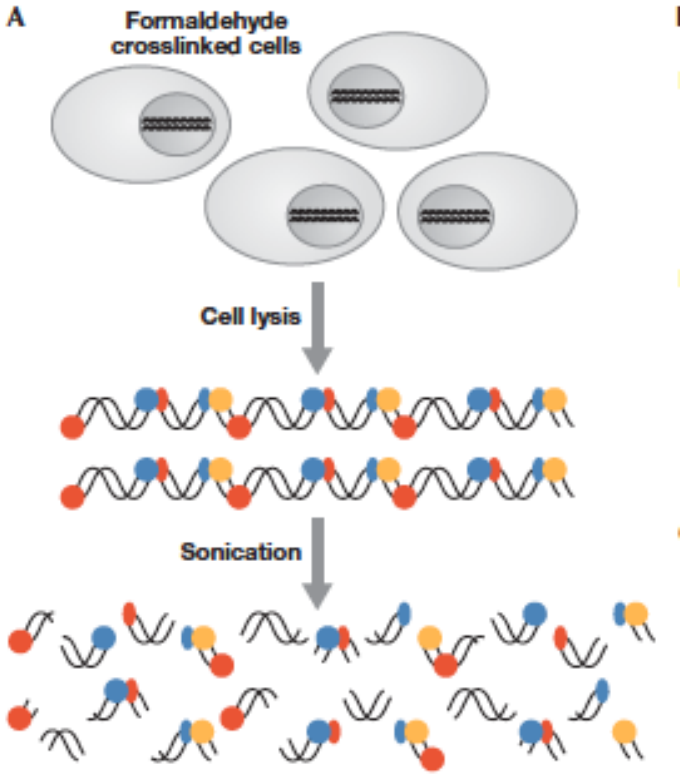


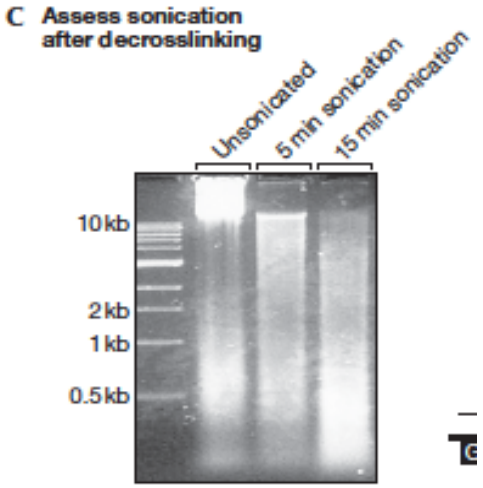
Question: Does a Transcription factor bind to a specifc gene promoter??



CHROMATIN IMMUNOPRECIPITATION

2. Cross linking followed by sonication (fragmentation of chromatin)



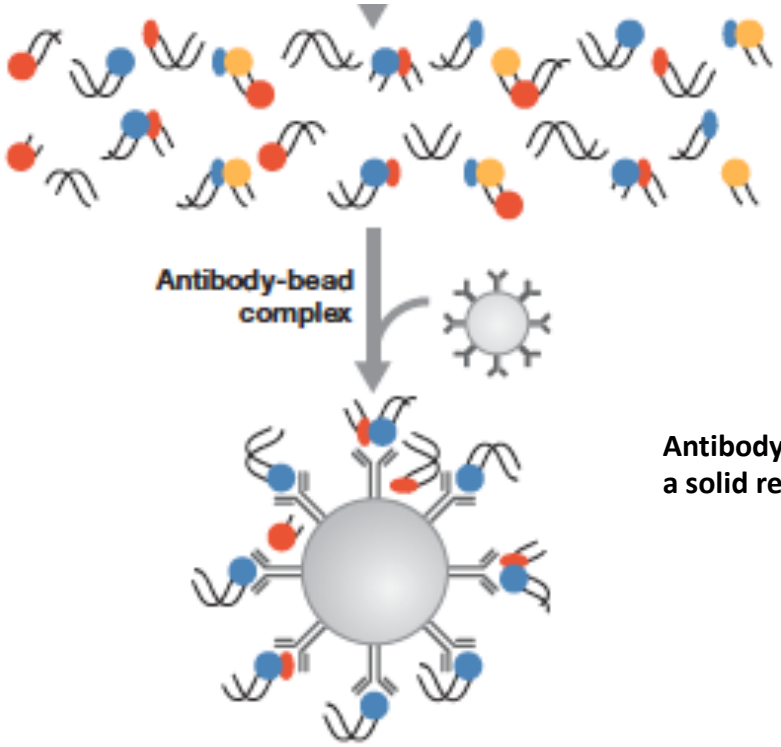


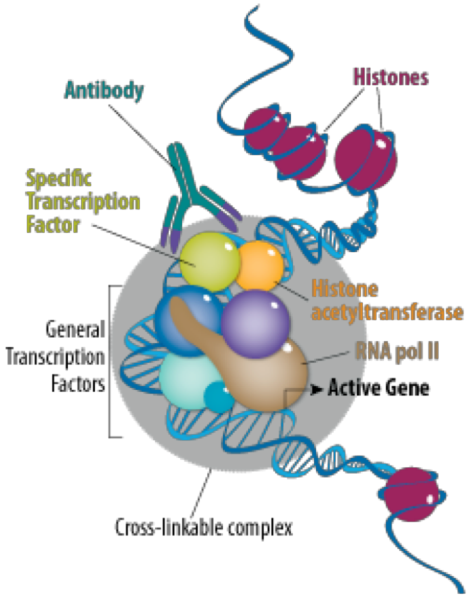
Chromatin is bulky → needs to be cut into small pieces to become soluble

CHROMATIN IMMUNOPRECIPITATION

3. Immunoprecipitation (IP)

The protein of interest is immunoprecipitated together with the crosslinked DNA: Modified histones; epignetic writers, epigentic readers





Antibody is coupled to a solid resin (agarose, magentic beads)

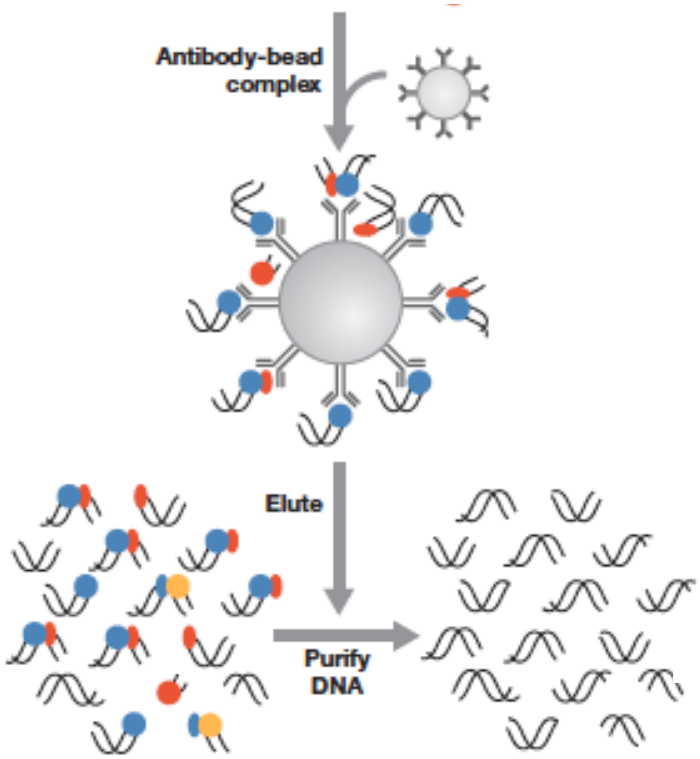
CHROMATIN IMMUNOPRECIPITATION

4.Decrosslinking of PFS crosslinked chrmatin and and purification of the DNA

-Reverse the FA: 65C, high salt concentrations: crosslink break

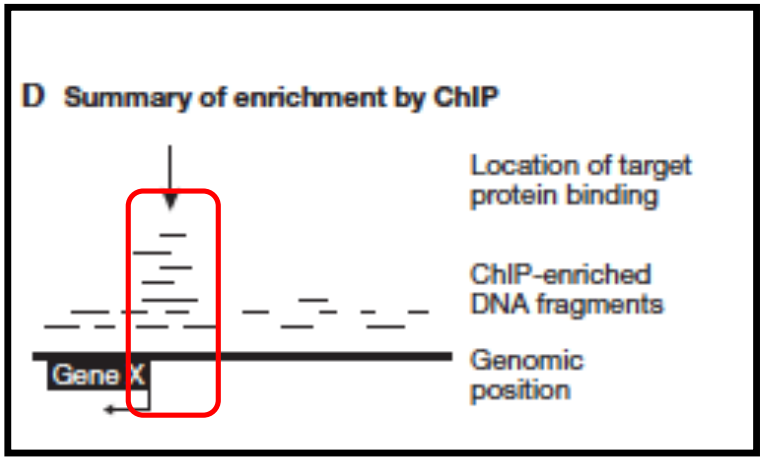
-RNAse and Protease treatment

- Purification of DNA



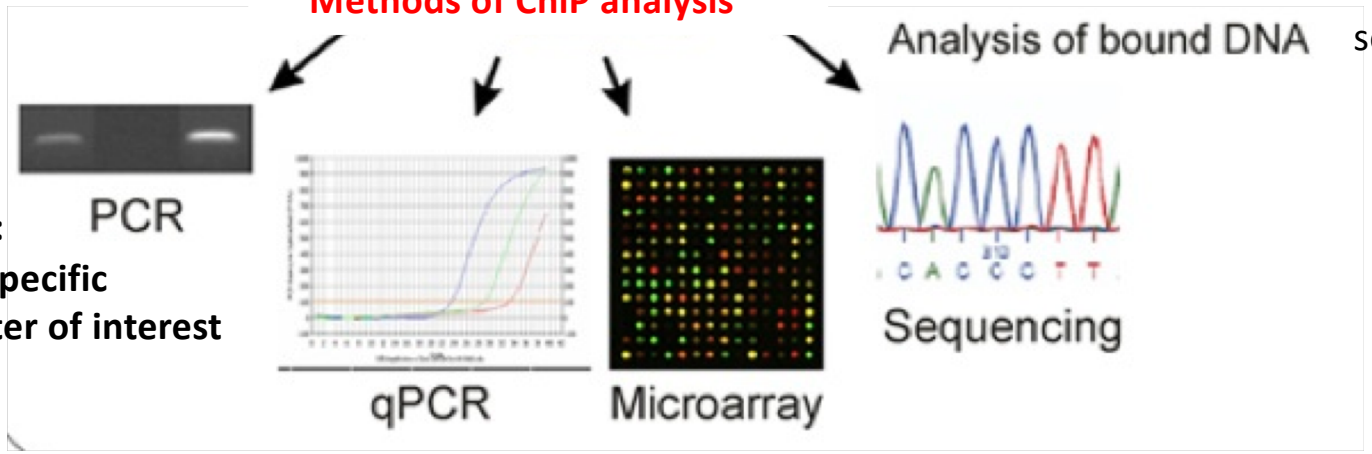
CHROMATIN IMMUNOPRECIPITATION

Identification of DNA regions associated with the protein/modification of interest



If TF specifically binds to the promoter,
ChIP gives a high number of genomic fragments in this region
(remember: DNA is fragemted in a non-sequence specific manner by sonication)

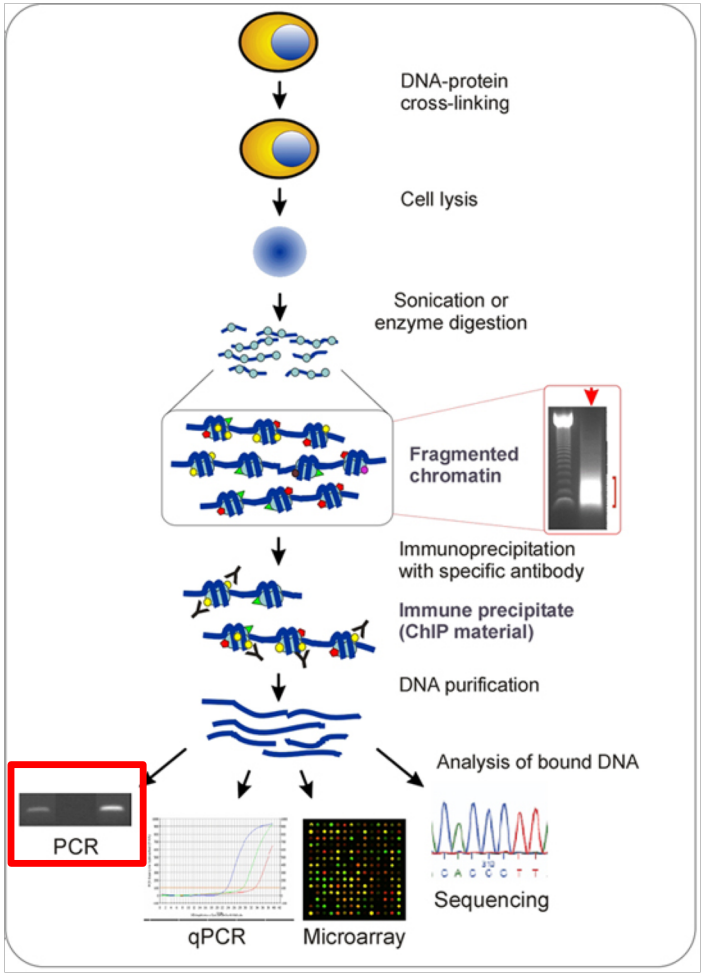
Methods of ChiP analysis



Easiest detection method:
Make PCR using primers specific
For the respective promoter of interest

Lecture 3: Hallmark discovery and analysis of histone modifications

CHROMATIN IMMUNOPRECIPITATION (ChIP) → DETAILLED ANALYSIS
→ Localization of protein at a defined region - seqeunce



1. Crosslink chromatin
(treatment of cells with Paraformaldehyde

2. Sonicate crosslinked cells

4. Recover antibody-chromatin complexes using a resin that binds to the constant region of antibodies

5. Elute chromatin at high salt concentration and revert crosslinks at high temperature

6. Digest protein with protease K and RNA with RNase

5. Elute chromatin at high salt concentration and revert crosslinks at high temperature

6. Digest protein with protease K and RNA with RNase

7. Purify DNA and precipitate DNA

8. Measure the amount of immunoprecipitated DNA By PCR; microarray or NGS

METHODS:

Quantitative PCR: design PCR oligos to amplify defined sequences in immunoprecipitates. Per PCR reaction only one Locus can be examined by real-time PCR

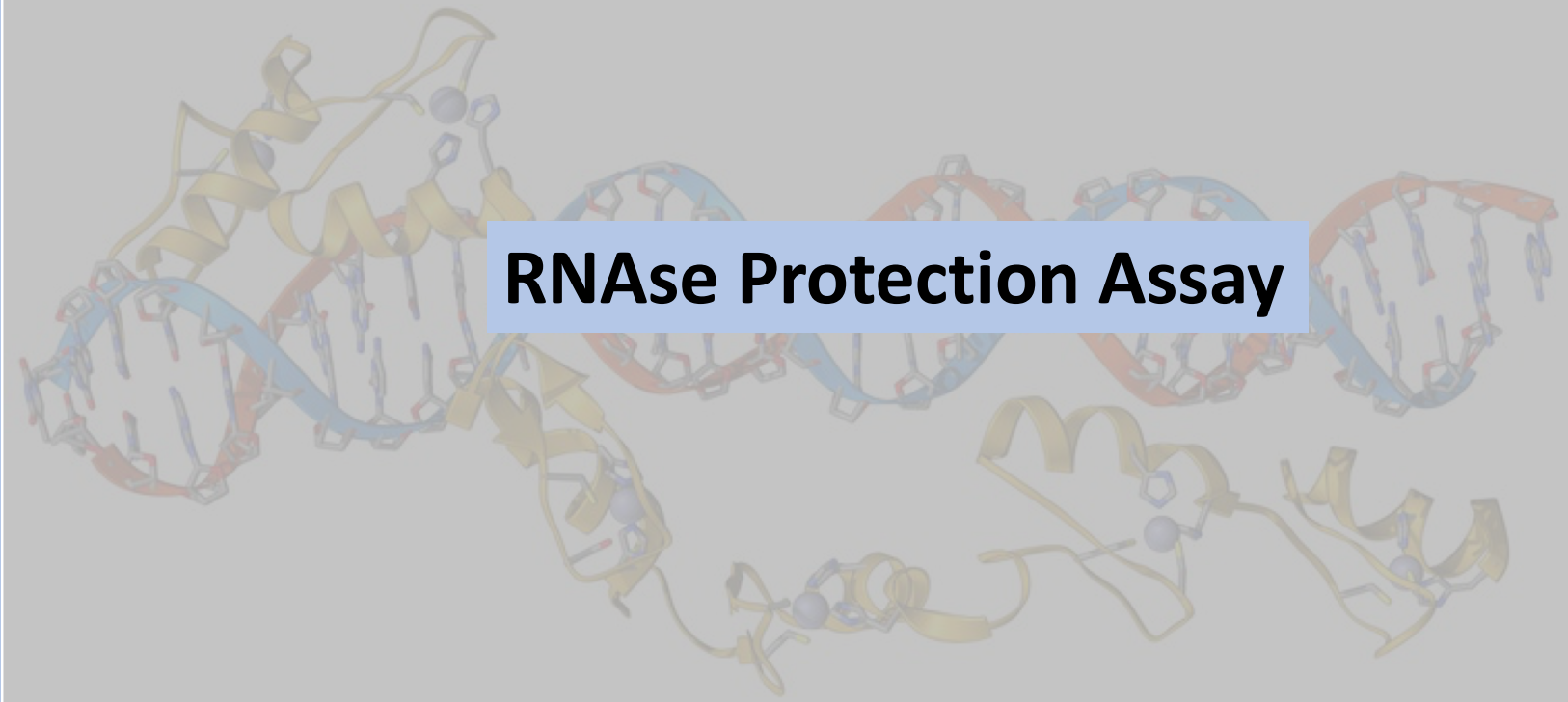
FROM

DNA

TO

RNA

Northern Blotting



RNAse Protection Assay

EMSA for RNA (same as DNA)

RNAse protection assay

The RNase Protection Assay (RPA) is a specific, sensitive, and qualitative method for the detection, mapping, and quantitation of specific mRNAs.

The RPA is at least 10-fold more sensitive than Northern blot analysis and is more accurate and direct than RT-PCR analysis.

In addition, RPA provides information that cannot be obtained reliably by other methods:

For example, **mapping transcription start sites, studying intron-exon junctions**, and **detecting very small differences in related transcripts** can be achieved with RPA. The RNase protection assay can be performed with either total RNA or poly A+ RNA, and the results are not dependent upon having purified or non-degraded RNA

CONCEPT: a labeled, single-stranded antisense RNA probe is allowed to hybridize to the target RNA. RNA probe molecules and transcripts that do not form hybrids are degraded by a mixture of RNases. The final inactivation of RNases and the precipitation of protected RNA hybrids are performed simultaneously. Electrophoresis followed by autoradiography reveals the presence, size, and relative level of RNA that was protected by the antisense probe.

Anti-sense RNA preparation

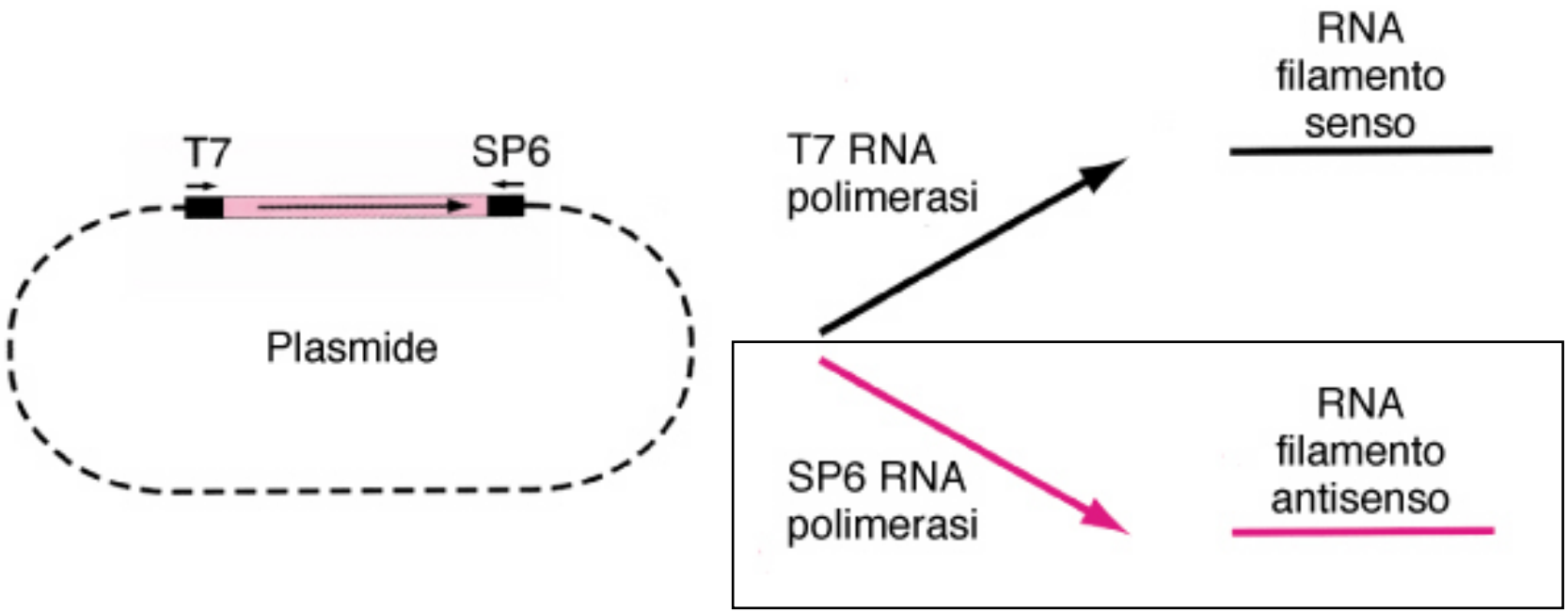


Figura 13.2 Produzione di un RNA antisenso

- Run on polyacrylamide gel
- Cut out radioactive, in vitro trasncribed RNA
- Purify RNA
- Deterimine concentration by measuring incorporated radioactive nucleotides

Radioactive ^{32}P-NTP is incorporated during in vitro transcription (body labelling)

RNAse protection

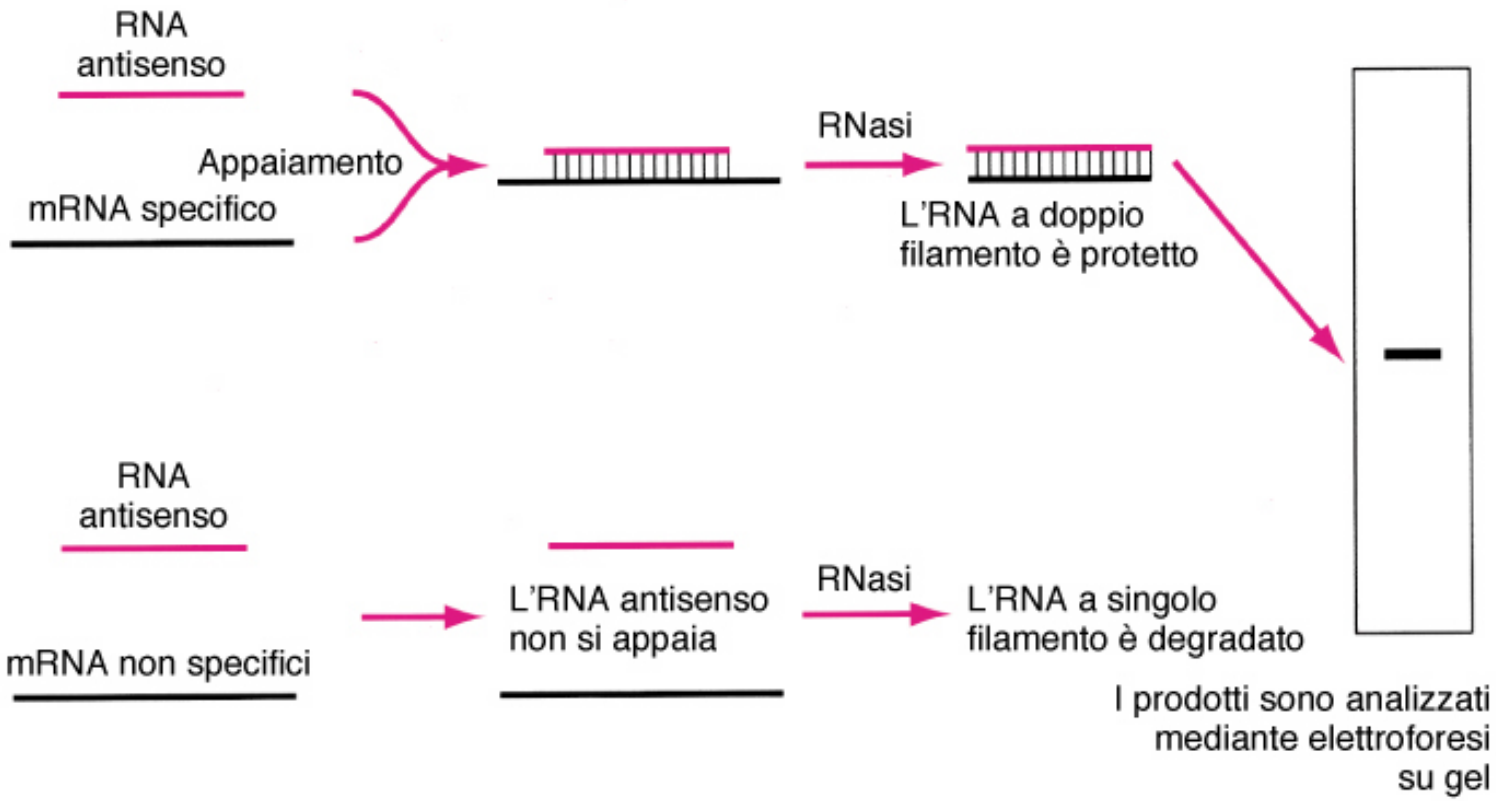
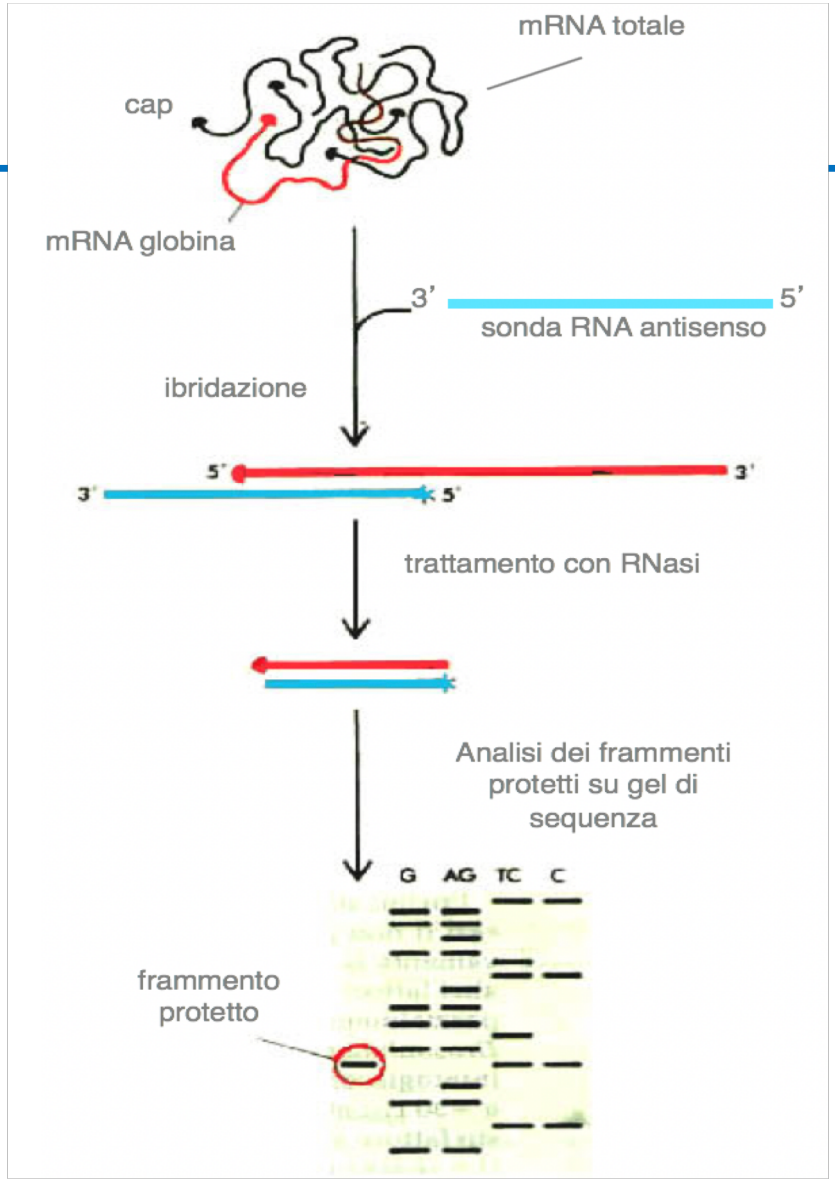


Figura 13.3 Analisi mediante protezione all'RNasi

RNAse protection



RNAse protection assay protocol

RNase Protection Assay

1) Isolation of RNA sample(s) to be examined for target expression
2) Synthesis of a labeled antisense RNA probe complementary to a several-hundred-base region of the target mRNA,
3) Hybridization of the labeled probe to a total RNA sample,
4) Treatment of the sample with single-strand-specific RNase to degrade unhybridized probe and target
5) Separation of the remaining protected probe::target hybrids on a denaturing polyacrylamide gel
6) Detection/quantitation of the RNase-resistant "protected" probe