

Isolation of Nucleic acids

The choice of the isolation protocol depends on:

- Tissue Type*
- Yield provided by the method used*
- Purity and length of isolated nucleic acids*
- Cost*
- Possibility of automation*



General procedures: liquid phase



- *Cell Lysis : digestion of cellular and nuclear membranes and proteins (Proteinase K)*
- *Separation of nucleic acids from proteins (phenol extraction: chloroform): aqueous phase / interphase / phenol phase: chloroform*
- *DNA / RNA precipitation (alcohols)*
- *Spectrophotometric quantification and purity measurement ($A_{260\text{ nm}}$ and $A_{280\text{ nm}}$)*

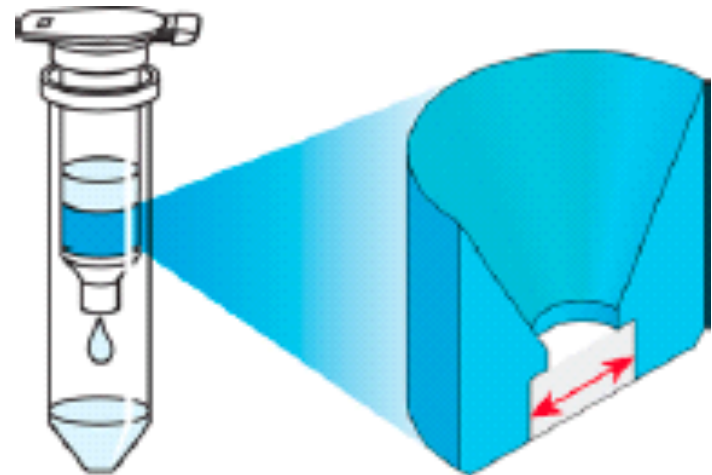
Solid phase

Use of extraction Kit based on using columns with specific filters for nucleic acid binding

Digestion with proteinase K

Washing for elimination of proteins

DNA elution



DNA extraction from fresh tissues

§-Tissue homogenization (or cells washing)

- Tissue homogenization or pellet cells and wash it with PBS.

§-Proteinase K digestion

- Put in 10X volume Proteinase K solution with SDS and enzyme.

Proteinase K solution: Proteinase K final 1 mg/ml + 10 mM Tris-HCl pH 7.4 + 10 mM EDTA + 150 mM NaCl + 0.4 % SDS (add just before use)

- O.N. incubation at 37-45°C with stirring (minimum 4 hrs).

§-Phenol/Chloroform extraction

- Extraction with phenol-Tris (pH 8)/chloroform (50%/50%) 1:1 in volume.

§-DNA precipitation

- Precipitate aqueous phase w/ 1 volume of isopropanol. Wash w/ 70% ethanol.

§-DNA pellet solubilization

- Recover pellet in H₂O or TE buffer (better) and store at -20° C.

DNA extraction from blood

§- Red cells lysis (blood in citrate, heparin or EDTA)

***-Add 45 ml lysis buffer to 5 ml blood, mix by inversion and leave at RT for 5'.
Centrifuge X 1000 g at 4°C for 6'.***

Red cells lysis buffer: 0.32 M saccharose + 10 mM tris-HCl pH 7.5 + 5 mM MgCl₂ + 1% triton X 100.

-Wash the pellet with PBS and centrifuge as before.

§- Proteinase K digestion

***-Recover pellet in 600 µl of buffer A and add SDS final 0.5% and proteinase K
1 mg/ml. Incubation at 37°C O.N. (at least 4 hrs).***

Buffer A: 10 mM Tris-HCl pH 8 + 400 mM NaCl + 2 mM EDTA.

§- Protein salting-out

***-Add 220 µl of sature solution of NaCl, vigorous stirring for 15'' and centrifuge
15' X 14000 rpm at 4° C.***

§- DNA precipitation

***-Recover supernatant and precipitate with 1 volume isopropanol at rt with
immediate precipitation. Centrifuge and wash pellet with 1 ml of 70% ethanol.***

§- DNA pellet solubilization

-DNA is solubilized in 500 µl of TE or H₂O.

DNA extraction from blood- silica use

1. 200 μ L whole blood mixed with 100 μ L lysis buffer^a **or** 25 μ L mononuclear cells were added to 100 μ L binding solution^b. Mix and incubate for 3 min at room temperature. Centrifuge for 15 s at 65 \times *g* and discard supernatant.
2. Add 100 μ L lysis buffer and mix. Centrifuge for 15 s at 65 \times *g* and discard supernatant. Repeat once.
3. Add 100 μ L washing solution^c and mix. Centrifuge for 15 s at 65 \times *g* and discard supernatant. Repeat once.
4. Add 100 μ L absolute ethanol and mix. Centrifuge for 15 s at 65 \times *g*. Discard supernatant and vacuum-dry the pellet.
5. Add 50 μ L elution buffer^d, resuspend the pellet, and incubate for 3 min at 65°C. Centrifuge for 1 min at 65 \times *g*, and transfer supernatant in a clean tube.

^a3 M guanidine thiocyanate, 20 mM EDTA, 10 mM Tris-HCl, pH 6.8, 40 mg/mL Triton® X-100, 10 mg/mL DTT

^b40 mg/mL silica (Sigma) directly suspended in the lysis buffer

^c25% absolute ethanol, 25% isopropanol, 100 mM NaCl, 10 mM Tris-HCl, pH 8.0

^d10 mM Tris-HCl, pH 8.0, 1 mM EDTA

All solutions were prepared in double-distilled water and sterilized by filtration and remain stable for more than six months.

RNA DEGRADATION



RNAses

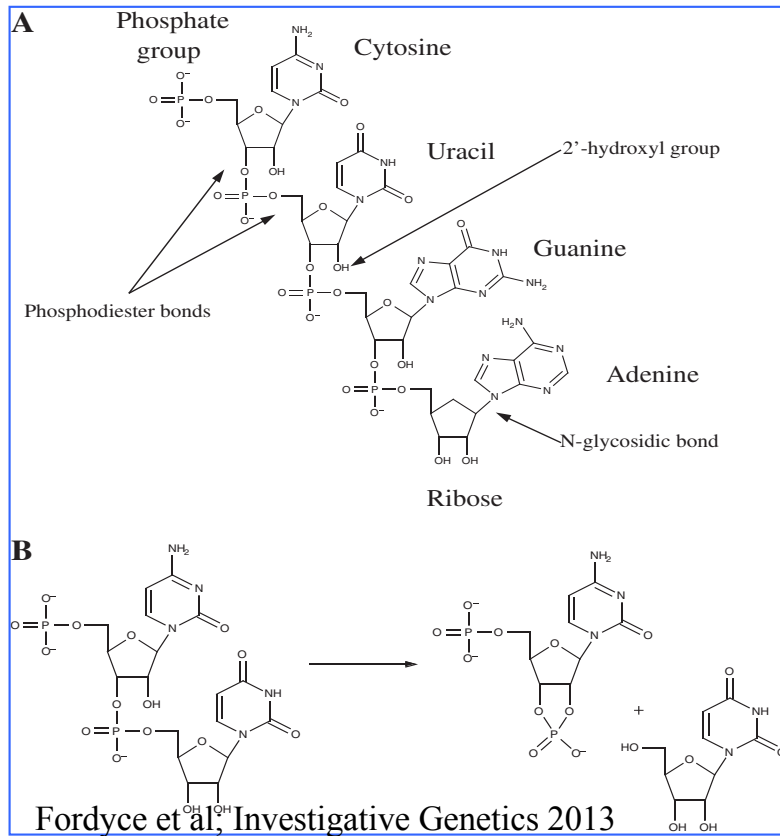


Temperature



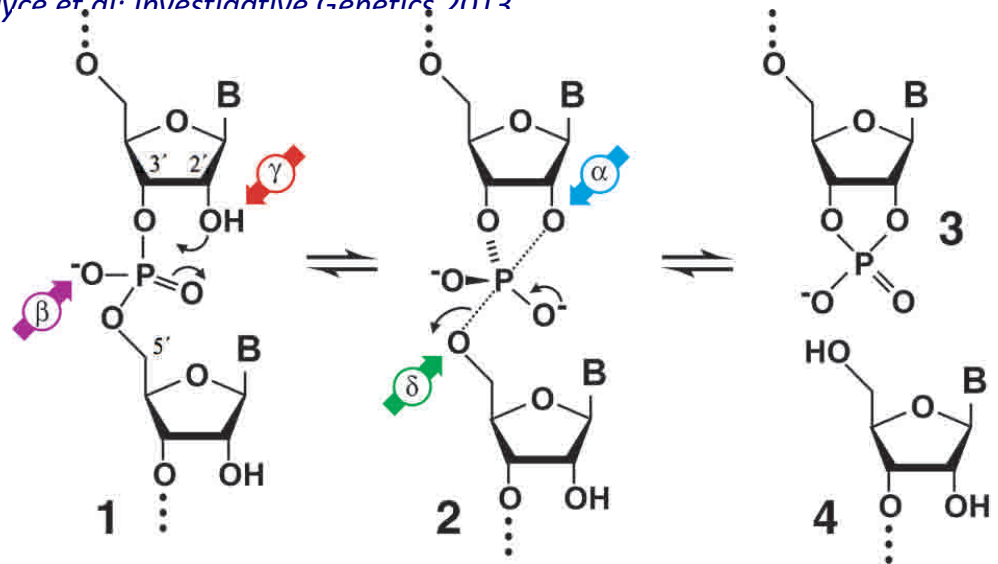
Alkaline environment

RNA degrades differently from DNA



- The 2'-OH allows the RNA molecule to be more easily degraded via hydrolysis than DNA.
- The phosphodiester bond in RNA can be broken during hydrolysis.
- The N-glycosidic bond is stronger in RNA than DNA
- The chemical process of hydrolysis, where the 2'-hydroxyl group has attacked the adjacent phosphodiester bond, cleaving the backbone of the RNA.

For dyce et al., Investigative Genetics 2013



RNAse A increases the rate of RNA cleavage by internal phosphotransfer

3-RNA extraction from fresh tissues

§-Tissue homogenization (or cells washing)

§-Cell lysis

§-Phenol/Chloroform extraction

§-RNA precipitation

§-RNA pellet solubilization

* Chomczynski P; Sacchi N. Single-step method of RNA isolation by acid guanidinium thiocyanate-phenol-chloroform extraction. Anal Biochem, 162: 156-159; 1987

3-RNA extraction from fresh tissues

§-Tissue homogenization (or cells washing)

-For tissues homogenization in liquid nitrogen / wash the cells with PBS (keep always in ice and use ice cold solution).

§-Cell lysis

-Put it in D solution* (1 ml for 20 million cells) with β -mercapto-ethanol (0.36 ml for 5 ml solution) (1 ml per 2.0×10^7 cells) for 5' in ice.

**D solution: 250 g guanidinium thiocyanate + 293 ml of DEPC- H_2O + 17.6 ml of sodium citrate 0.75 M pH 7.0 + 26.4 ml sarcosyl 10% at 65°C.*

§-Phenol/Chloroform extraction

-Add 1/10 volume of Sodium Acetate 2M with pH 4.0-4.5. (acid pH is necessary to obtain a good separation at the interface) and extract w/ 1 volume of phenol/ H_2O and chloroform (7:3 ratio). Keep in ice for 20' and centrifuge for 20'.

§-RNA precipitation

-Add 1 volume of isopropanol, ON at -20°C -centrifuge -Wash w/ chilled 75% EtOH.

§-RNA pellet solubilization

-Resuspend in DEPC H_2O .

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