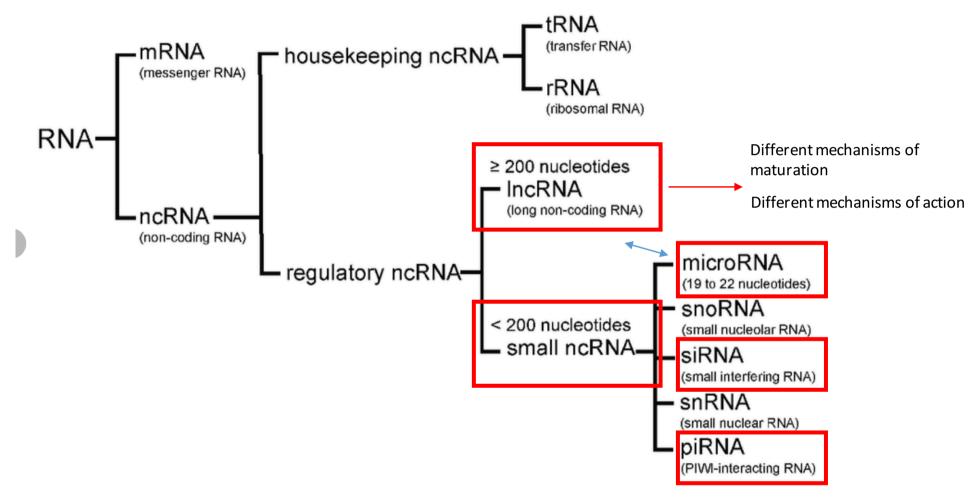
### **DIFFERENT CATEGORIES OF ncRNAs**

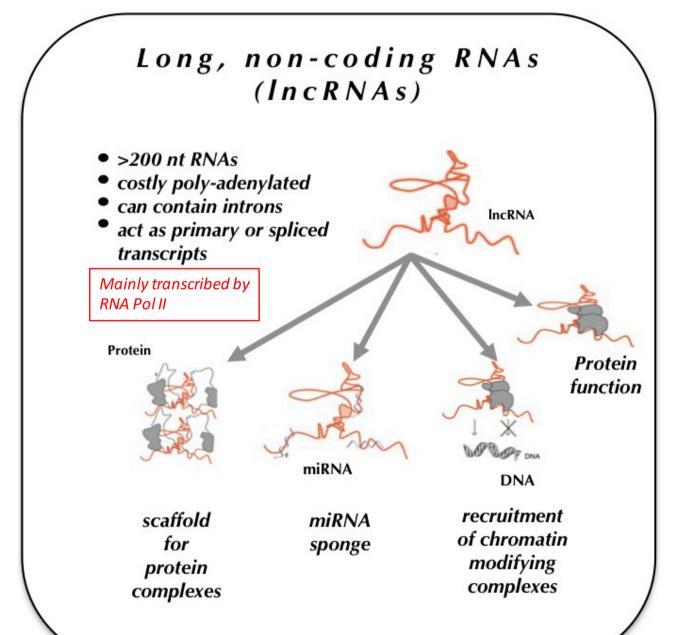


RNA categories. RNAs are divided into two major classes: messenger RNA (mRNA) and non-coding RNA (ncRNA). NcRNAs include housekeeping ncRNA, which consists of transfer RNA (tRNA) and ribosomal RNA (rRNA), and regulatory ncRNA. Regulatory ncRNAs are classified into long ncRNA (lncRNA) and small ncRNA. Small ncRNAs are subclassified into microRNA, small nucleolar RNA (snoRNA), small interfering RNA (siRNA), small nuclear RNA (snRNA), and PIWIinteracting RNA (piRNA).

# **IncRNAs**

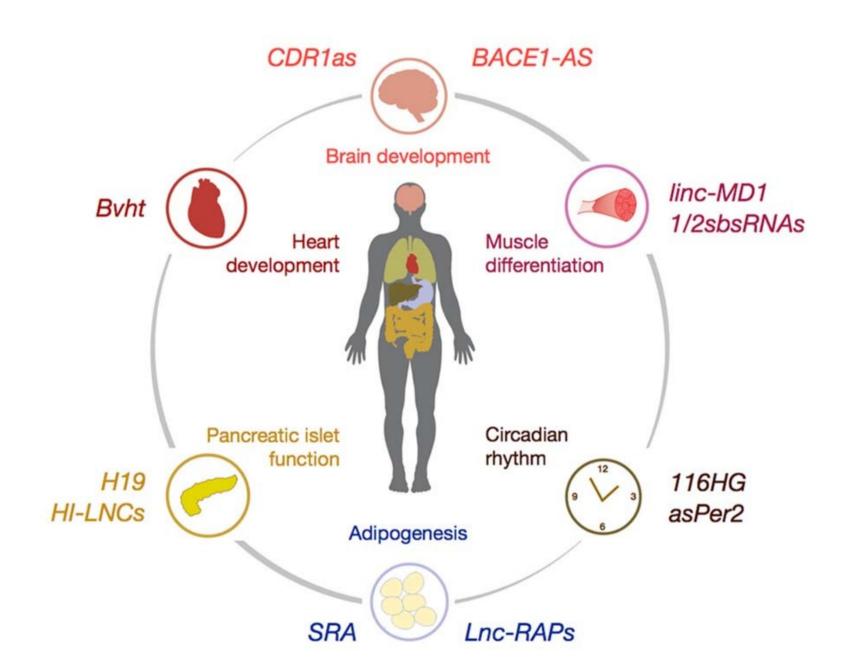
long, non-coding RNAs

# **Characteristics of IncRNAs**



nuclear and/or cytoplasmatic localiztion

# **IncRNAs** in physiology



Resource

# The GENCODE v7 catalog of human long noncoding RNAs: Analysis of their gene structure, evolution, and expression

Thomas Derrien,<sup>1,11</sup> Rory Johnson,<sup>1,11</sup> Giovanni Bussotti,<sup>1</sup> Andrea Tanzer,<sup>1</sup> Sarah Djebali,<sup>1</sup> Hagen Tilgner,<sup>1</sup> Gregory Guernec,<sup>2</sup> David Martin,<sup>1</sup> Angelika Merkel,<sup>1</sup> David G. Knowles,<sup>1</sup> Julien Lagarde,<sup>1</sup> Lavanya Veeravalli,<sup>3</sup> Xiaoan Ruan,<sup>3</sup> Yijun Ruan,<sup>3</sup> Timo Lassmann,<sup>4</sup> Piero Carninci,<sup>4</sup> James B. Brown,<sup>5</sup> Leonard Lipovich,<sup>6</sup> Jose M. Gonzalez,<sup>7</sup> Mark Thomas,<sup>7</sup> Carrie A. Davis,<sup>8</sup> Ramin Shiekhattar,<sup>9</sup> Thomas R. Gingeras,<sup>8</sup> Tim J. Hubbard,<sup>7</sup> Cedric Notredame,<sup>1</sup> Jennifer Harrow,<sup>7</sup> and Roderic Guigó<sup>1,10,12</sup>

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22:1775-1789 © 2012, Published by Cold Spring Harbor Laboratory Press; ISSN 1088-9051/12; www.genome.org

Genome Research 1775 www.genome.org

BASED ON ENCODE DATA -> GENCODE ANNOTATION OF IncRNAS

FIRST DETAILLED, GENOME-WIDE ANNOTATION OF IncRNAs

IncRNAs can be

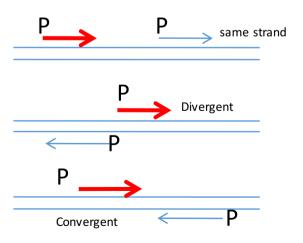
### 1. Intergenic (lincRNA):

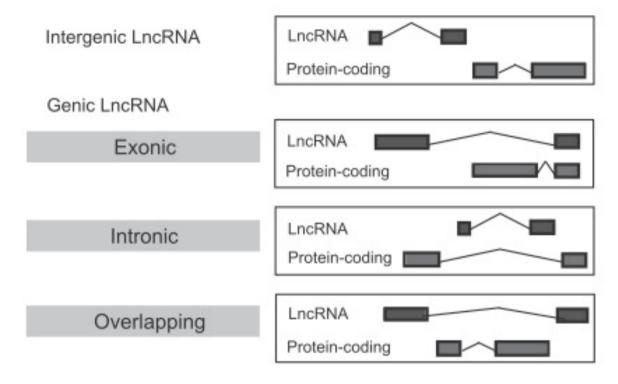
Do not intersect with protein coding gene

#### 2. Genic:

Intersect a protein coding gene

- Exonic
- Intronic
- overlapping





	Gencode	e LncRNAs	trans	cripts	(14,8	80)		
Int	Intergenic (9,518)			Genic (5,362)				
Same Strand	Convergent	Divergent	Exonic (2,411)		Intronic (2,784)		Overlapping (167)	
4,165	1,937	3,416	S	AS	S	AS	S	AS
4,105			NA	2,411	563	2,221	52	115

Closest protein coding gene

IncRNA

P: promoter

Why?

Why?

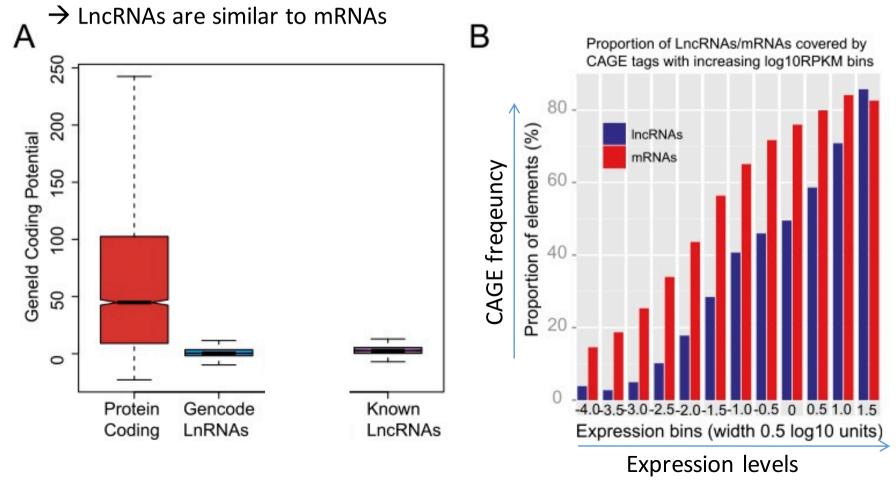
Why?

A. LncRNAs do not have coding potential:

→ Longest possible ORF was searched in mRNA/IncRNAs

CODING POTENTIAL INITIATION - EXPRESSION

B. CAGE tags can be found in IncRNAs → defined transcriptional start site CAGE tag frequency increases with increased IncRNA expression levels. mRNAs are characterized by more CAGE tags



# TRASCRITTOMICA Schedule lectures— AA 2020/2021

#### October (18 hours) L1: 05.10.2020: 14-16(2h) L2: 08.10.2020: 10:45-12:45 (2h) L3: 09.10.2020: 10:45-12:45 (2h) L4: 12.10 2020: 14-16 (2h) L5: 15.10.2020: 10:45-12:45 (2h) L6: 16.10.2020: 10:45-12:45 (2h) L7: 19.10.2020: 14-16(2h) L8: 22.10.2020: 10:45-12:45 (2h) L9: 23.10.2020: 10:45-12:45 (2h) November (16 hours) L10: 05.11.2020: 10:45-12:45 (2h) Seminar 1 L11: 06.11.2020: 10:45-12:45(2h) Seminar 2 L12: 12.11.2020: 10:45-12:45 (2h) Seminar 3 L13: 13.11.2020: 10:45-12:45 (2h) Seminar 4 L14: 19.11.2020: 10:45-12:45 (2h) Seminar 5 L15: 20.11.2020: 10:45-12:45 (2h) Seminar 6 L16: 26.11.2020: 10:45-12:45 (2h) Seminar 7 L17: 27.11.2020: 10:45-12:45 (2h) Seminar 8 December (12 hours) L18: 04.12.2020: 14-16 (2h) → EXTRA SEMINAR L19: 09.12.2020: 08:45-10:45(2h) Seminar 9 (change with Sistemi Modelli $\rightarrow$ 03.12.2020 (10:45 – 12:45) L20: 10.12.2020: 16-18 (2h) Seminar 10 L21: 11.12.2020: 11-13 (2h) Seminar 11 L22: 17.12.2020: 16-18 (2h) Seminar 12 L23: 18.12.2020: 16-18 (2h) Seminar 13 L24: XX.YY,ZZZZPresentation Studenti fuori Trieste: Seminae 14 January XX: 15.01.2021: 10:45-12:45 (2h) XX: 21.01.2021: 10:45-12:45 (2h) 48 ore = 6CFU

Edificio C1, Aula L

**PPT SLIDES:** 

**MOODLE FEDERALE** 

PASSWORD: Trascrittomica

Prof. Stefan Schoeftner E-mail: sschoeftner@units.it

I only reply to official students' emails: @units.it (no @gmail; @libero....)

Students' representatives: luca.secco@studenti.units.it

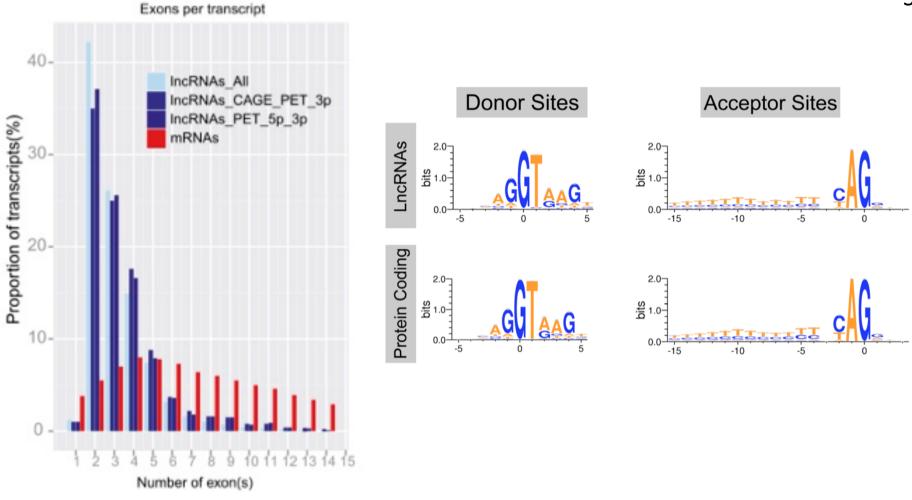
Students' representatives: federico.fierli@.studenti.units.it

# TRASCRITTOMICA **STUDENT SEMINARS**

- 1. Reduce from 17  $\rightarrow$  14 groups
- 2. Form groups only with stundets that do not come to Trieste

Numero gruppo	Studente 1 + email istituzionale	Studente 2 + email istituzionale	Studente 3 + email istituzionale
1	Eleonora Capezzali s266393@ds.units.it	Viviana Ippolito viviana.ippolito@studenti.units.it	Giulia Canarutto s231374@ds.units.it
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3	Agostino Campanile AGOSTINO.CAMPANILE@studenti.units.it	Alessia Pesaresi alessia.pesaresi@studenti.units.it	Patrik Rossi patrik.rossi@studenti.units.it
4	Antonio Manai s231230@ds.units.it	Beatrice Alessandrini beatrice.alessandrini@studenti.units.it	
5	Sonia Panico s268005@ds.units.it	Fracesca Cuomo s267148@ds.units.it	Lara Favero s239076@ds.units.it
6	Luca Secco LUCA.SECCO@studenti.units.it	Sara Maggiore SARA.MAGGIORE@studenti.units.it	Francesco Furio D'Amico FRANCESCOFURIO.D'AMICO@studenti.units.it
7	Silvia Aldrovandi silvia.aldrovandi@studenti.units.it	Matteo Colombo matteo.colombo@studenti.units.it	Richard Camara s271237@ds.units.it
8	Nicoletta Franco NICOLETTA.FRANCO@studenti.units.it	Camilla D'Angelo CAMILLA.D'ANGELO@studenti.units.it	Mbarsid Racaku s265766@ds.units.it
9	Catello Guida s270324@ds.units.it	Domenico Loperfido domenico.loperfido@studenti.units.it	Angelo Acito angelo.acito@studenti.units.it
10	Karim Rahhali KARIM.RAHHALI@studenti.units.it	Elena Petrini ELENA.PETRINI@studenti.units.it	William Bongiovanni WILLIAM.BONGIOVANNI@studenti.units.it
11	Giulia Visani s269271@ds.units.it	Roberta Noè s270858@ds.units.it	Camilla Volponi s270857@ds.units.it
12	Alessia Ferraro alessia.ferraro@studenti.units.it	Cinzia Bessone cinzia.bessone@studenti.units.it	Elena Di Mattia elena.dimattia@studenti.units.it
13	Federico Fierli federico.fierli@studenti.units.it	Domiziano Dario Tosi domizianodario.tosi@studenti.units.it	Giuseppe Castiglione giuseppe.castiglione@studenti.units.it
14	Rossana Putino rossana.putino@studenti.units.it	Gloria Leva gloria.leva@studenti.units.it	Elena Peresani s230866@ds.units.it
15	Camillo Balanzin s239118@ds.units.it	Michele Esposito s260507@ds.units.it	Martin Belliu MARTIN.BELLIU@studenti.units.it
16	Elisabetta Molteni ELISABETTA.MOLTENI@studenti.units.it	Federica Ruggiero FEDERICA.RUGGIERO@studenti.units.it	Maria Alessia Cucco s260331@ds.units.it
17	Ilaria Frascolla ILARIA.FRASCOLLA@studenti.units.it	Elisabetta Giordano ELISABETTA.GIORDANO@studenti.units.it	Sefora Naomi Agrò SEFORANAOMI.@studenti.units.it

**SPLICING** 

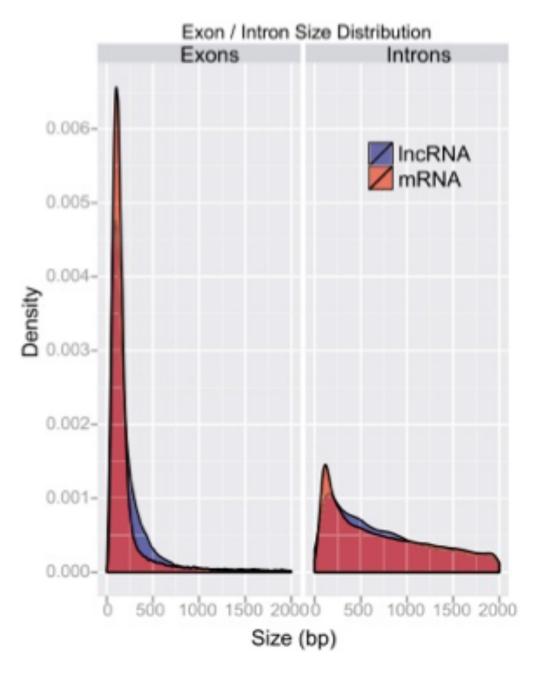


#### **SPLICING:**

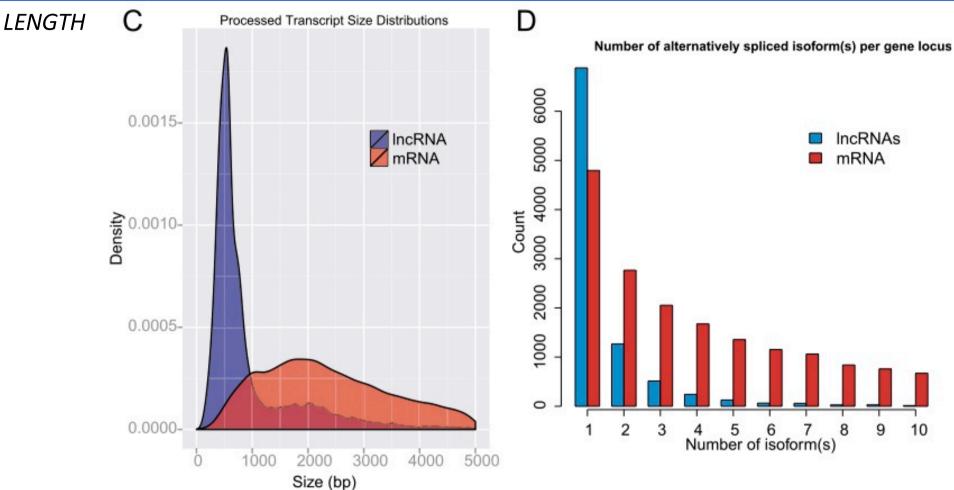
- 98% of IncRNAs are spliced,
- Relevant Splice-site prerequisites at splice donor/acceptor are conserved
- Remarkable tendency of IncRNAs to have only 2 exons (42%, mRNAs: 6%)

BUT: LncRNAs contain fewer INTRONs!! most lncRNA have only 1 intron!!!; mRNAs 3-6

Introns/Exons from IncRNAs are slightly longer



EXON INTRON LENGTH



- IncRNAs are on average much shorter: ca. 500nt
- mRNAs are longer and have wider size distribution

- IncRNAs are uniform → little alternative splicing
- mRNAs: large variety of alternative splicing

**ISOFORMS** 

#### **CONSERVATION**

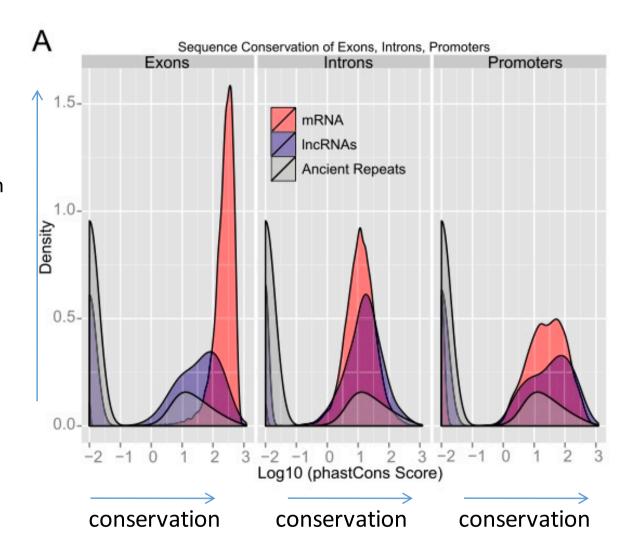
#### **EXONS:**

mRNA: high conservation IncRNA: reduced conservation **But:** conservation is higher than mRNA intron conservation

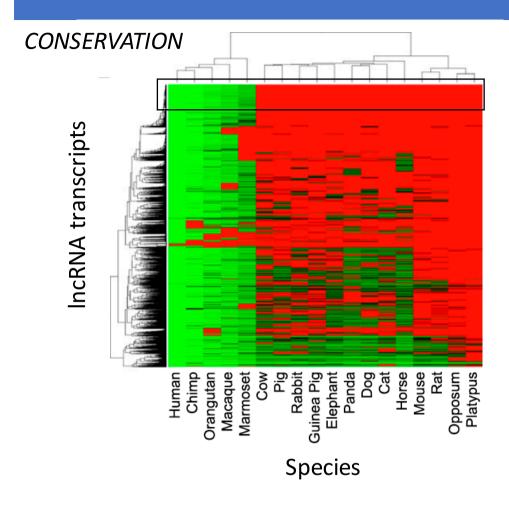
#### **INTRONS:**

mRNA: higher conservation

than IncRNAs

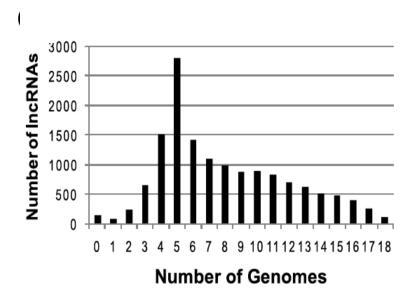


# **Evolutional conservation of IncRNAs**



BLAST of human lncRNAs against all available mammalian genomes and identify homologous genes in other species:

30% of IncRNA transcripts (n = 4546) appear to be primate specific (green=high conservation).



Most IncRNA transcripts are conserved in 4-6 species

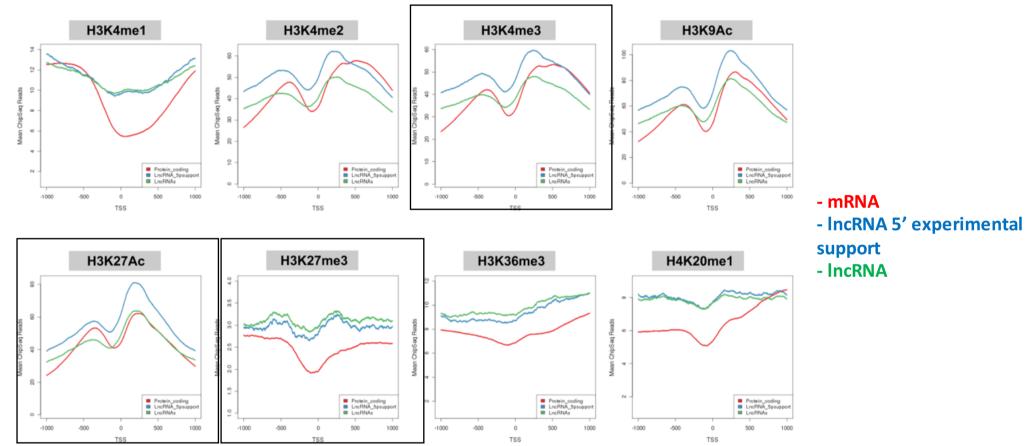
A total of **0.7%** (101) of transcripts appear to be **specific to the human** lineage.

A similar number (134; 1.0%) is found in all of the 18 species analyzed here.

IncRNAs show relevant species specificity

# Chromatin signatures at transcriptional start sites are conserved between mRNAs And IncRNAs: MOST IncRNAs ARE TRANSCRIBED BY RNA Pol II

**EPIGENTICS** 



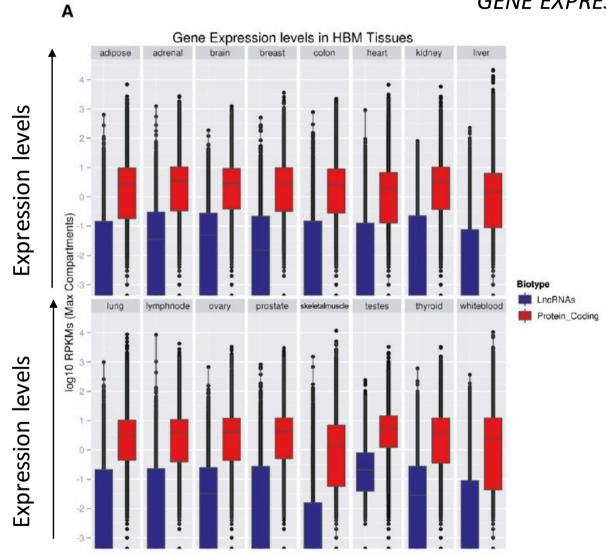
Chromatin signatures around TSS of protein-coding and lncRNA transcripts expressed in the same cell lines where the signatures were monitored by ChipSeq. Shown on the y-axis is the average density of reads covering the TSS of various gene sets, with position plotted on the x-axis (bp relative to positive strand TSS). Protein coding genes are plotted in red, Gencode v7 lncRNAs in green, and lncRNAs with 5' experimental support (n=2,793) in blue. N.B. A more extensive analysis of histone modifications in multiple cell types is available at http://big.crg.cat/bioinformatics and genomics/lncrna data.

K562 human immortalised myelogenous leukemia cell line

NOTE: Histone exclusion at TSS: (Not shown: RNA Pol II peaks at transcription start site of most lncRNAs)

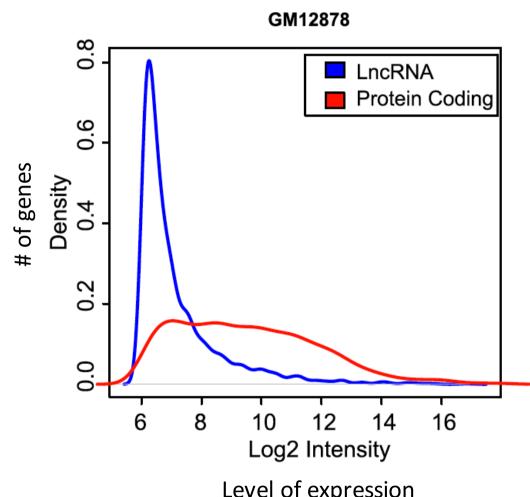
Absolute expression of IncRNAs is much lower than mRNA expression

GENE EXPRESSION



GENE EXPRESSION

**Absolute expression** of IncRNAs is much lower than mRNA expression



Level of expression

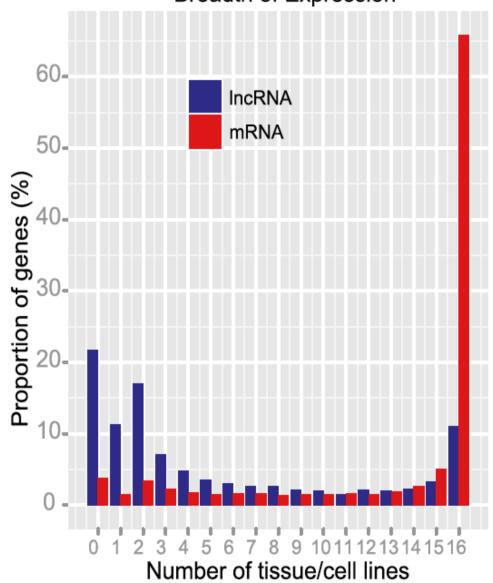
#### SPECIFICITY OF GENE EXPRESSION

Breadth of Expression

mRNAs show broad expression in different tissues – 65% of mRNAs are expressed in 16 tissues

IncRNAs show tissue/cell specific expression – 50% of IncRNAs expressed in 4-5 tissues.

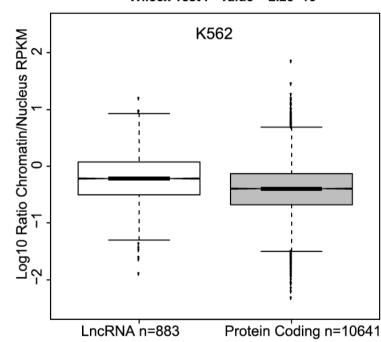
HIGH VALUE OF IncRNAs AS BIOMARKER – EXPRESSION SIGNATURES IN CANCER



LOCALIZATION

# Determination of transcript abundance Chromatin/nucleus

Wilcox Test P-value < 2.2e-16

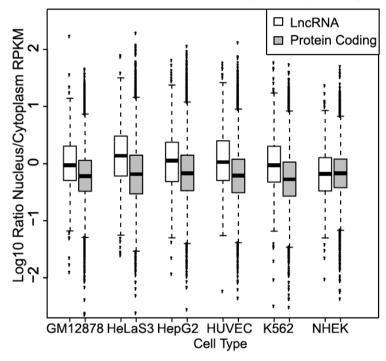


# IncRNAs locate more frequently to chromatin than mRNAs

Data: RNA seq of RNAs located in the nucleoplasma and sticking to chromatin

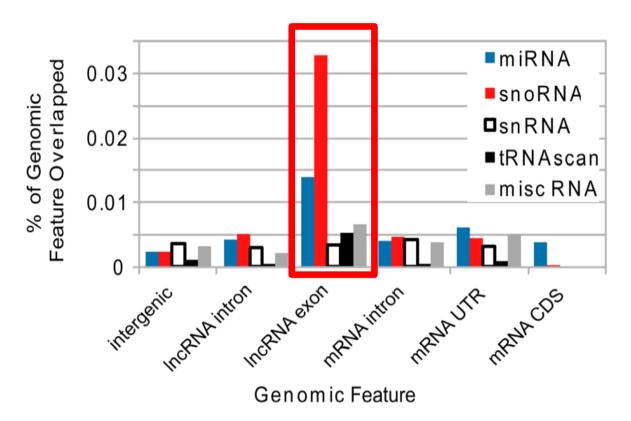
#### Determination of transcript abundance Nucleus/cytoplasma

Wilcox Test P-value < 2.2e-16 for all pairs except NHEK (P=0.7)



# IncRNAs locate more frequently to the nucleus than mRNAs

Data: RNAseq of RNAs located in the nucleoplasma and cytoplasma



**Figure S7: LncRNAs as a source of small RNAs.** Shown is the proportion of nucleotides in exons/introns/(UTRS) from protein-coding genes and lncRNAs that overlap different classes of annotated small RNAs. Note that all values refer to cases where the small RNA is on the same strand as the indicated lncRNA/mRNA.

# IncRNA exons have a remarkable potential to encode functional small RNAs (sRNAs)

# HIGHER THAN EXONS OF mRNAS!!!!

miRNA: small RNAs that target mRNAs to reduce protein expression

**snoRNA**: Small nucleolar RNAs (snoRNAs) are a class of small RNA molecules that primarily guide chemical modifications of other RNAs, mainly ribosomal RNAs, transfer RNAs and small nuclear RNAs.

**snRNA**: Small nuclear ribonucleic acid (snRNA), also commonly referred to as U-RNA, is a class of small RNA molecules that are found within the splicing speckles and Cajal bodies of the cell nucleus in eukaryotic cells.

tRNA: translation

miscRNAs: MiscRNA is short for miscellaneous RNA, a general term for a series of miscellaneous small RNA. It serves a variety of functions, including some enzyme-like catalysis and processing RNA after it is formed. Besides, some of these small RNAs may serve as switches.

IncRNAs represent a big class of functional elements that

- show controlled gene expression
- are processed
- lack protein coding potential
- defined localization
- frequently encode small RNAs
- low overall conservation
- 35% of IncRNAs are primate specific
- expression is rather low but controlled!
- mostly transcribed by RNA Pol II

# FUNCTION: FOR THE VAST MAJORITY OF IncRNAs THE BIOLOGICAL FUNCTION IS UNKNOWN!!!!