

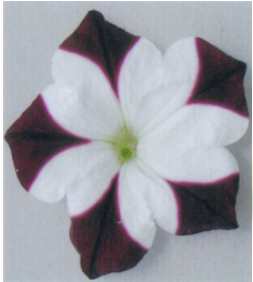
The background features a glowing blue DNA double helix on the left side, set against a grid pattern. Scattered throughout are various molecular models, including ball-and-stick structures and clusters of spheres, all rendered in shades of blue and teal.

LOSS OF FUNCTION APPROACHES

**SIRNA AND
GENOME EDITING BY CRISPS/CAS9**

STEFAN SCHOEFTNER

Co-suppression of gene expression



The discovery of RNAi was preceded first by observations of transcriptional inhibition by antisense RNA expressed in transgenic plants.

Reports of unexpected outcomes in experiments performed by plant scientists in the United States and the Netherlands in the **early 1990s**.



Attempted to overexpress **chalcone synthase** (anthocyanin pigment gene) in petunia. (trying to darken flower color)
Caused **the loss** of pigment .

Further investigation of the phenomenon in plants indicated that the downregulation was due to **post-transcriptional inhibition of gene expression** via an increased rate of mRNA degradation.



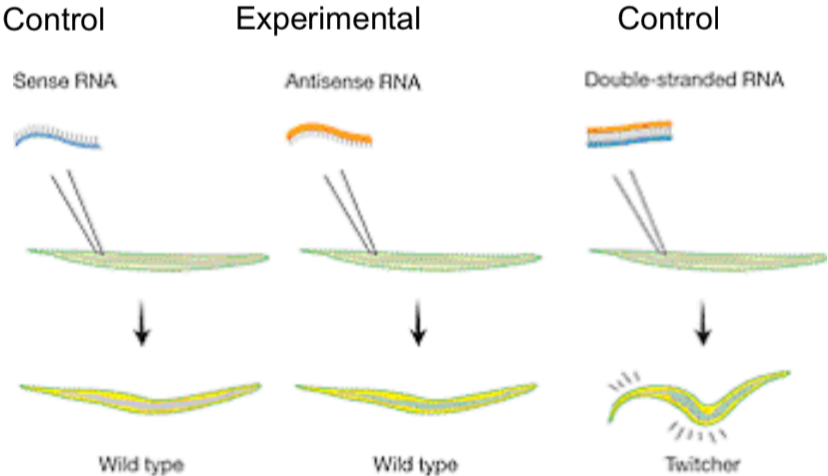
This phenomenon was called **co-suppression of gene expression**, because **suppressed expression of both endogenous gene and transgene** but the molecular mechanism remained unknown

RNA interference (RNAi)

Previously known as co-suppression or post transcriptional gene silencing (PTGS), now is known as **RNA interference (RNAi)** as a process within living cells that moderates the activity of their genes.

Accidental Discovery of RNAi

- Goal: silence endogenous mRNAs with antisense RNA
- The *unc-22* gene encodes a myofilament protein.
- Decrease in *unc-22* activity is known to produce severe twitching movements.



Fire *et al.*
Nature 1997

dsRNA strongly knocked down expression!!!!

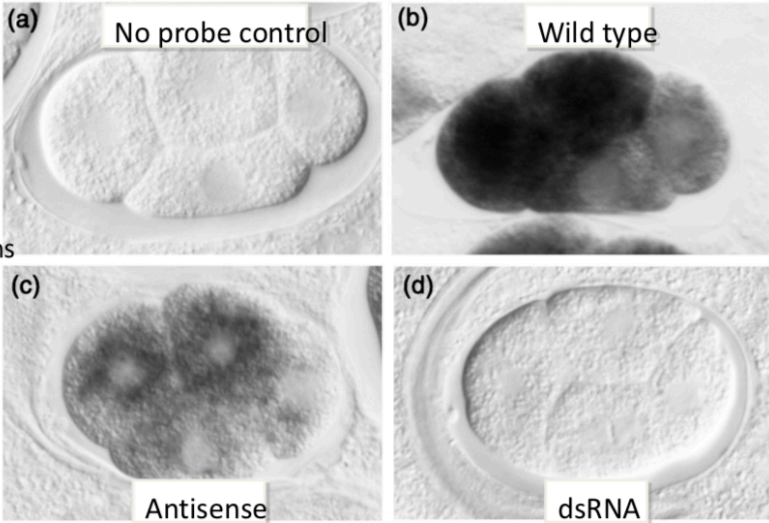
Phenotypic effect after injection of ssRNA or dsRNA (*unc-22*) into the gonad of *C. elegans*.

RNA interference (RNAi)

Injection of dsRNA in *C. elegans* Shown To Cause Destruction of Specific mRNA

• Mello and colleagues, 1998

- Injection in gonads of dsRNA for *mex-3* (abundant RNA) gave much more efficient inhibition in embryos than antisense RNA
- dsRNA had to include exons; introns and promoter didn't work
- Effect was incredibly potent and even spread to other cells within the worm
- Termed 'RNA Interference'
- Incredibly useful as a tool for molecular biology



© Fire, A., S. Xu, M.K. Montgomery, S.A. Kostas, S.E. Driver, and C.C. Mello. Potent and specific genetic interference by double-stranded RNA in *Caenorhabditis elegans*. "Nature" 391 (1998) 1. 3, p. 809. Copyright © Macmillan Magazines, Ltd.

in situ hybridization four-cell stage embryo

Fire et al. Nature 1998

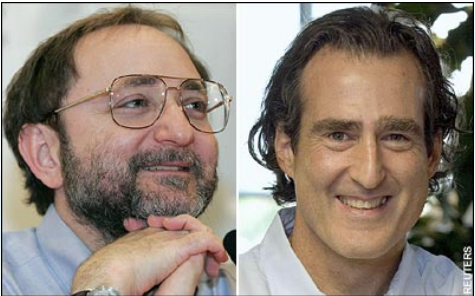
- dsRNA from mature mRNA elicits RNAi
- dsRNA from introns does not
- RNAi results in decreased mRNA levels
- RNAi is heritable (for a few generations)
- RNAi only requires a few molecules of dsRNA per cell
- RNAi is applicable to many different transcripts

RNA interference (RNAi)

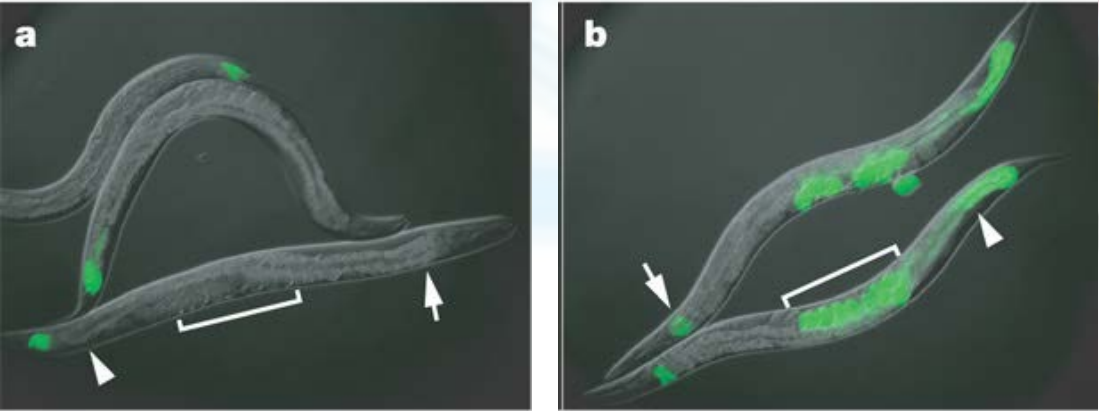
In **2006**, Andrew Fire and Craig C. Mello shared the Nobel Prize in Physiology or Medicine for their work on RNAi in the nematode worm *C. elegans*.

RNAi in *C. elegans*

– Silencing of a green fluorescent protein (**GFP**) reporter in *C. elegans* occurs when animals feed on bacteria expressing GFP dsRNA (**a**) but not in animals that are defective for RNAi (**b**).



Andrew Fire Craig Mello

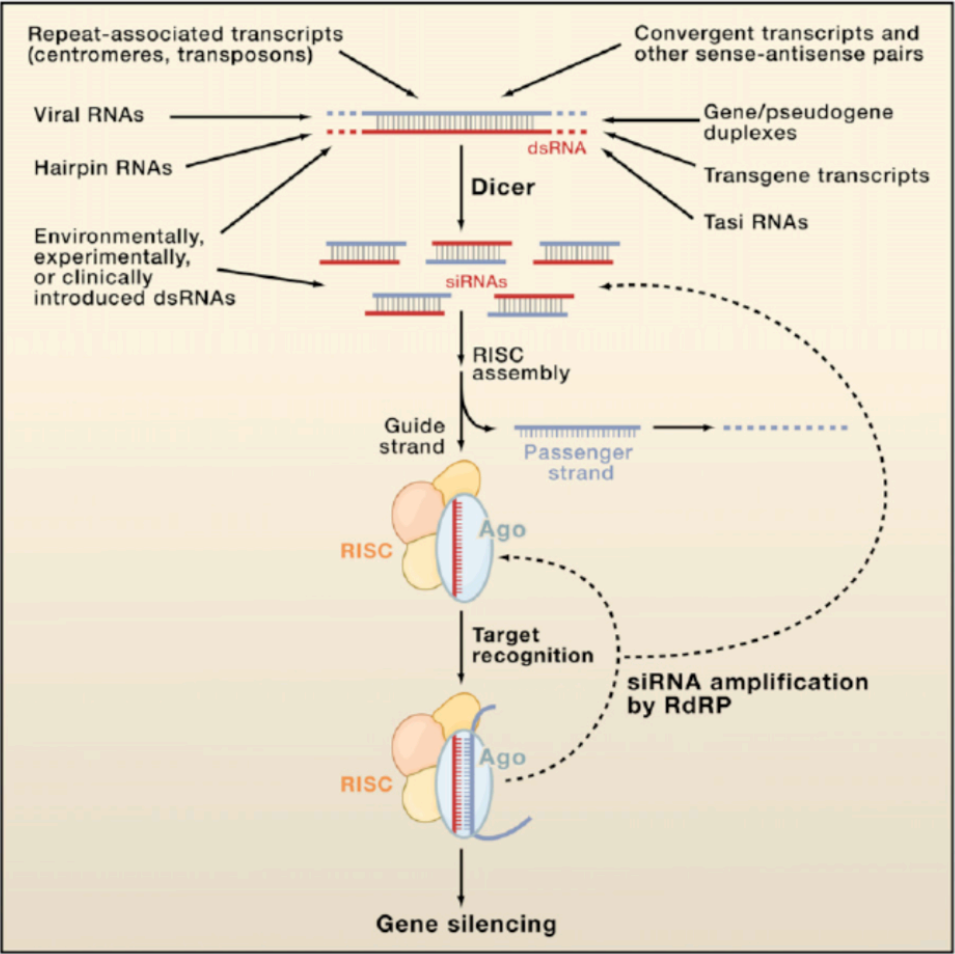


The lack of GFP-positive embryos in a (bracketed region) demonstrates the systemic spread

Sources of dsRNA

Sources of dsRNA

- Some dsRNAs have viral origin, but not all
- Genomic repetitive sequences also are source of siRNA
- Some even regulate other genes (ta-siRNA for trans-acting in plants)
- exo siRNAs (viral etc)
- endo siRNAs –the precursor has a nuclear phase (hairpins, sense-antisense transcripts etc)



Carthew and Sontheimer, Cell (2009) 136, 642-655.

siRNA and miRNA

Two types of RNA molecules involved:

- **small interfering RNA (siRNA)**
- **microRNA (miRNA)**

They bind to other specific mRNAs and modulate their activity.

RNA interference has played an important role in defending cells against parasitic nucleotide sequences – viruses and transposons – but also in directing development as well as gene expression in general.

In 2001 first report of RNAi in MAMMALS

letters to nature

Nature 411, 494 - 498 (2001); doi:10.1038/35078107

Duplexes of 21-nucleotide RNAs mediate RNA interference in cultured mammalian cells

SAYDA M. ELBASHIR*, JENS HARBORTH†, WINFRIED LENDECKEL*, ABDULLAH YALCIN*, KLAUS WEBER† & THOMAS TUSCHL*

* Department of Cellular Biochemistry; and

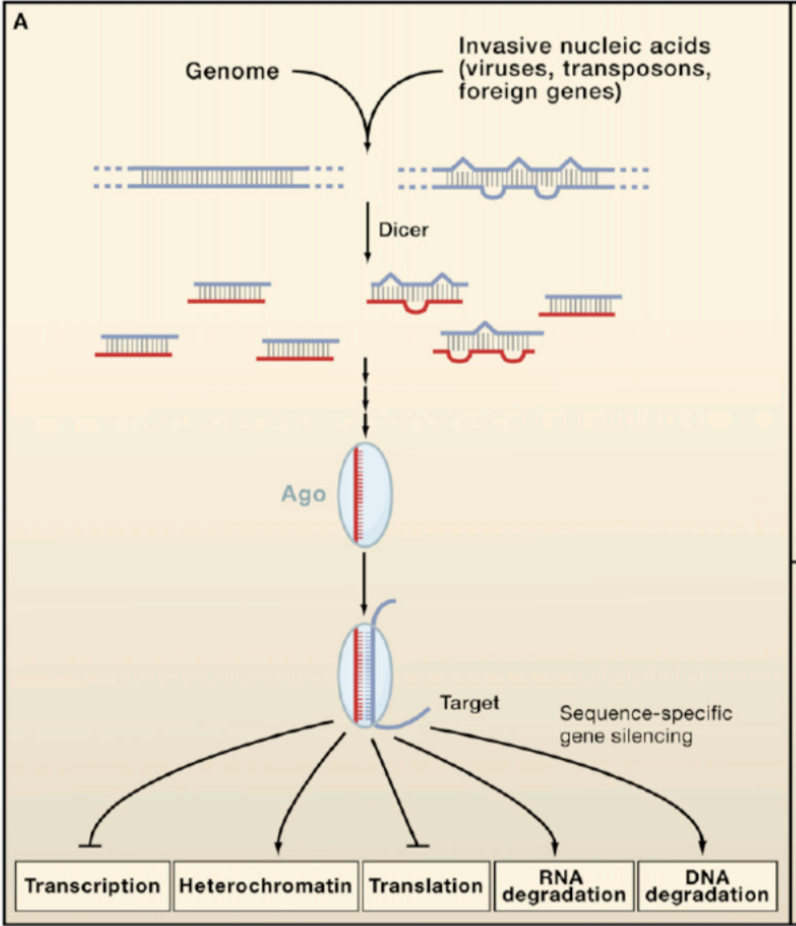
† Department of Biochemistry and Cell Biology, Max-Planck-Institute for Biophysical Chemistry, Am Fassberg 11, D-37077 Göttingen, Germany

siRNA and miRNA

Common Features of Pathways for siRNA and miRNA

Both pathways include:

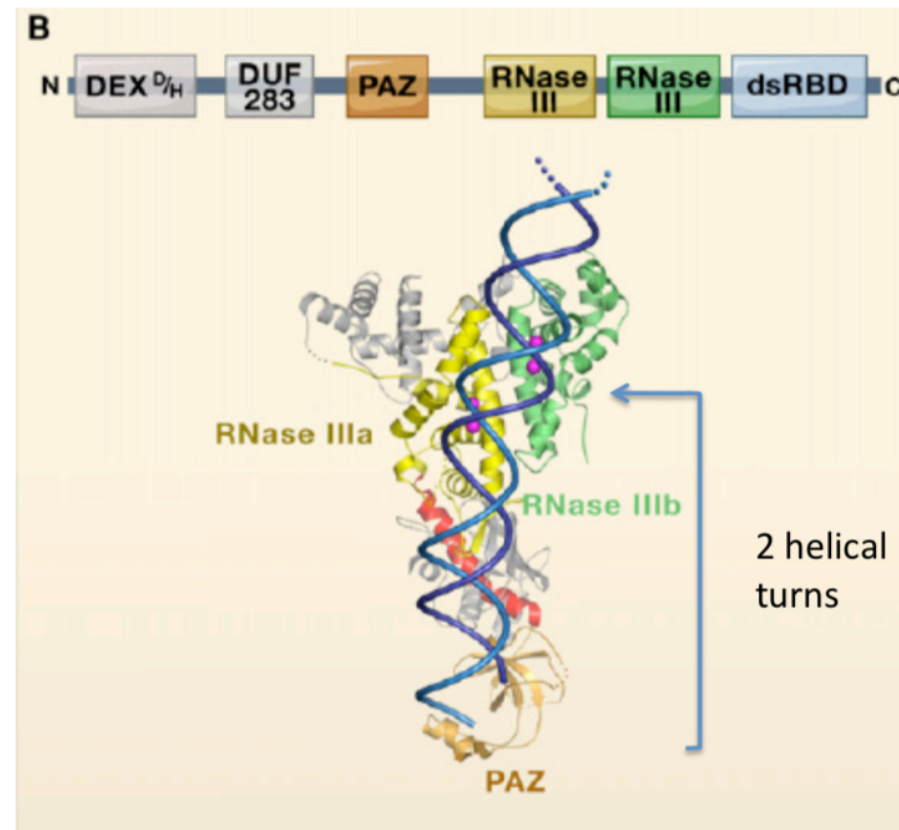
- 1. • dsRNA 'trigger'
- 2. • Dicer processing enzyme
- 3. • Argonaute (Ago)-containing complex to carry out effector function



Carthew and Sontheimer, Cell (2009) 136, 642-655.

DICER: Producer of Small (21-23 bp) RNA fragments

- Structure solved by Doudna and colleagues (2006)
- PAZ domain binds RNA end, RNase III domains cut RNA to produce 2 nt 3'-overhang
- Roles of other domains (not present in structure) remain unclear



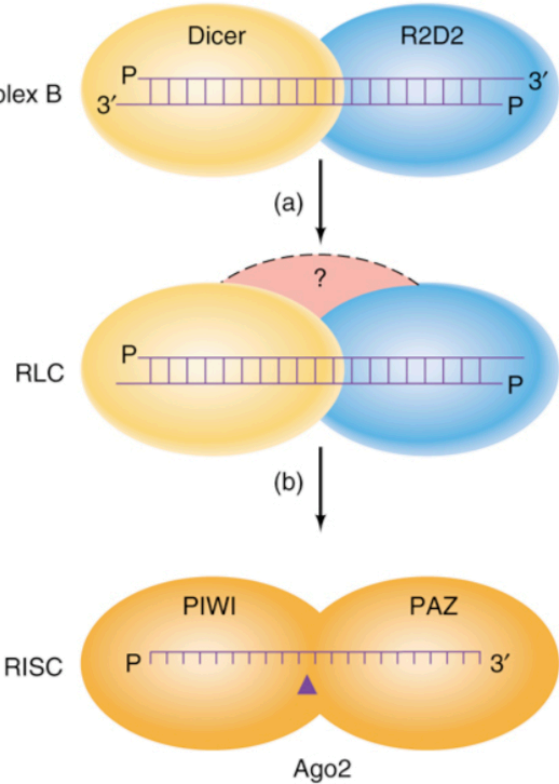
Carthew and Sontheimer, Cell (2009) 136, 642-655.

DICER partners and RISC

Assembly of the RNA-Induced Silencing Complex (RISC) Involves Additional Proteins

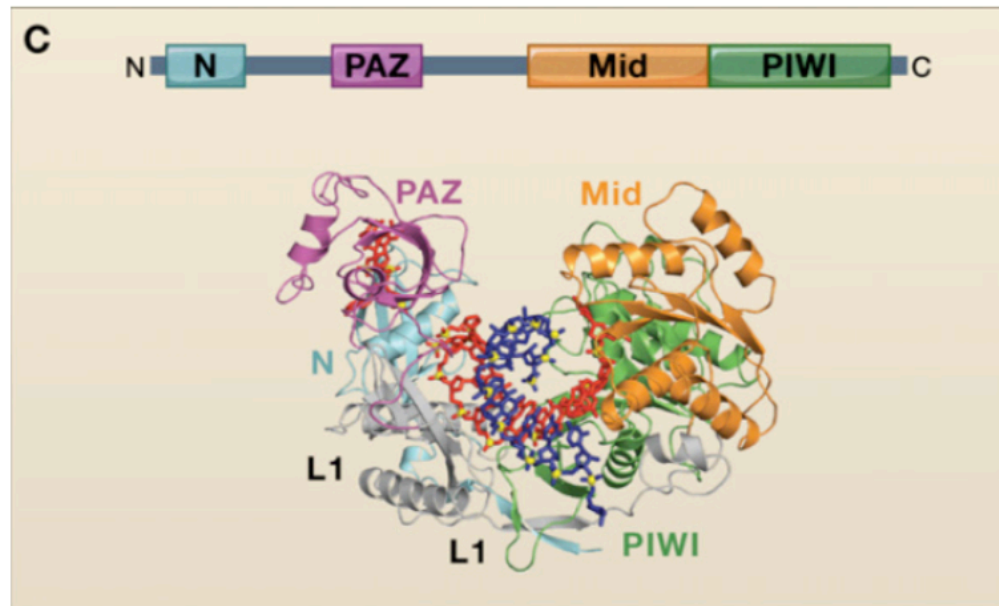
- Processing of dsRNAs into RISC requires accessory proteins: TRBP (R2D2 in *Drosophila*) forms complex with Dicer
- Other unknown proteins bind to form RISC Loading Complex
- Ago2 cleaves the passenger strand, leading to its ejection

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ARGONAUTE: Central component of RISC

- One strand of the dsRNA produced by Dicer is retained in the RISC complex in association with Argonaute
- Structure first solved by Leemor-Tor and colleagues (2004), more recent structures by Patel and colleagues include RNAs mimicking guide ssRNA and target mRNA
- The PAZ domain has RNA 3' end binding activity
- In structure without mRNA, guide strand nucleotides 2-6 have bases exposed and available for base-pairing
- PIWI domain adopts RNase H fold and in some Ago proteins can cleave the 'passenger strand' : i.e. the mRNA



Carthew and Sontheimer, Cell (2009) 136, 642-655.

siRNA: Exogenous dsRNA molecules

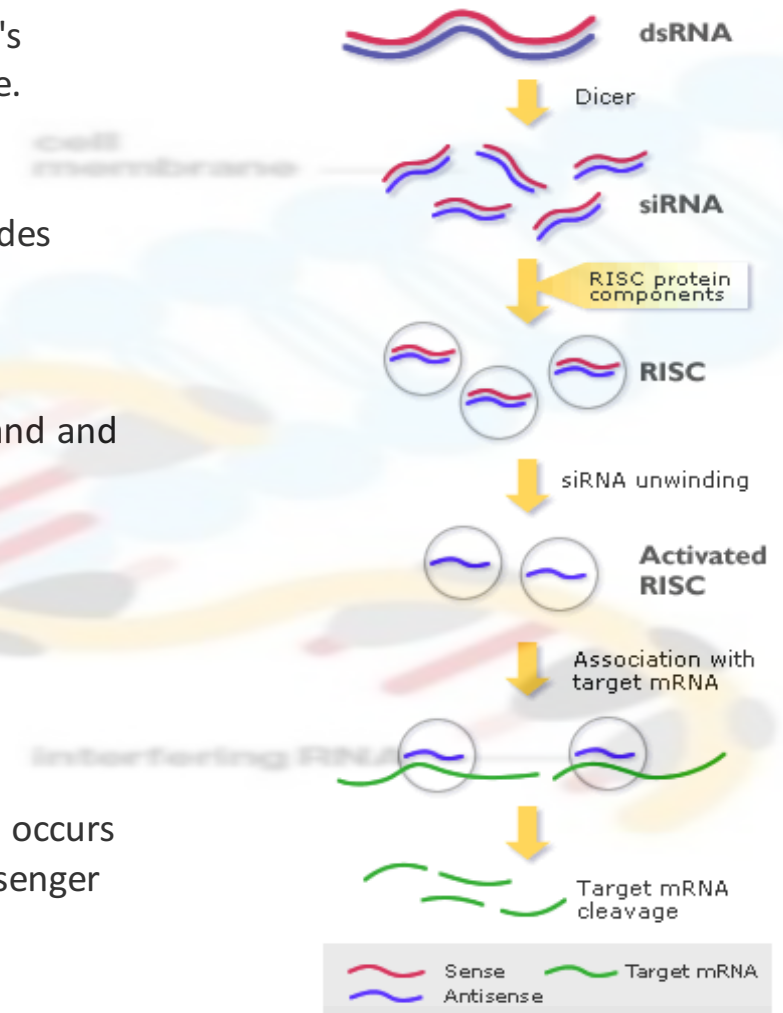
RNAi is controlled by **RISC** and is initiated by short dsRNA molecules in a cell's cytoplasm, where they interact with the catalytic RISC component argonaute.

dsRNAs is cleaved by the **Dicer enzyme** into short fragments of ~20 nucleotides that are called **siRNAs**.

Each siRNA is unwound into two single-stranded (ss) ssRNAs (**passenger** strand and the **guide** strand).

The passenger strand is degraded (red), and **the guide strand (blue) is incorporated into the RNA-induced silencing complex (RISC)**.

The most well-studied outcome is post-transcriptional gene silencing, which occurs when the guide strand base pairs with a complementary sequence in a messenger RNA molecule (green) and induces **cleavage by Argonaute**, the catalytic component of the RISC complex.



miRNA: Endogenous RNA silencing

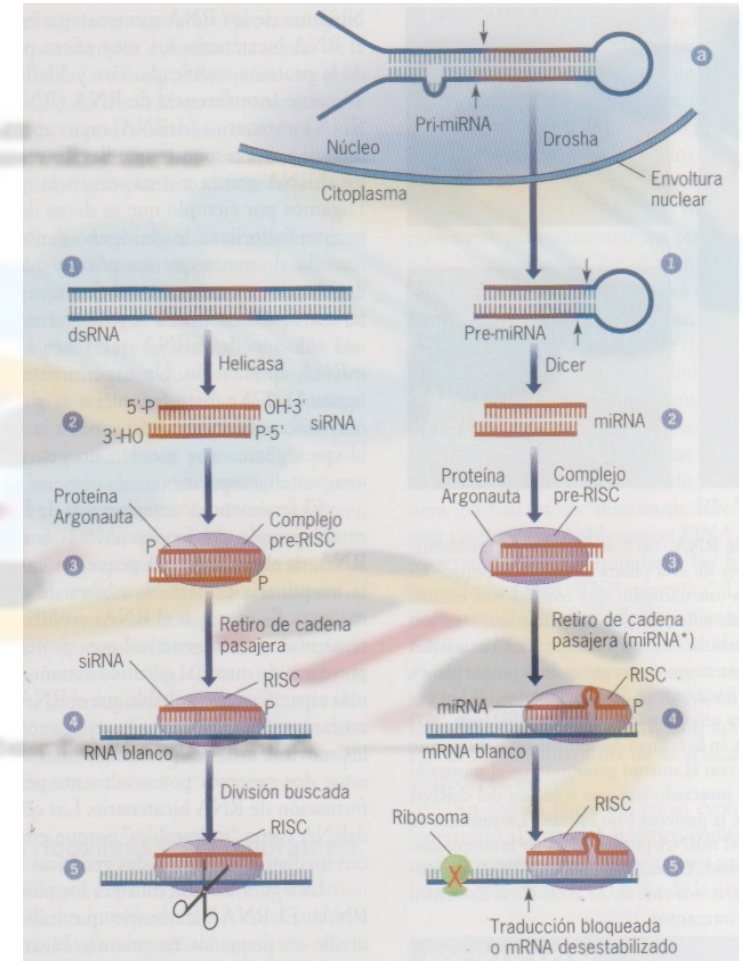
miRNAs are genomically encoded non-coding RNAs that regulate gene expression, particularly during development.

Mature miRNAs are structurally similar to siRNAs produced from exogenous dsRNA but **must undergo post-transcriptional modification**.

miRNA's are expressed from longer RNA-coding gene as a primary transcript (**pri-miRNA**) which is processed within the cell nucleus to a 70 bp stem-loop structure (**pre-miRNA**) by the microprocessor complex (RNase III **Drosha** and dsRNA binding protein DGCR8).

The dsRNA molecule is bound and cleaved by Dicer to produce the mature miRNA molecule that can be integrated into the RISC complex; thus, **miRNA and siRNA share the same cellular machinery downstream of their initial processing**.

miRNAs typically inhibit the translation of many different mRNAs with similar sequences. In contrast, siRNAs typically inhibit only a single, specific target.



dsRNA in the nucleus: silencing by formation of heterochromatin

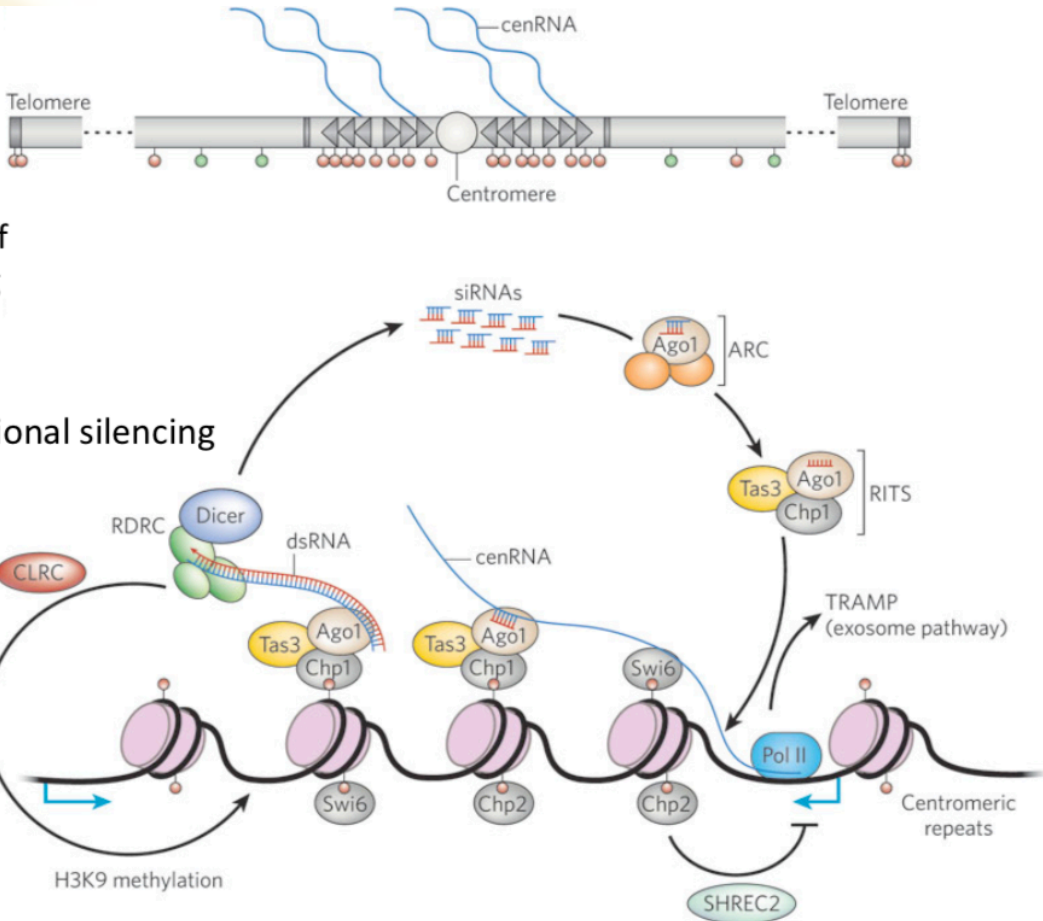
- Pathway best understood in *S. pombe*

- Silencing involves formation of heterochromatin and resulting transcriptional repression

the RNA-induced transcriptional silencing complex (RITS)

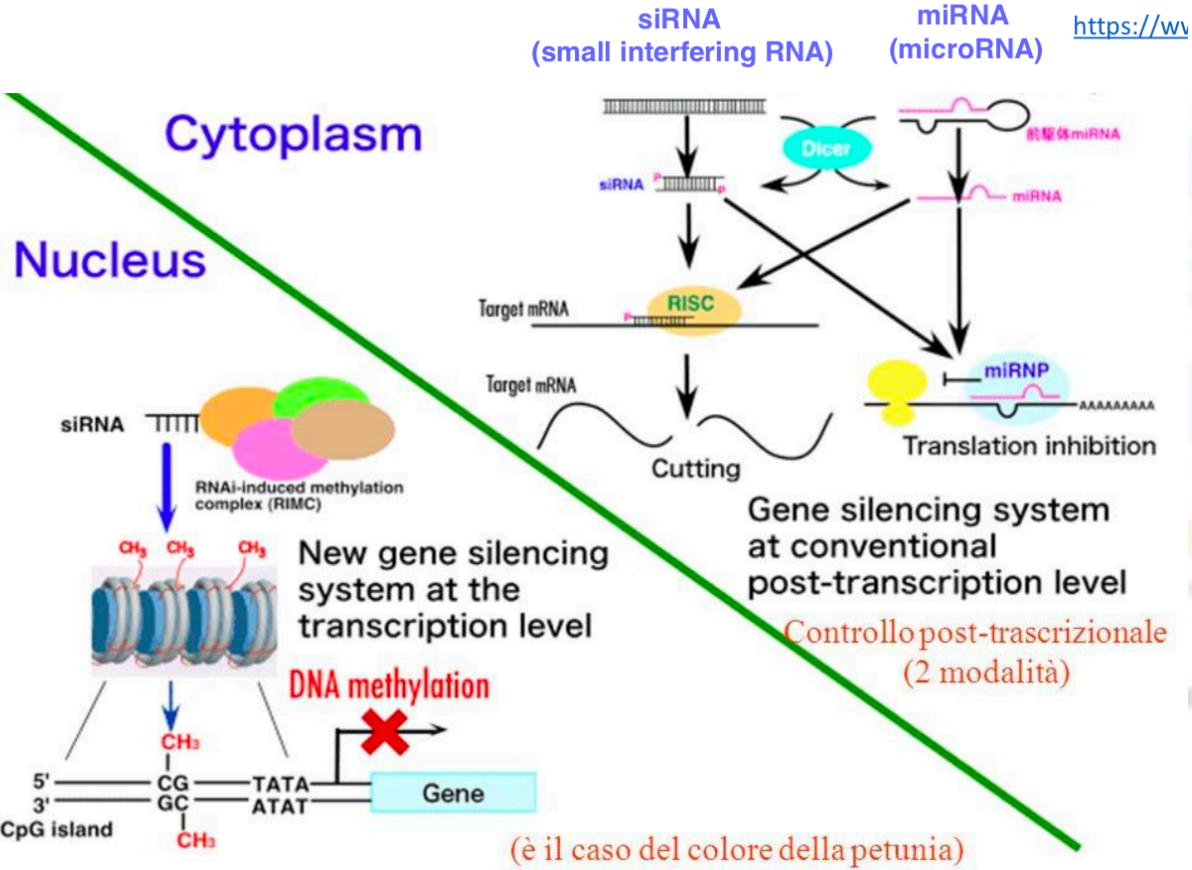
Clr4 methyltransferase complex (CLRC)

RNA-directed RNA polymerase complex (RDRC)



Moazed, *Nature* (2009) 457, 413-420

RNAi models



A fluorescence microscopy image showing several cells with a dense network of green-stained cytoskeletal filaments, likely microtubules or actin. The cells are spread out on a dark background, and the green signal highlights the intricate internal structure of the cytoskeleton.

RNAi
dalla teoria alla pratica di laboratorio

Come disegnare un siRNA in lab

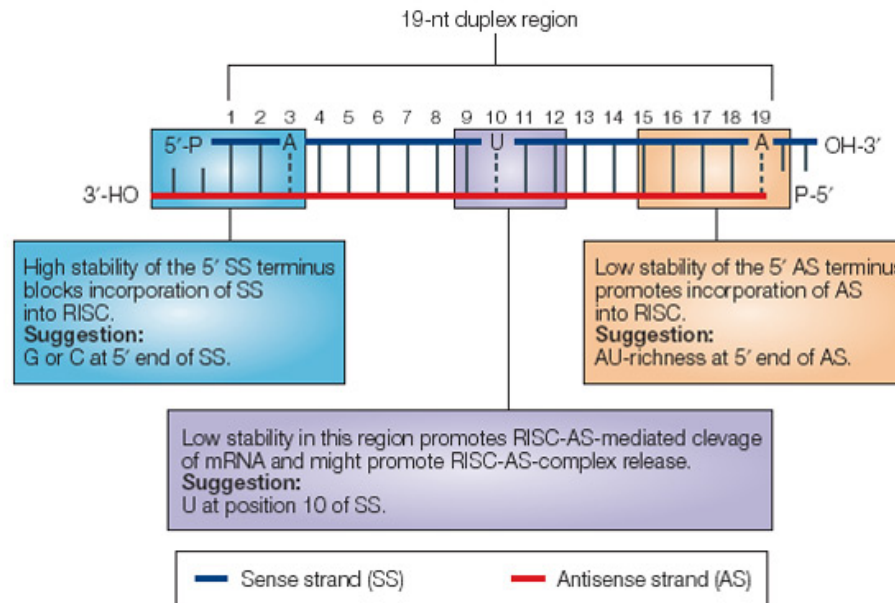
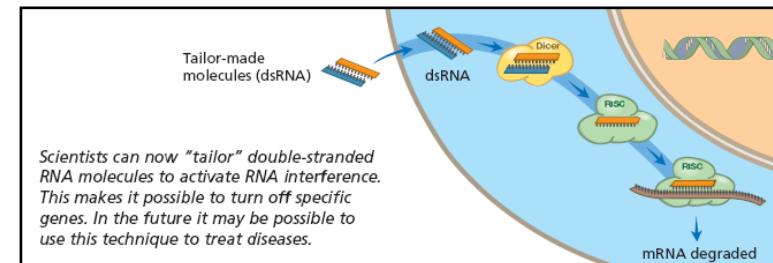
siRNA PROFILING TECHNOLOGIES

Disegno di siRNA

- il siRNA possiede un gruppo UU al terminale 3'
- 19 nucleotidi
- un contenuto in G/C < 50% è preferibile.

Algoritmo di nuova generazione

- Tm dell'siRNA
- Effetti della posizione nucleotidica
- Contenuto nucleotidico dei 3' overhangs
- Distribuzione nucleotidica
- Controllo della specificità



Come disegnare un siRNA in lab

Preventing Off-Target Effects

Overabundance of the siRNA activates the interferon pathway, as antiviral response



Low concentrations (~5-30nM) of single siRNA minimizes:

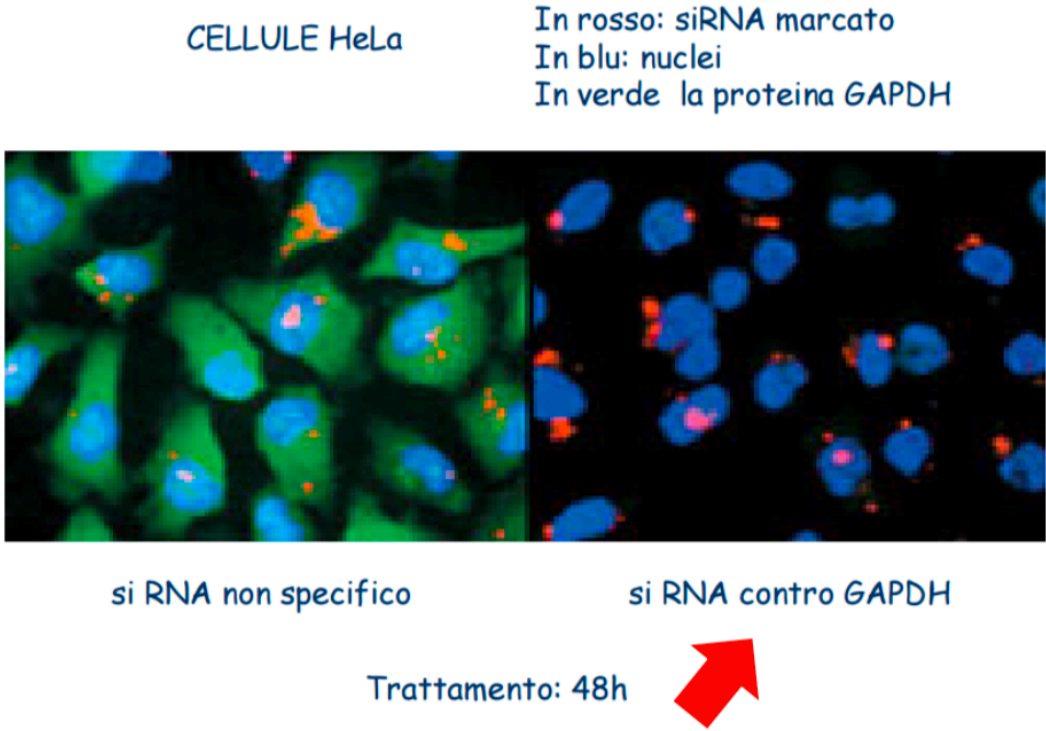
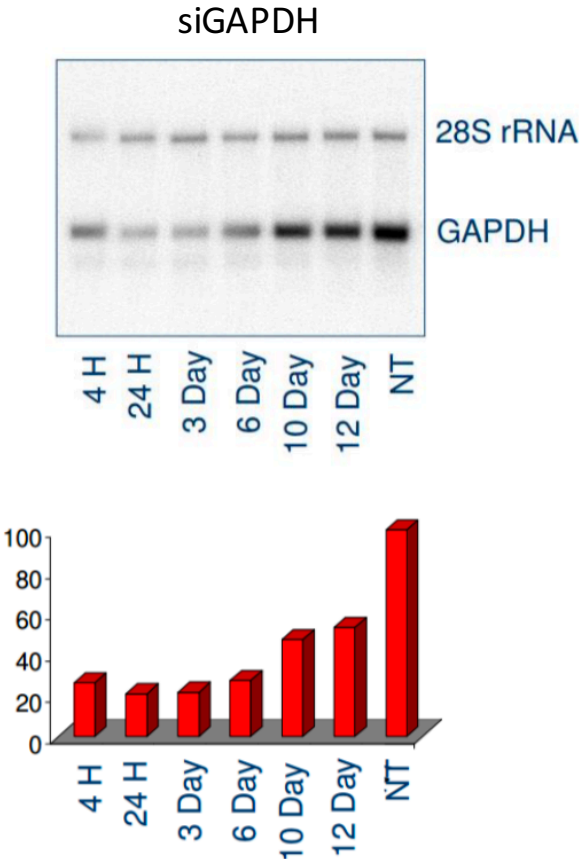
- chances of off-target effect
- induction of interferon response

It is currently preferable to use **ONE** highly potent siRNA than a **MIXTURE** of siRNAs that raise overall siRNA conc.



Verify specificity of RNAi effect by testing independent siRNAs to the same target

Durata del silenziamento transiente



Trasfezione con il siRNA: ottimizzazione delle condizioni

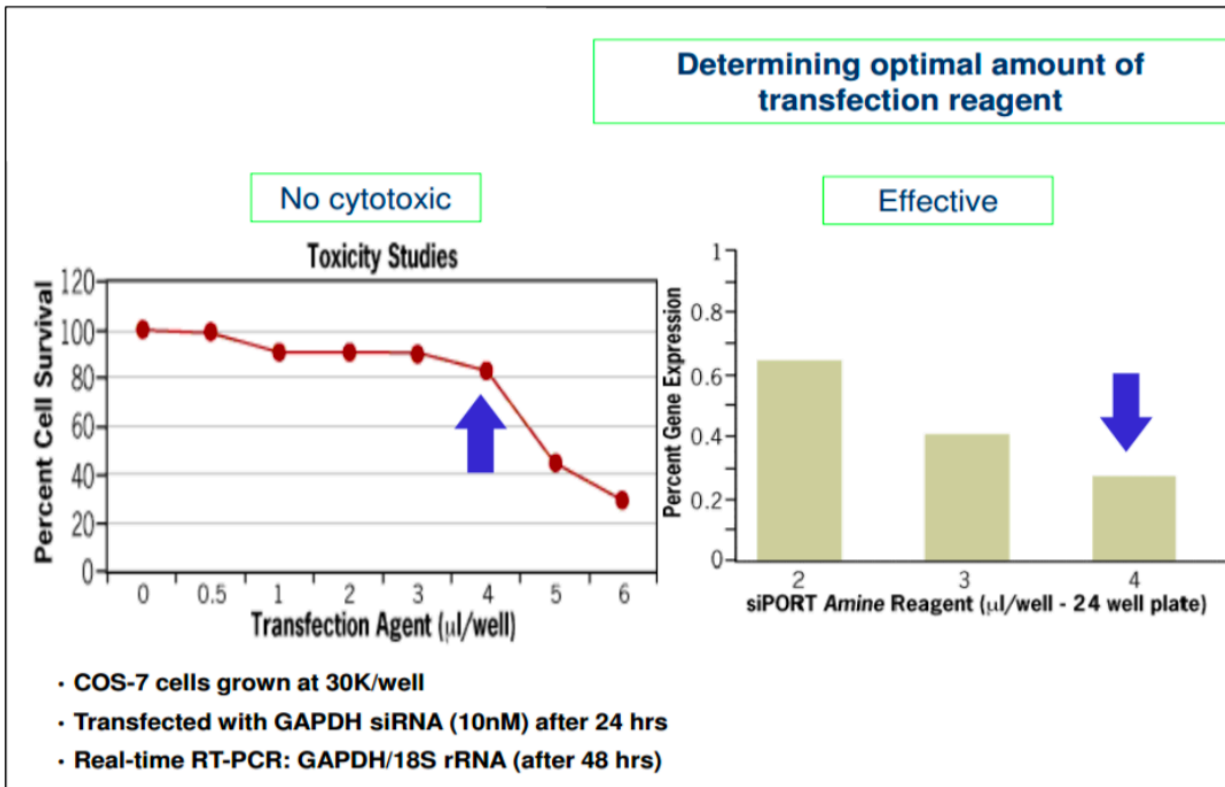


Prevenire effetti di spegnimento del target:

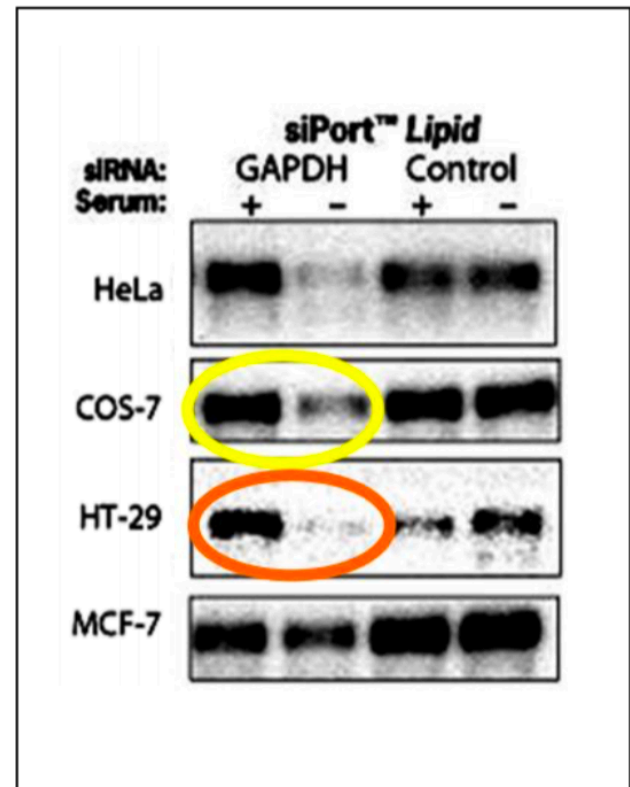
- Basse concentrazioni** (~5-30 nM) di siRNA per minimizzare l'attivazione l'**interferon pathway** come risposta anti-virale
- E' preferibile usare un solo siRNA molto efficiente piuttosto che una miscela di siRNA meno potenti, la MIXTURE fa aumentare la concentrazione totale
- Usare RNAi specifici, dopo aver effettuato test di siRNA differenti sullo stesso mRNA bersaglio

Trasfezione con il siRNA: ottimizzazione delle condizioni

Scelta del reagente trasfettante

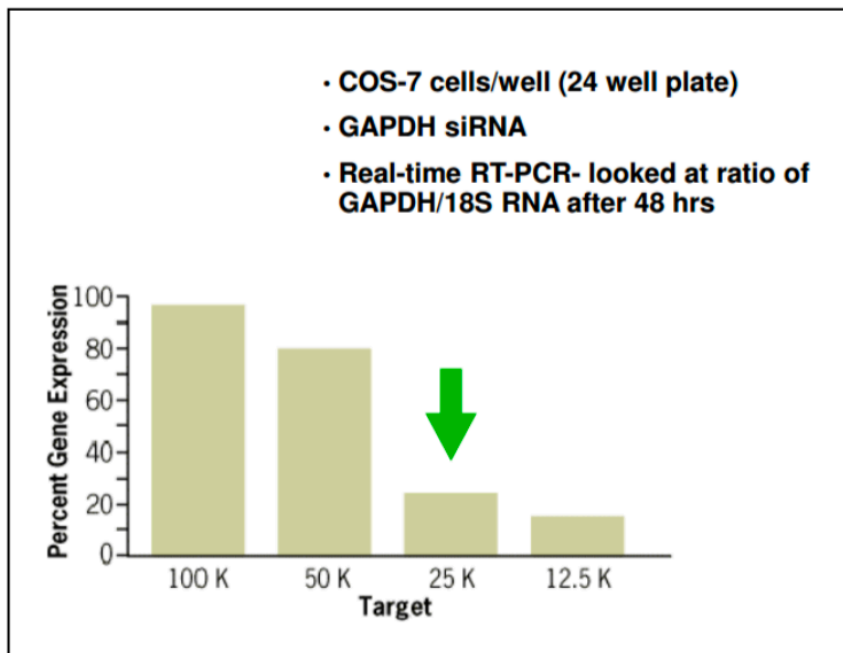


Presenza ed assenza di siero

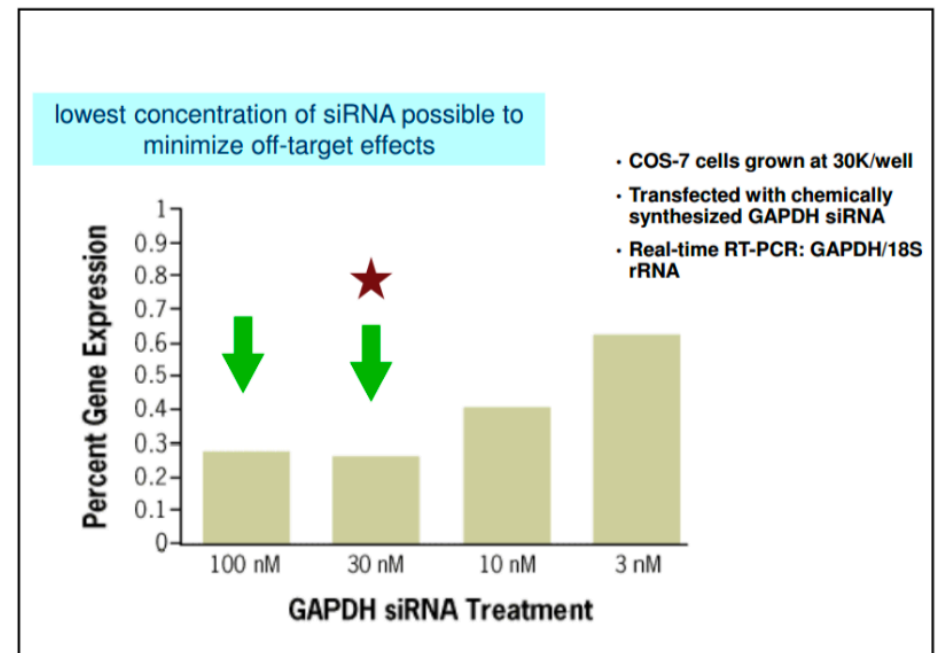


Trasfezione con il siRNA: ottimizzazione delle condizioni

Determinare la densità di semina delle cellule



Concentrazione di siRNA



Dal transiente alla trasfezione con vettori

PRO

- ❑ La trasfezione con siRNA è davvero molto efficiente in molti tipi di cellule
- ❑ Coi siRNA il silenziamento è immediato

CONTRO

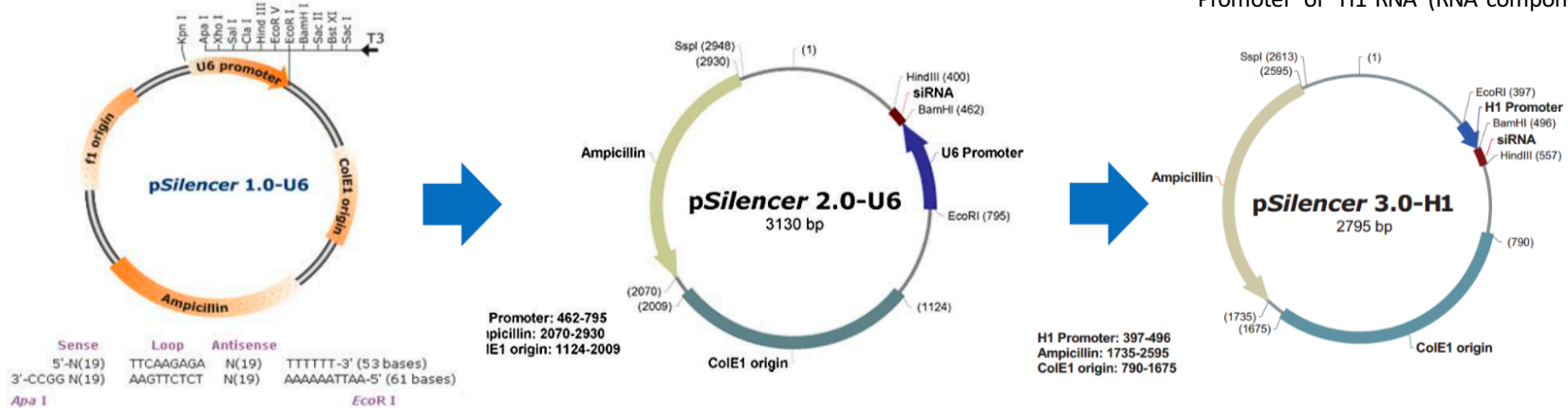
- ❑ Alcune cellule sono refrattarie alla trasfezione e la loro elettroporazione spesso causa morte cellulare
- ❑ I siRNA sono stabili, **ma la trascrizione può risultare transiente se le cellule si duplicano molto in fretta diluendo il silenziamento e la vita media della proteina**

Superamento del problema mediante.....

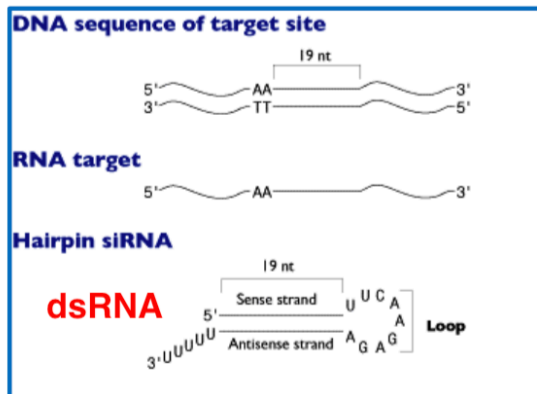
Dal transiente alla trasfezione con vettori

Silenziamento tramite vettori a DNA

Important:
 Use RNA Pol III promoters (short RNA)
 Promoter or U6 snRNA
 Promoter of H1 RNA (RNA component of human RNase P)



Mimicking
miRNA
production



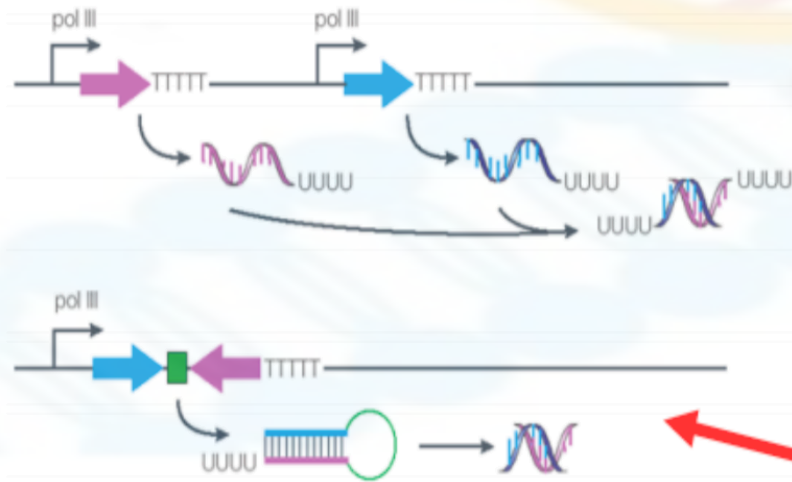
- Una sequenza stampo per un “hairpin siRNA” viene clonata in un opportuno vettore per trascrivere una molecola di RNA
- Produzione di siRNA *in vivo direttamente* nelle cellule trasfettate
- Trasfezione stabile nella linea cellulare di cui si vuol silenziare il gene target dell’RNAi
- Silenziamento a lungo termine del gene target**

Dal transiente alla trasfezione con vettori

Sintesi di siRNA *in vivo*

❏ Nessuna sequenza richiesta dopo start site per la trascrizione

❏ TTTT: sufficiente per terminazione



❏ **Clonati in vettori plasmidici**
con promotori adatti per la
produzione di RNA

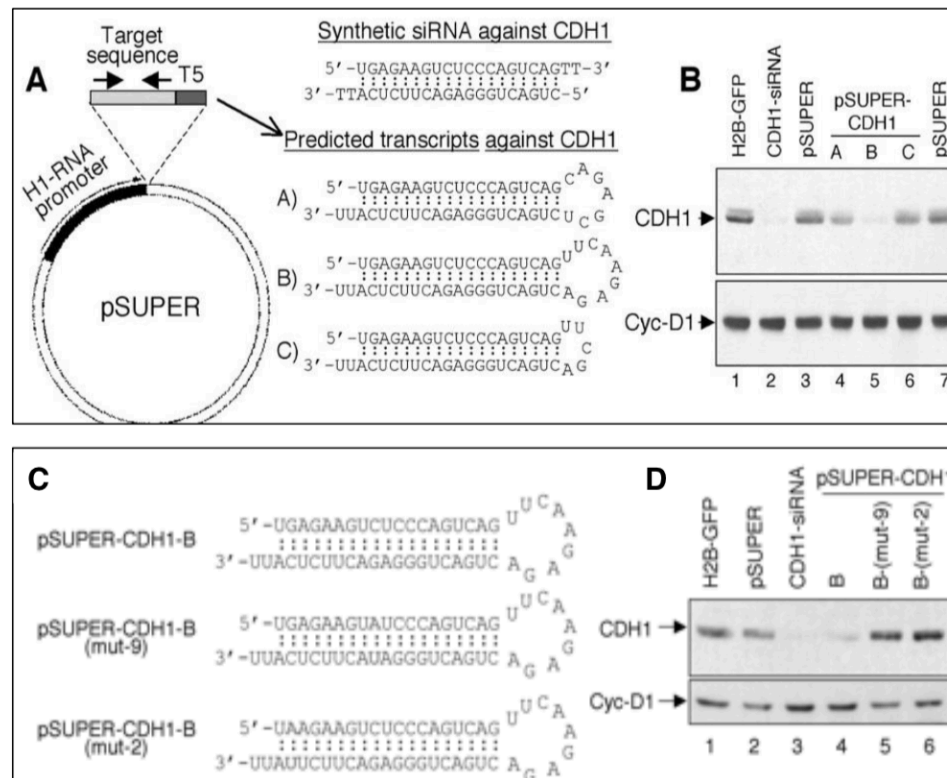
❏ **Clonati in vettori virali**

- **Oncoretrovirus:** MoMuLV o MSCV, le cellule devono duplicanti per poter essere infettate
- **Lentivirus:** HIV-1, per infettare cellule quiescenti

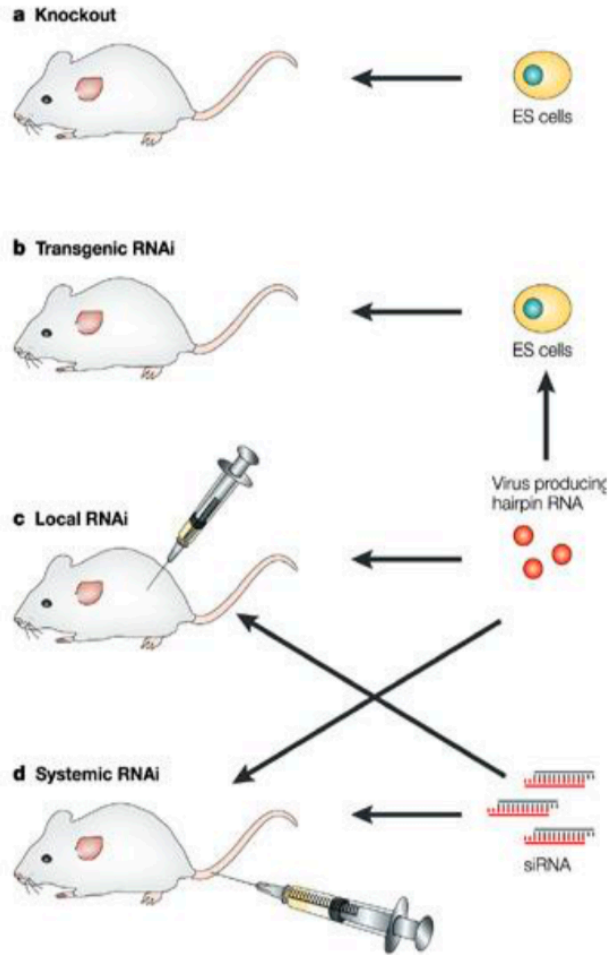
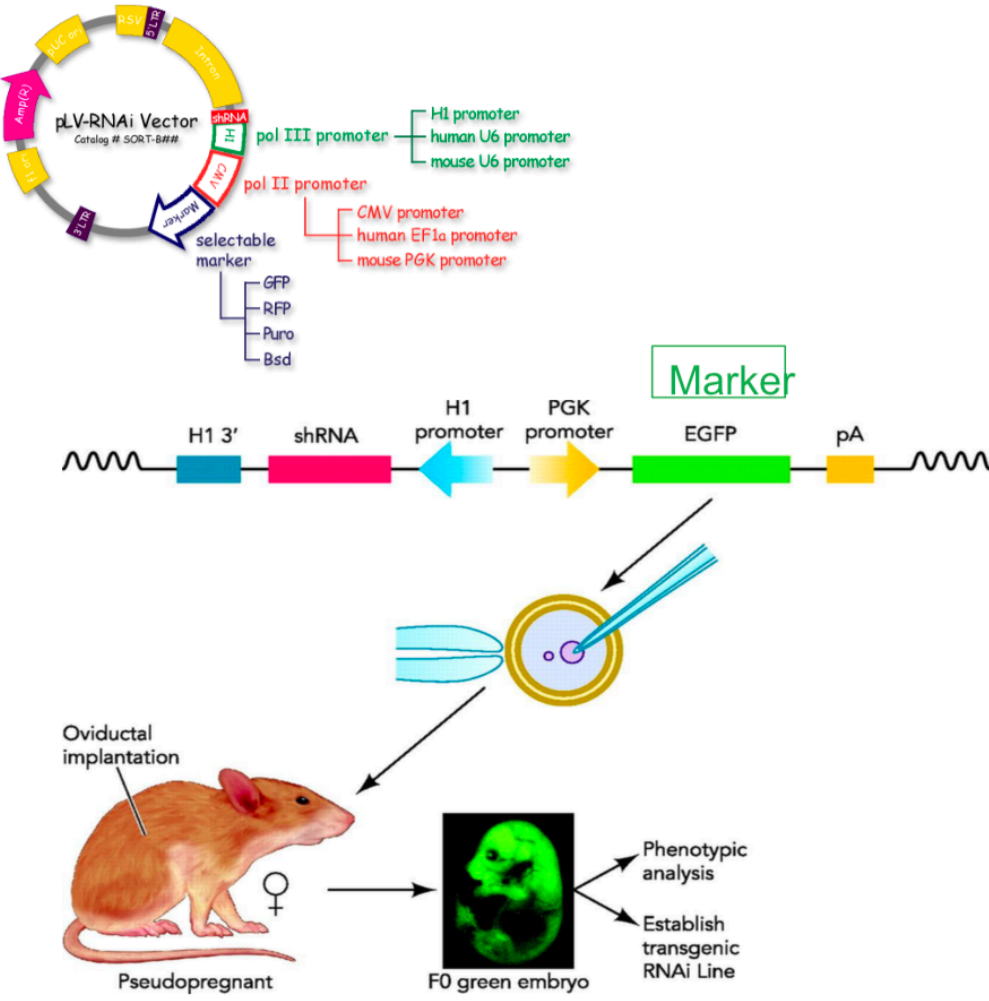
More efficiently processed by DICER!!

Espressione stabile di shRNA

Un sistema per l'espressione stabile di *short interfering RNA* in cellule di mammifero: vettore plasmidico



Espressione siRNA in vivo



Trasfezione con siRNA: le APPLICAZIONI

- Silenziamento genico specifico, efficiente e stabile nel tempo (economico e veloce)
- È un approccio di «genetica inversa»
- Screening delle funzioni genomiche (Genome-wide functional screenings)
- Terapia genica (es. antitumorale)
- Creazione di modelli per lo studio di agenti farmacologici (es. murini)
- Rivoluzione nello studio dei meccanismi di regolazione dell'espressione genica

siRNA library design

1. Grazie ai siRNA è possibile **silenziare uno alla volta** tutti i geni di un organismo.
2. Una tipica applicazione consiste **nell'identificare quali geni sono coinvolti in un certo processo**
3. Il punto di partenza è una **libreria di siRNA, specifica per un singolo gene del genoma**. Oggi esistono librerie in grado di coprire la maggior parte dei geni umani (≈ 20.000 siRNA).

siRNA "potenzialmente" funzionale:

- La regione target deve essere a valle del codone di inizio, ad una distanza che varia da 50 a 100bp.
- Lunghezza compresa fra 19-22 bp.
- Contenuto in GC fra il 35-55%
- 2-nt 3' overhangs di residui di uridina
- 5'-phosphate and 3'-hydroxyl group.

- **Stability**
- **Access to RISC**

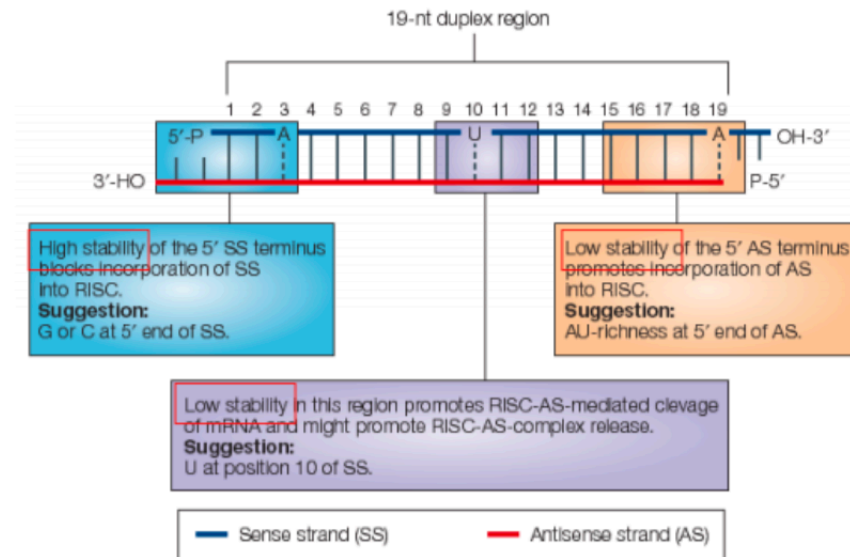


Figure 2 | **The generation of effective siRNA.** A small interfering RNA (siRNA) is a 21–23-nucleotide (nt) dsRNA that contains: a 19-nt duplexed region, symmetric 2–3-nt 3' overhangs, and 5'-phosphate (P) and 3'-hydroxyl (OH) groups. The positions of each nucleotide in the 19-nt duplexed region of the sense strand are shown. On the basis of recently established design criteria, an effective siRNA has high stability at the 5' terminus of the sense strand (blue box), lower stability at the 5' antisense terminus (orange box) and at the cleavage site (purple box). In addition, the sequence-specific preferences at the following positions on the sense strand are important: the presence of an A at position 19, an A at position 3, a U at position 10 (BOX 2 lists other parameters). RISC, RNA-induced silencing complex.

Mittal, Nature Review Genet, 2004

Limitazioni dei siRNA

Impossibile studiare geni essenziali
per la sopravvivenza cellulare (*housekeeping*) e sviluppo



Sviluppo di nuovi vettori
per l'espressione condizionale-inducibile dei shRNA

(tet OFF/ON H1 and U6 promoter system)

siRNA vs. oligonucleotidi antisense (a ssDNA)

Similarità

- Lunghezza
- Metodologia di *delivery* comune
- Induzione di silenziamento genico a livello post-trascrizionale
- Digestione di mRNA bersaglio da parte di endonucleasi
- Possibilità di stabilizzare con basi modificate
- Bio-distribuzione simile

Differenze

- Doppio filamento vs. singolo filamento
- Maggiore stabilità del siRNA
- Maggiore efficacia delle molecole in cellule in coltura
- Meccanismo d'azione mediato da RISC

