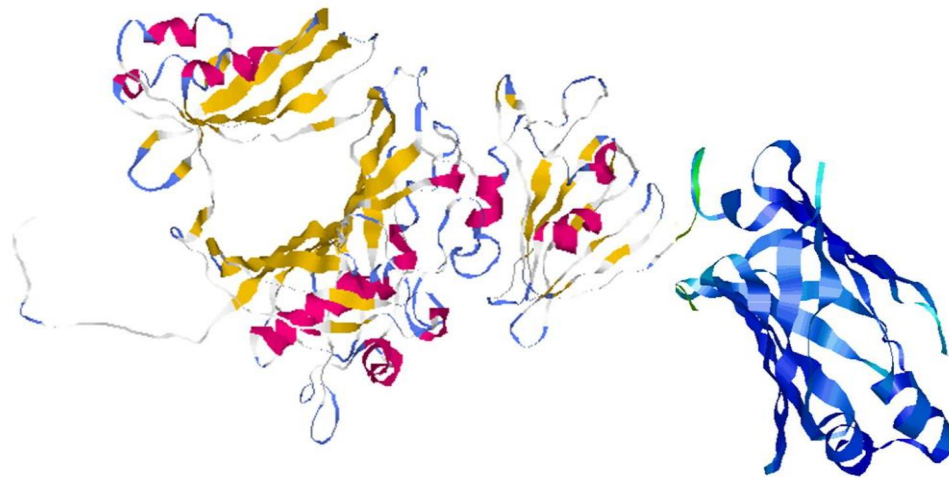


# **Solution Properties of the new Fusion Protein HUG**



**CASO STUDIO 2:**  
Ricerche svolte dal 2010 ad oggi

**HELP** (*Human Elastin-like Polypeptide*), a synthetic polypeptide based on the **VAPGVG** hexapeptidic motif that is found in the human elastin repetition domains, shows very interesting chemical-physical properties in solution and in particular it has the peculiar thermal behavior defined as the **reverse phase transition**:

- at temperatures below the transition temperature ( $T_t$ ), the biopolymer is soluble in aqueous solutions where the free chains of HELP exist in a disordered and completely hydrated state.
- at temperatures above  $T_t$ , these chains show a more orderly structure ( $\beta$ -spiral) stabilized by intramolecular hydrophobic interactions that favor their association and the formation of an amorphous solid phase.

# HELP

## HELP

MW 44885.7 536 aa, Theoretical pI: 11.68

MRGSHHHHHHGSAAAAAAAAAKAAKAAQFGLVPGVGVAPGVGVAPGVGVAPGVGLAPGVGVAPGV  
GVAPGVGVAPGIAPAAAAAAKAAKAAQFGLVPGVGVAPGVGVAPGVGVAPGVGLAPGVGVAPGV  
GVAPGVGVAPGIAPAAAAAAKAAKAAQFGLVPGVGVAPGVGVAPGVGVAPGVGLAPGVGVAPGV  
GVAPGVGVAPGIAPAAAAAAKAAKAAQFGLVPGVGVAPGVGVAPGVGVAPGVGLAPGVGVAPGV  
GVAPGVGVAPGIAPAAAAAAKAAKAAQFGLVPGVGVAPGVGVAPGVGVAPGVGLAPGVGVAPGV  
GVAPGVGVAPGIAPAAAAAAKAAKAAQFGLVPGVGVAPGVGVAPGVGVAPGVGLAPGVGVAPGV  
GVAPGVGVAPGIAPAAAAAAKAAKAAQFGLVPGVGVAPGVGVAPGVGVAPGVGLAPGVGVAPGV  
GVAPGVGVAPGIAPGV

Alpha helix	(Hh)	:	137	is	25.56%
3 <sub>10</sub> helix	(Gg)	:	0	is	0.00%
Pi helix	(Ii)	:	0	is	0.00%
Beta bridge	(Bb)	:	0	is	0.00%
Extended strand	(Ee)	:	0	is	0.00%
Beta turn	(Tt)	:	0	is	0.00%
Bend region	(Ss)	:	0	is	0.00%
Random coil	(Cc)	:	399	is	74.44%
Ambiguous states (?)		:	0	is	0.00%
Other states		:	0	is	0.00%

**Total number of negatively charged residues (Asp + Glu): 0**

**Total number of positively charged residues (Arg + Lys): 17**

# HELP

## Target Sequence:

10 20 30 40 50 60 70  
MRGSHHHHHH GSAAAAAAA KAAAKAAQFG LVPGVGVAPG VGVAPGVGVÄ PGVGLAPGVG VAPGVGVAPG

80 90 100 110 120 130 140  
VGVAPGIAPÄ AAAAAKAAAK AAQFGLVPGV GVAPGVGVAP GVG VAPGVGL APGVGVAPGV VAPGVGVAP

150 160 170 180 190 200 210  
GIAPAAAAAA KAAAKAAQFG LVPGVGVAPG VGVAPGVGVÄ PGVGLAPGVG VAPGVGVAPG VGVAPGIAPÄ

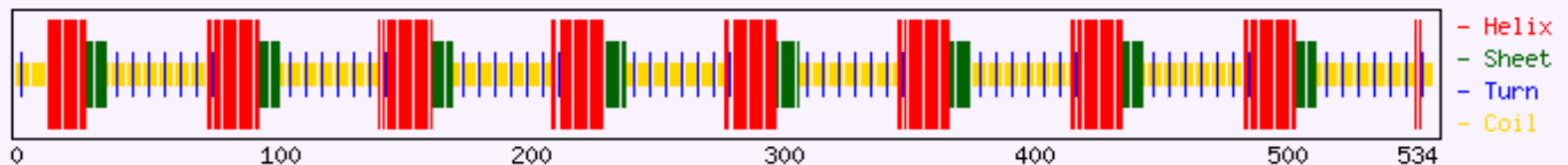
220 230 240 250 260 270 280  
AAAAAKAAAK AAQFGLVPGV GVAPGVGVAP GVG VAPGVGL APGVGVAPGV GVAPGVGVAP GIAPAAAAAA

290 300 310 320 330 340 350  
KAAAKAAQFG LVPGVGVAPG VGVAPGVGVÄ PGVGLAPGVG VAPGVGVAPG VGVAPGIAPÄ AAAAAKAAAK

360 370 380 390 400 410 420  
AAQFGLVPGV GVAPGVGVAP GVG VAPGVGL APGVGVAPGV GVAPGVGVAP GIAPAAAAAA KAAAKAAQFG

430 440 450 460 470 480 490  
LVPGVGVAPG VGVAPGVGVÄ PGVGLAPGVG VAPGVGVAPG VGVAPGIAPÄ AAAAAKAAAK AAQFGLVPGV

500 510 520 530  
GVAPGVGVAP GVG VAPGVGL APGVGVAPGV GVAPGVGVAP GIAP



Secondary Structure

In **2013**, a new protein, called **UnaG**, was identified and isolated from the muscle of the Japanese eel (*Anguilla japonica*) (N:B: è la prima proteina fluorescente derivata dai vertebrati).

This free-fatty acid binding protein binds unconjugated bilirubin (Br) in a highly specific and selective way, emitting strong fluorescence (Kumagai et al., 2013).

The UnaG gene was been cloned to the C-terminal of the gene encoding the HELP polypeptide obtaining a new functionalized synthetic polypeptide called **HUG**, an acronym that indicates **HELP-UnaG** sequence.

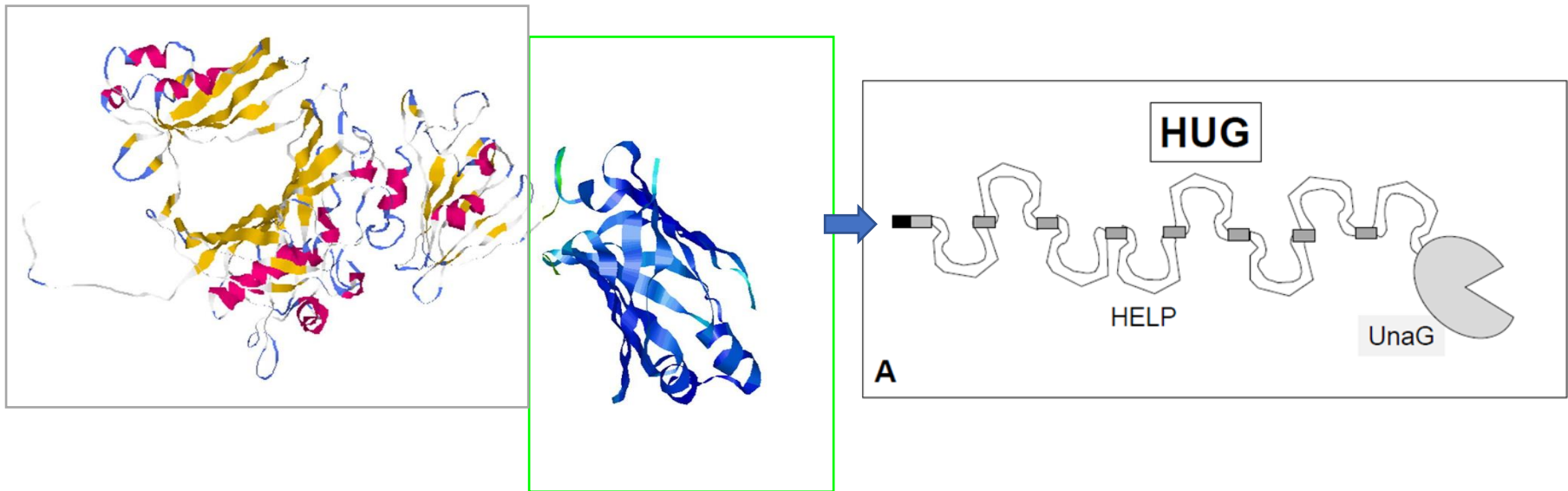
**HUG (HELP-UnaG)** was been produced from a synthetic gene of the HELP polypeptide fused with the **139** amino acids coding sequence of the UnaG bilirubin-binding protein

Its molecular property appears highly interesting because unconjugated bilirubin (indirect bilirubin) is hardly measured with traditional colorimetric assay.

# HUG

A bi-functional, synthetic protein

HUG is composed of a bilirubin-binding domain (UnaG) fused with a scaffold (HELP)



HUG is the acronym of **HELP-UnaG**

# HUG

**HELP-UnaG** (clon. /10/2014) Number of amino acids: 675, Molecular weight: 60406.47, Theoretical pI: 9.88

MRGSHHHHHHGSAAAAAAAAAKAAKAAQFGLVPGVGVAPGVGVAPGVGVAPGVGLAPGVGVAPGVGVAP  
GVGVAPGIAPAAAAAAKAAKAAQFGLVPGVGVAPGVGVAPGVGVAPGVGLAPGVGVAPGVGVAPGVGV  
APGIAPAAAAAAKAAKAAQFGLVPGVGVAPGVGVAPGVGVAPGVGLAPGVGVAPGVGVAPGVGVAPGI  
APAAAAAAKAAKAAQFGLVPGVGVAPGVGVAPGVGVAPGVGLAPGVGVAPGVGVAPGVGVAPGIAPAA  
AAAAKAAKAAQFGLVPGVGVAPGVGVAPGVGVAPGVGLAPGVGVAPGVGVAPGVGVAPGIAPAAAAAA  
**KAAKAAQFGLVPGVGVAPGVGVAPGVGVAPGVGLAPGVGVAPGVGVAPGVGVAPGIAPAAAAAAKAAA**  
**KAAQFGLVPGVGVAPGVGVAPGVGVAPGVGLAPGVGVAPGVGVAPGVGVAPGIAPAAAAAAKAAKAAQ**  
**FGLVPGVGVAPGVGVAPGVGVAPGVGLAPGVGVAPGVGVAPGVGVAPGIAPAAAAAAKAAKAAQ**  
**FGLVPGVGVAPGVGVAPGVGVAPGVGLAPGVGVAPGVGVAPGVGVAPGIAP**  
**GGMVEKFGVTWKIADSHNFGEYLKAIGAPKELSDGGDATTPTLYISQKDGDKMTVKIENGPPTFLDTQV**  
**KFKLGEEFDEFPSDRRKGVKSVVNLVGEKLVYVQKWDGKETTYVREIKDGKLVVTLTMGDVVAVRSYRR**  
**ATE**

Alpha helix	(Hh)	:	198	is	22.84%
$3_{10}$ helix	(Gg)	:	0	is	0.00%
Pi helix	(Ii)	:	0	is	0.00%
Beta bridge	(Bb)	:	0	is	0.00%
Extended strand	(Ee)	:	45	is	5.19%
Beta turn	(Tt)	:	0	is	0.00%
Bend region	(Ss)	:	0	is	0.00%
Random coil	(Cc)	:	624	is	71.97%
Ambiguous states (?)		:	0	is	0.00%
Other states		:	0	is	0.00%

Total number of negatively charged residues  
(Asp + Glu): 22

Total number of positively charged residues  
(Arg + Lys): 39

# HUG

```
      10      20      30      40      50      60      70
MRGSHHHHHH GSAAAAAAA KAAAKAAQFG LVPGVGVAPG VGVAPGVGVÄ PGVGLAPGVG VAPGVGVAPG

      80      90     100     110     120     130     140
VGVAPGIAPÄ AAAAAKAAAK AAQFGLVPGV GVAPGVGVAP GVGVAPGVGL APGVGVAPGV VAPGVGVAPG

     150     160     170     180     190     200     210
GIAPAAAAAA KAAAKAAQFG LVPGVGVAPG VGVAPGVGVÄ PGVGLAPGVG VAPGVGVAPG VGVAPGIAPÄ

     220     230     240     250     260     270     280
AAAAAKAAAK AAQFGLVPGV GVAPGVGVAP GVGVAPGVGL APGVGVAPGV GVAPGVGVAP GIAPAAAAAA

     290     300     310     320     330     340     350
KAAAKAAQFG LVPGVGVAPG VGVAPGVGVÄ PGVGLAPGVG VAPGVGVAPG VGVAPGIAPÄ AAAAAKAAAK

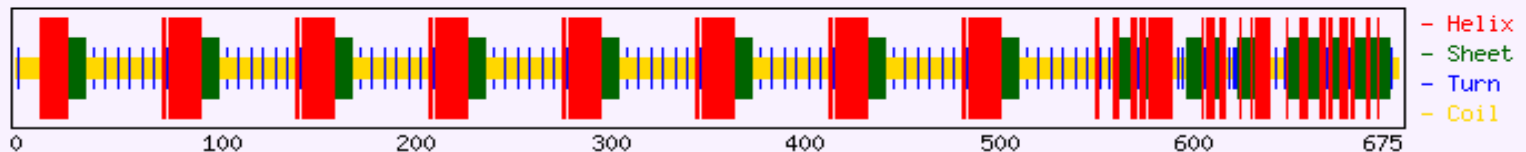
     360     370     380     390     400     410     420
AAQFGLVPGV GVAPGVGVAP GVGVAPGVGL APGVGVAPGV GVAPGVGVAP GIAPAAAAAA KAAAKAAQFG

     430     440     450     460     470     480     490
LVPGVGVAPG VGVAPGVGVÄ PGVGLAPGVG VAPGVGVAPG VGVAPGIAPÄ AAAAAKAAAK AAQFGLVPGV

     500     510     520     530     540     550     560
GVAPGVGVAP GVGVAPGVGL APGVGVAPGV GVAPGVGVAP GIAPGGMVEK FVGTWKIADS HNFGEYLKAI

     570     580     590     600     610     620     630
GAPKELSDGG DATTPTLYIS QKDGDKMTVK IENGPPTFLD TQVKFKLGEE FDEFPSDRRK GVKSVVNLVG

     640     650     660     670
EKLIVYQKWD GKETTYVREI KDGKLVVTLT MGDVVAVRSY RRATE
```





```

10      20      30      40      50      60      70
MRGSHHHHHH GSAAAAAAA KAAAKAAQFG LVPGVGVAPG VGVAPGVGVÄ PGVGLAPGVG VAPGVGVAPG

80      90      100     110     120     130     140
VGVAPGIAPÄ AAAAAKAAAK AAQFGLVPGV GVAPGVGVAP GVGVAPGVGL APGVGVAPGV VAPGVGVAP

150     160     170     180     190     200     210
GIAPAAAAAA KAAAKAAQFG LVPGVGVAPG VGVAPGVGVÄ PGVGLAPGVG VAPGVGVAPG VGVAPGIAPÄ

220     230     240     250     260     270     280
AAAAAKAAAK AAQFGLVPGV GVAPGVGVAP GVGVAPGVGL APGVGVAPGV VAPGVGVAPG GIAPAAAAAA

290     300     310     320     330     340     350
KAAAKAAQFG LVPGVGVAPG VGVAPGVGVÄ PGVGLAPGVG VAPGVGVAPG VGVAPGIAPÄ AAAAAKAAAK

360     370     380     390     400     410     420
AAQFGLVPGV GVAPGVGVAP GVGVAPGVGL APGVGVAPGV VAPGVGVAPG GIAPAAAAAA KAAAKAAQFG

430     440     450     460     470     480     490
LVPGVGVAPG VGVAPGVGVÄ PGVGLAPGVG VAPGVGVAPG VGVAPGIAPÄ AAAAAKAAAK AAQFGLVPGV

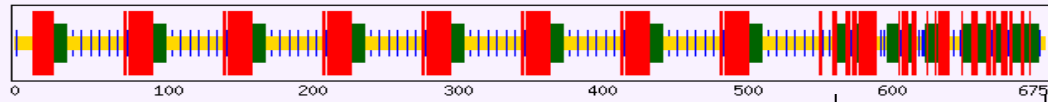
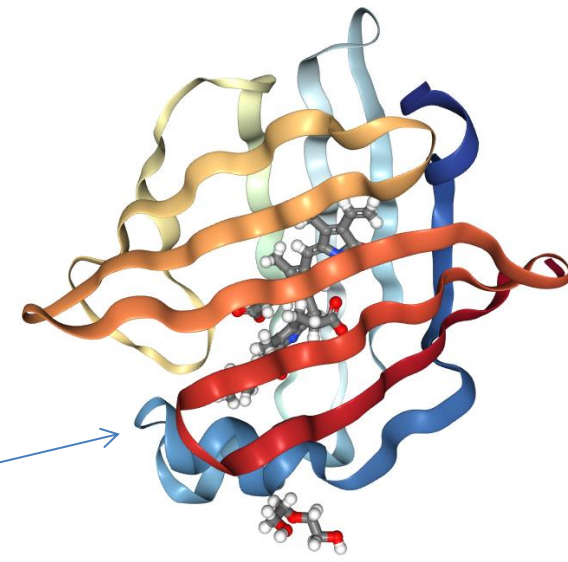
500     510     520     530     540     550     560
GVAPGVGVAP GVGVAPGVGL APGVGVAPGV GVAPGVGVAP GIAPGGMVEK FVGWTKIADS HNFGEYLKAI

570     580     590     600     610     620     630
GAPKELSDGG DATPTPLYIS QKDGDKMIVK IENGPTFLD TQVKFKLGEÄ FDEFPSDRRK GVKSVVNLVÄ

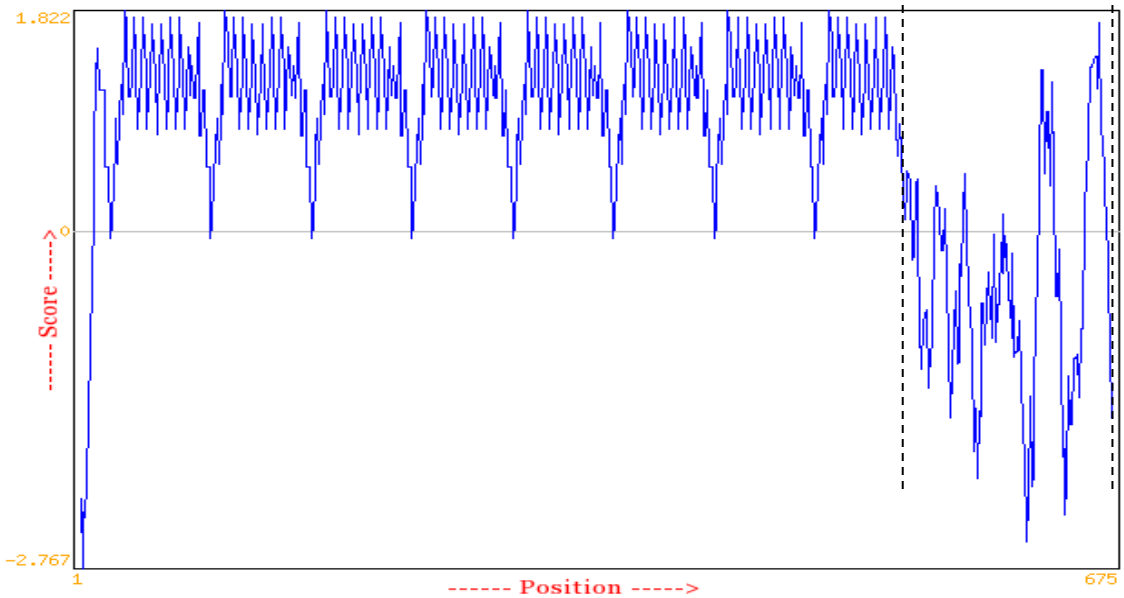
640     650     660     670
EKLVYVQKWD GKETTYVREI KDGKLVVILT MGDVVAVRSY RRATE

```

**UNAG**



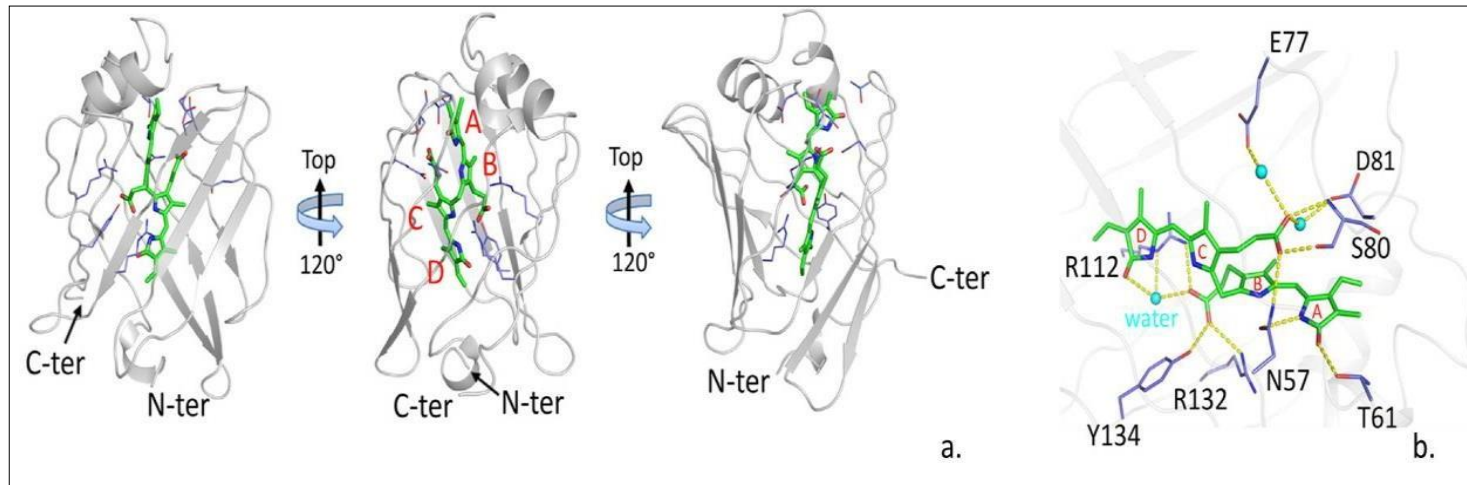
Kyte & Doolittle Hydrophobicity Plot



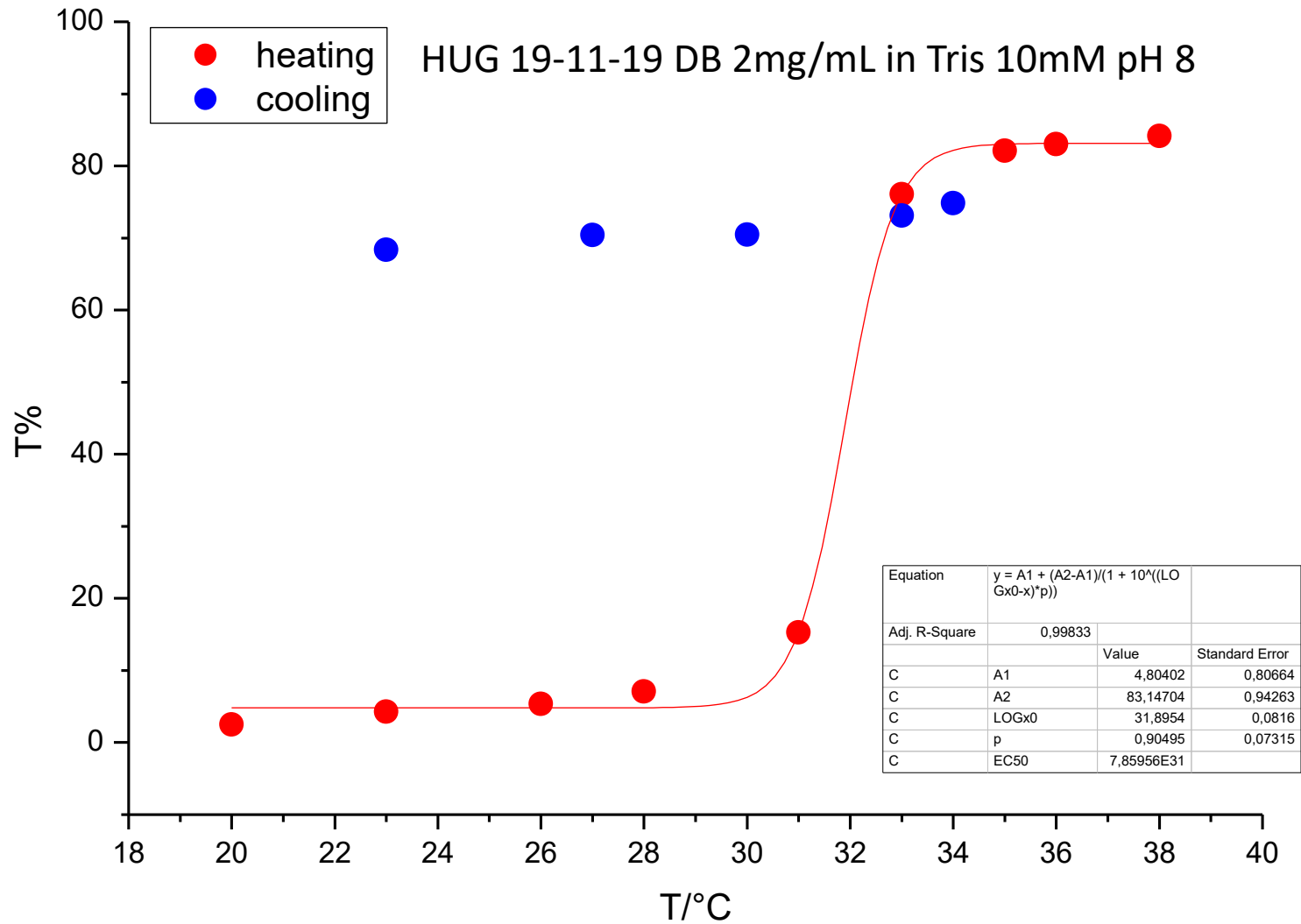
The **biosensor HUG** for the quantitative analysis of unconjugated bilirubin was developed in our laboratory (Dr. Antonella Bandiera and co-workers):

This new fusion protein preserves:

- 1) the reverse phase transition property of HELP
- 2) the highly specific capacity of UnaG to bind bilirubin and to become fluorescent.



# The reverse thermal transition of HUG



## **CHARACTERIZATION OF HUG**

- **Molecular mechanics and dynamic simulation**
- **Circular Dichroism (CD)**
- **Differential Scanning Calorimetry (DSC)**
- **Potentiometric titration of HUG and HELP**

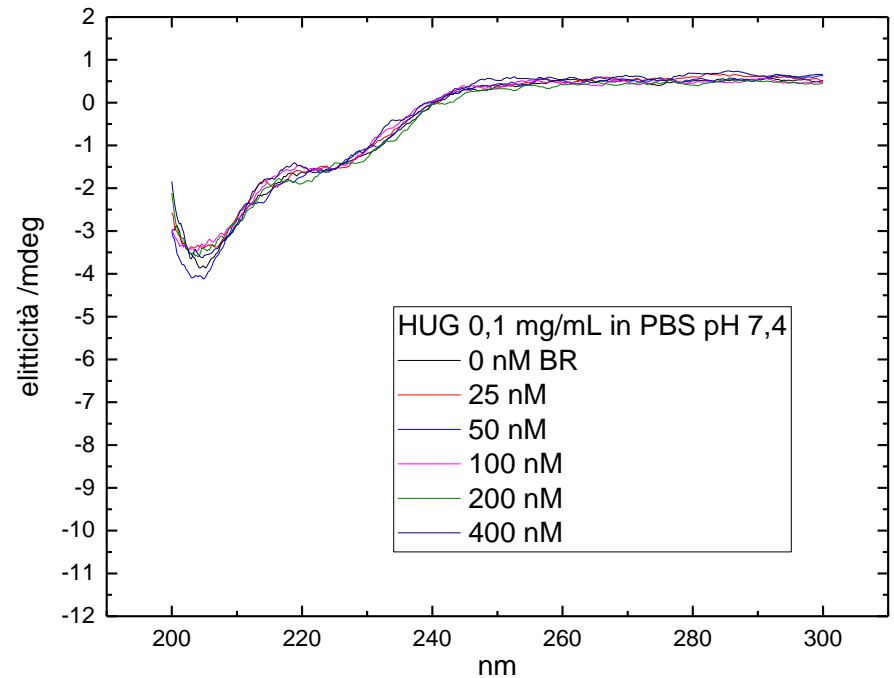
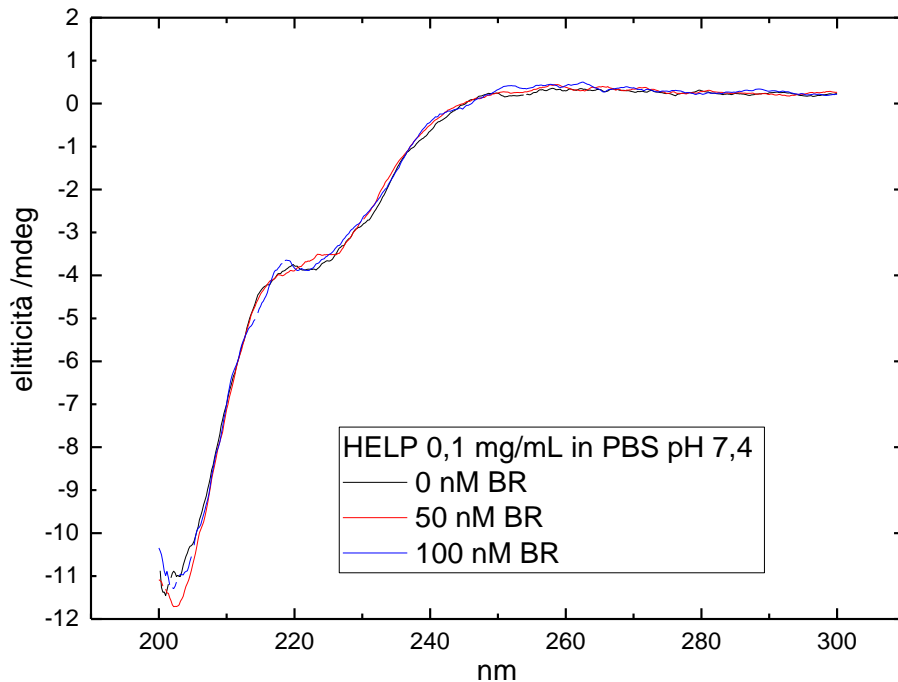
The physical and chemical characterization of the new fluorescent biosensor HUG. This step is very important for optimization of liquid assay of free bilirubin in biological samples

Analyzing quantitatively **the ligand binding to macromolecule** it is possible to focus on the application and importance of HUG biopolymer for bilirubin quantitative assessment in biological systems

**Fluorescence spectroscopy** is one of the most convenient methods for the evaluation of binding processes. Since the intensity and wavelength are very sensitive to the change of the environment due to the ligand binding, variation in fluorescence intensity as a function of ligand concentration provide information about the strength of the protein-ligand interaction.

# CIRCULAR DICHROISM

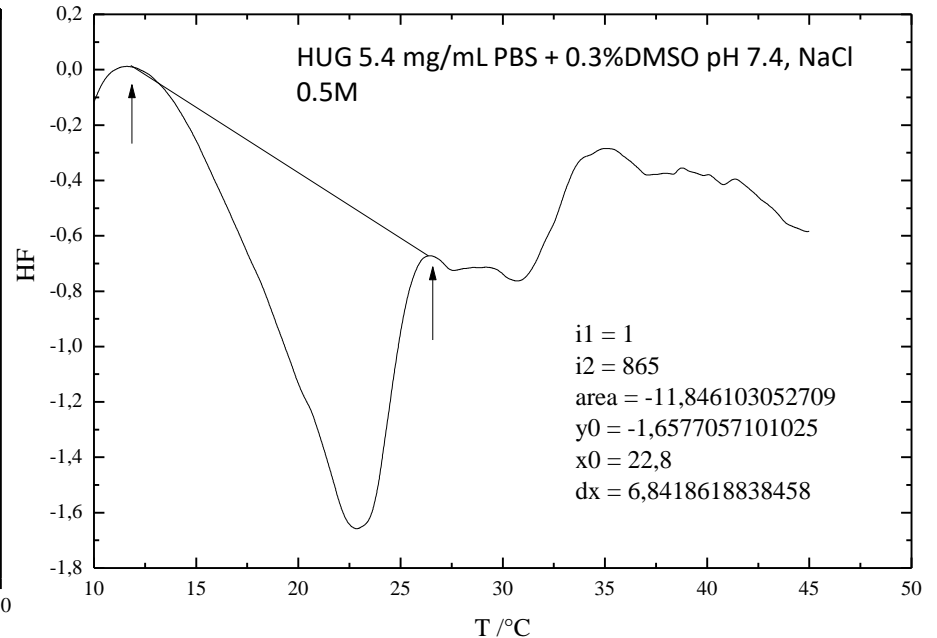
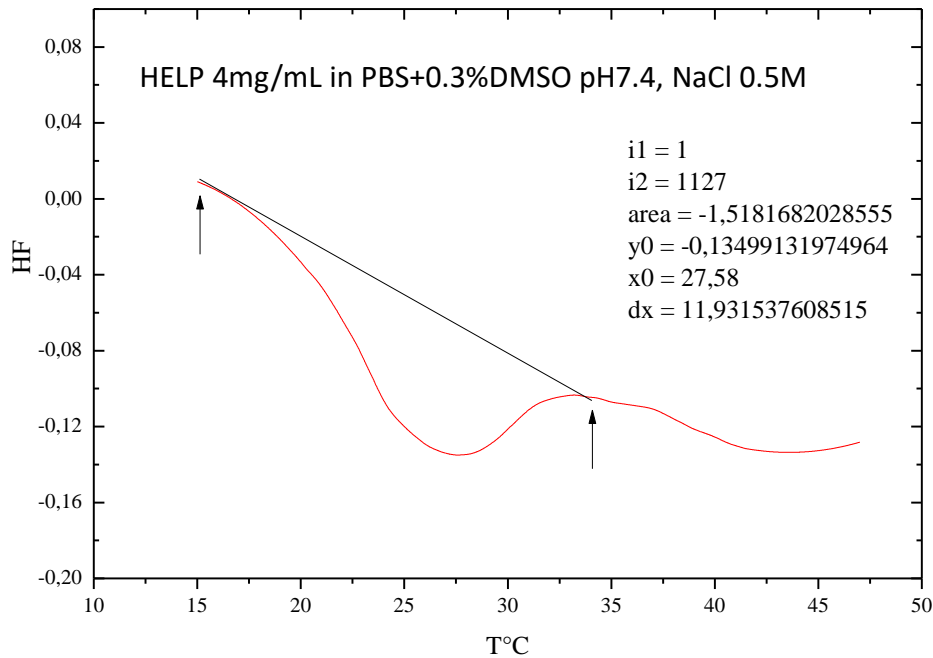
CD is an excellent tool for rapid determination of the secondary structure and folding properties of proteins that have been obtained using recombinant techniques. The most widely used applications of protein CD are to determine whether an expressed protein is folded. In addition, it can be used to study protein-ligand interactions.



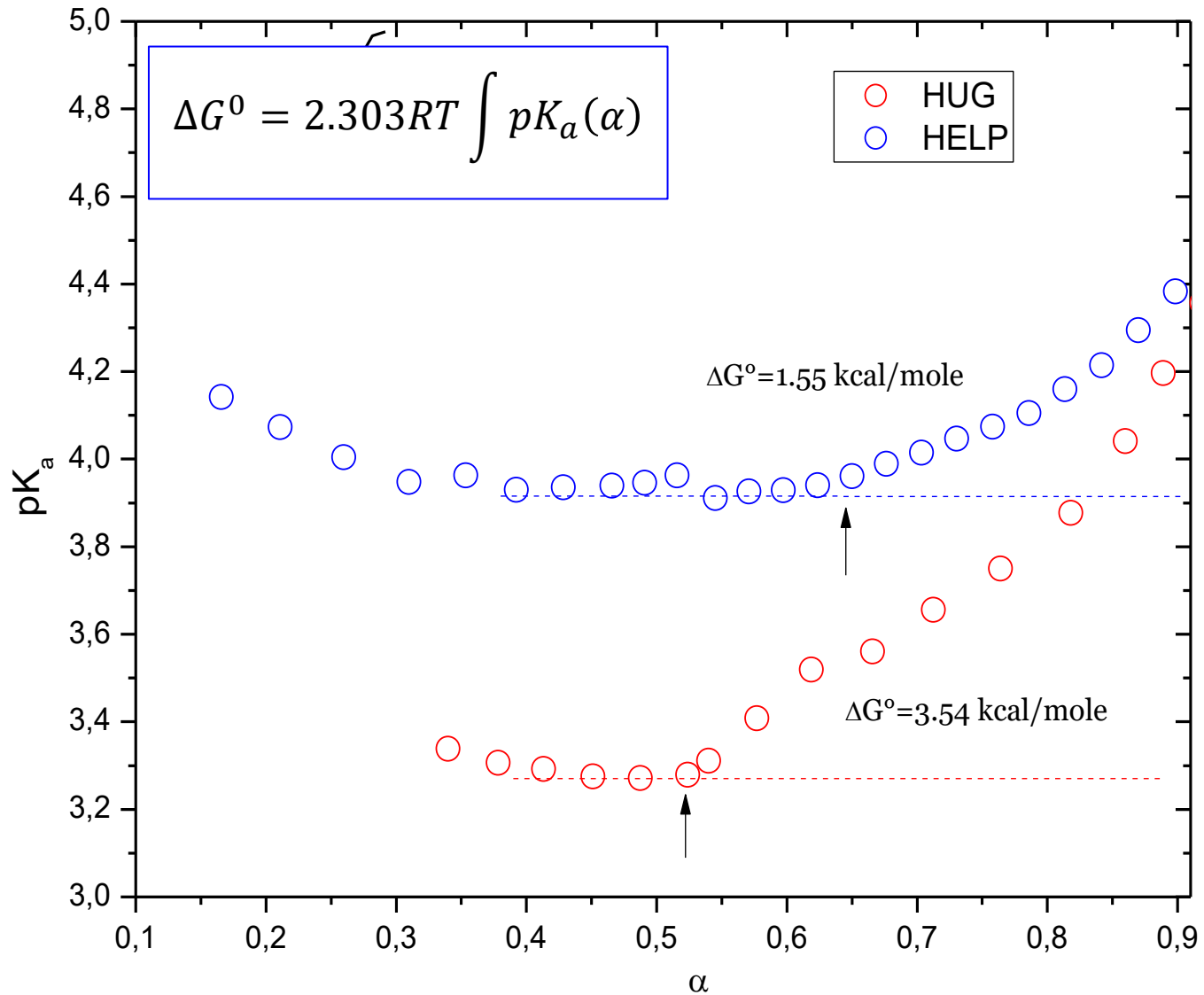
# DSC SETARAM (preliminary results)

The DSC approach employs a reference cell and a sample cell heated at a controlled rate. It can detect some of transitions such as melts, glass transitions, phase changes.

Calorimetric methods demonstrate an advantage over other methods in their ability to provide a relatively full picture of thermodynamic parameters during binding reactions, including the equilibrium binding constants, the enthalpy of binding reactions ( $\Delta H$ ) and the entropy change ( $\Delta S$ ).



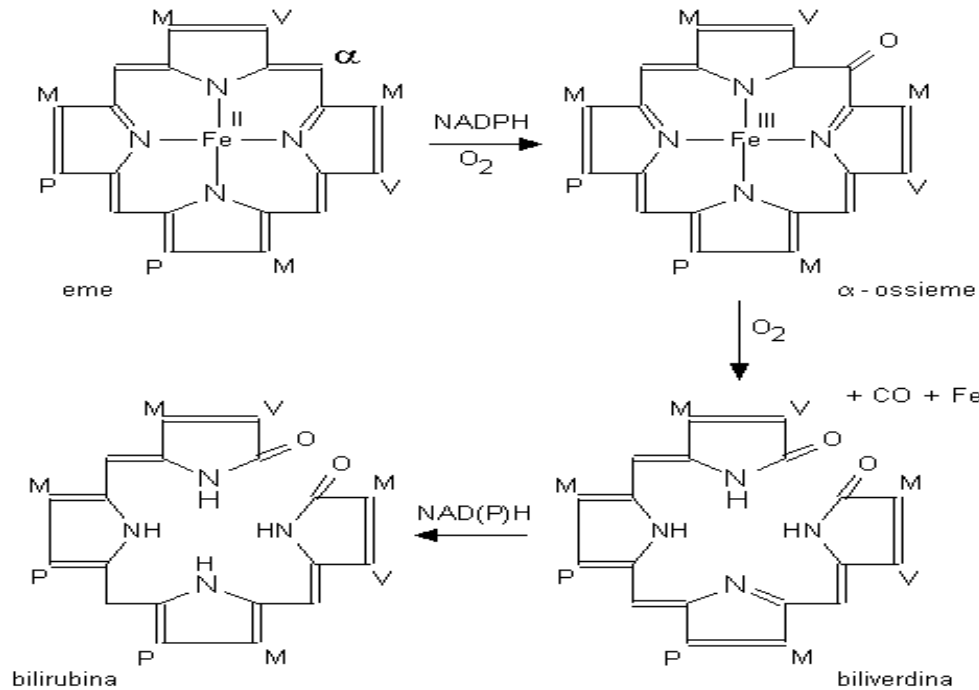
# Potentiometric titration of HUG and HELP





La **bilirubina** (dal latino bilis = bile e ruber = rosso) è un pigmento di colore rosso-arancione che deriva per l'80% dal catabolismo dell'eme dell'emoglobina (Hb) e per il restante 20% da altre emoproteine, come le mioglobine e i citocromi.

La bilirubina si ottiene dal processo di degradazione della ferroporfirina, o gruppo eme, dell'emoglobina, rilasciata durante la distruzione dei globuli rossi, o eritrociti, nella milza.



il metabolismo della bilirubina non coniugata avviene a livello epatico, dopo essere stata trasportata nel sangue da una proteina di trasporto, l'albumina, in virtù della sua insolubilità in acqua (liposolubilità);

## BILIRUBIN

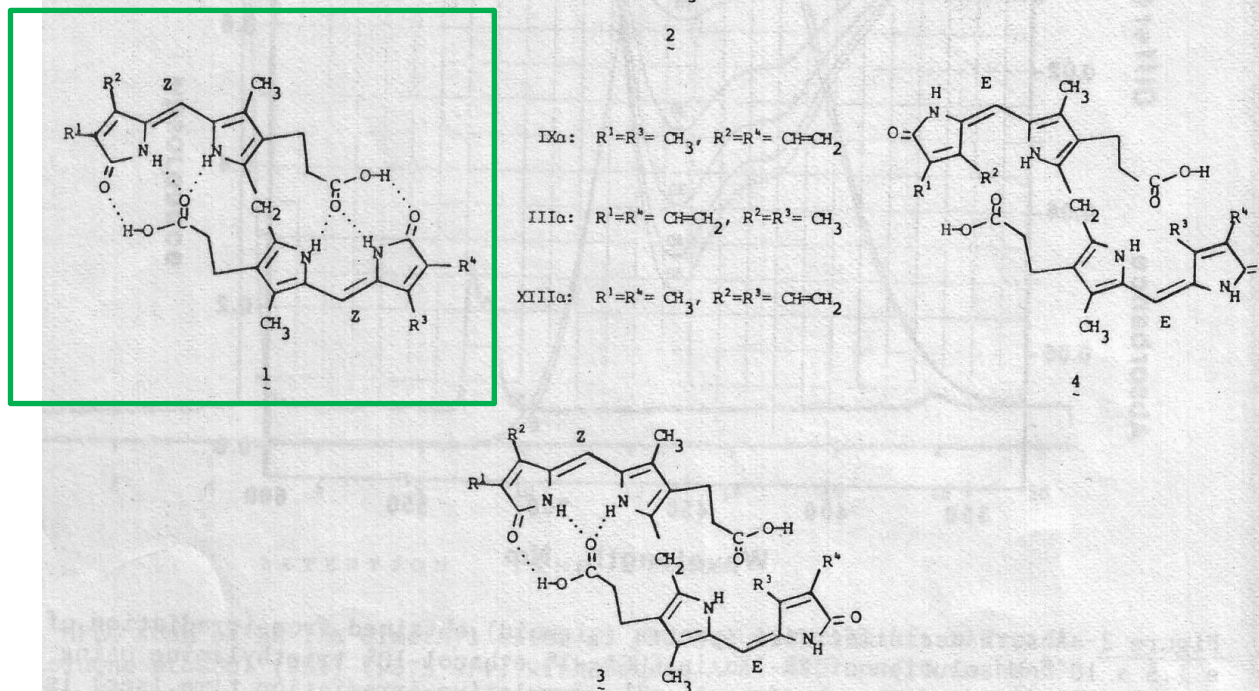
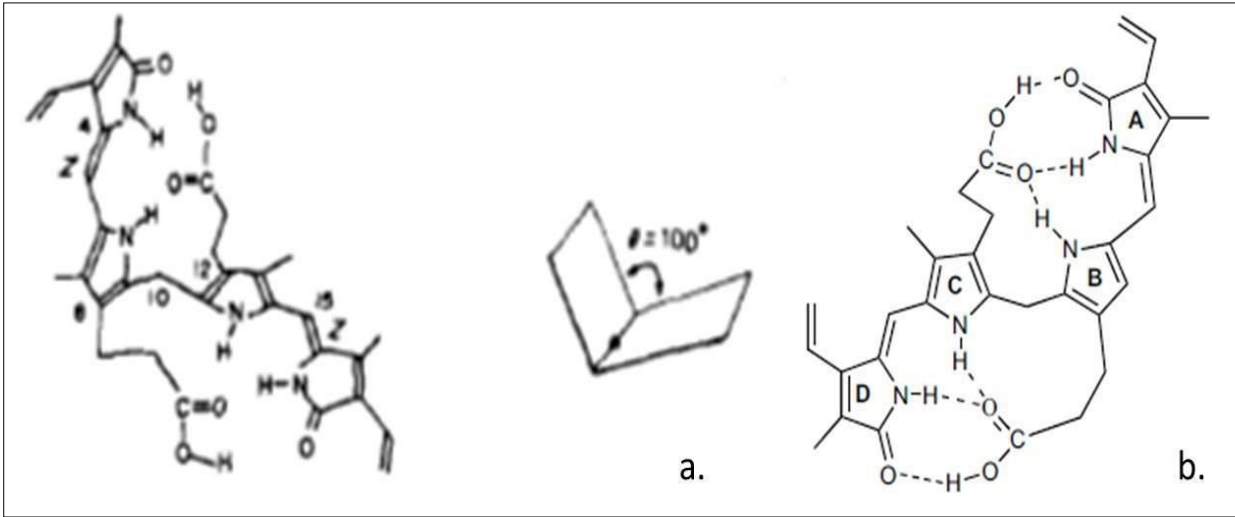
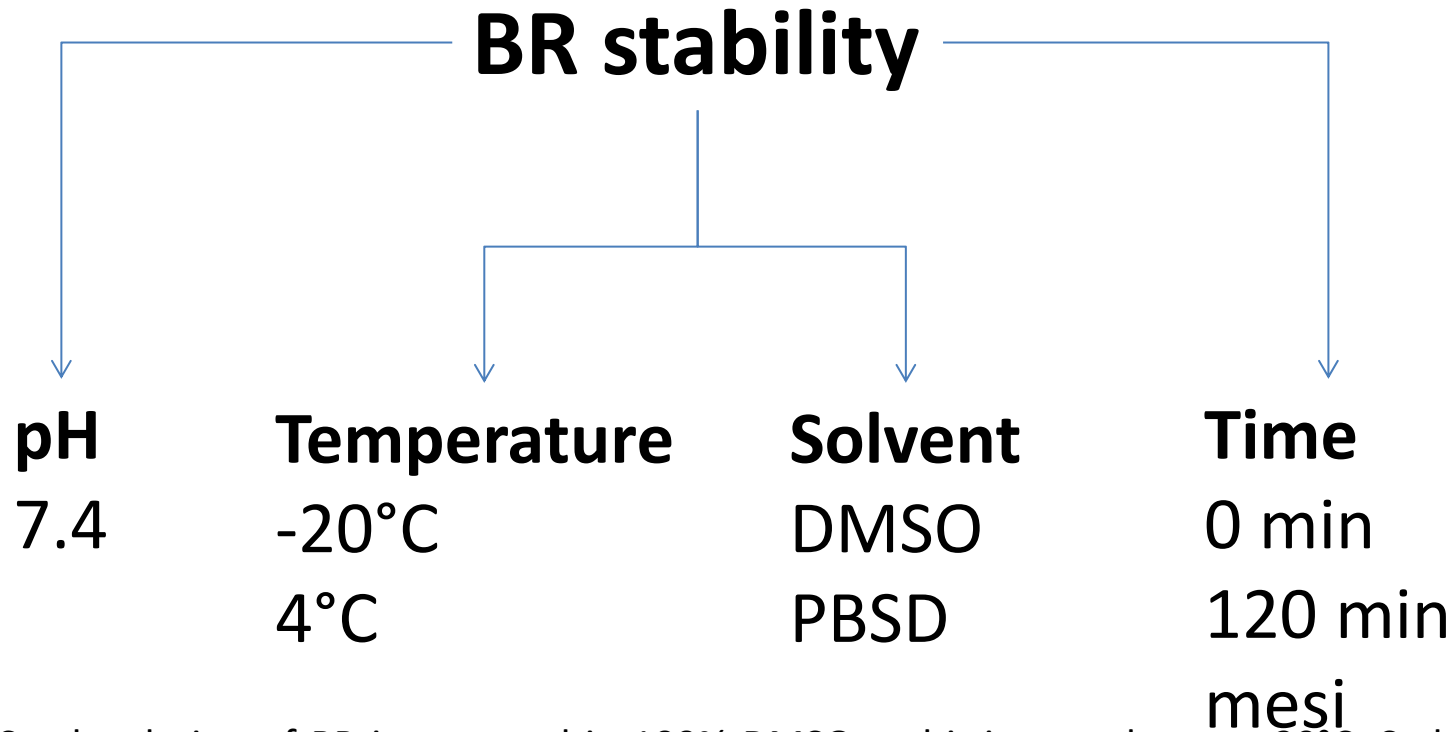


Figure 1 Configurational isomers of BR showing geometric isomerism about the 5,15 *meso* double bonds: 1 = Z-Z, 2 = E-Z, 3 = Z-E and 4 = E-E. The Z-Z configuration (1) is the stable ground state structure of BR (1). The E-Z (2), Z-E (3) and E-E (4) structures are the photochemically accessible isomers which, in the case of BR-IX $\alpha$ , we call collectively PBR. The E-Z (2) and Z-E (3) isomers are identical for (symmetrical) BR-III $\alpha$  and BR-XIII $\alpha$  but not for BR-IX $\alpha$ . In the corresponding BR dimethyl esters, the propionic acid groups become methyl propionate groups.



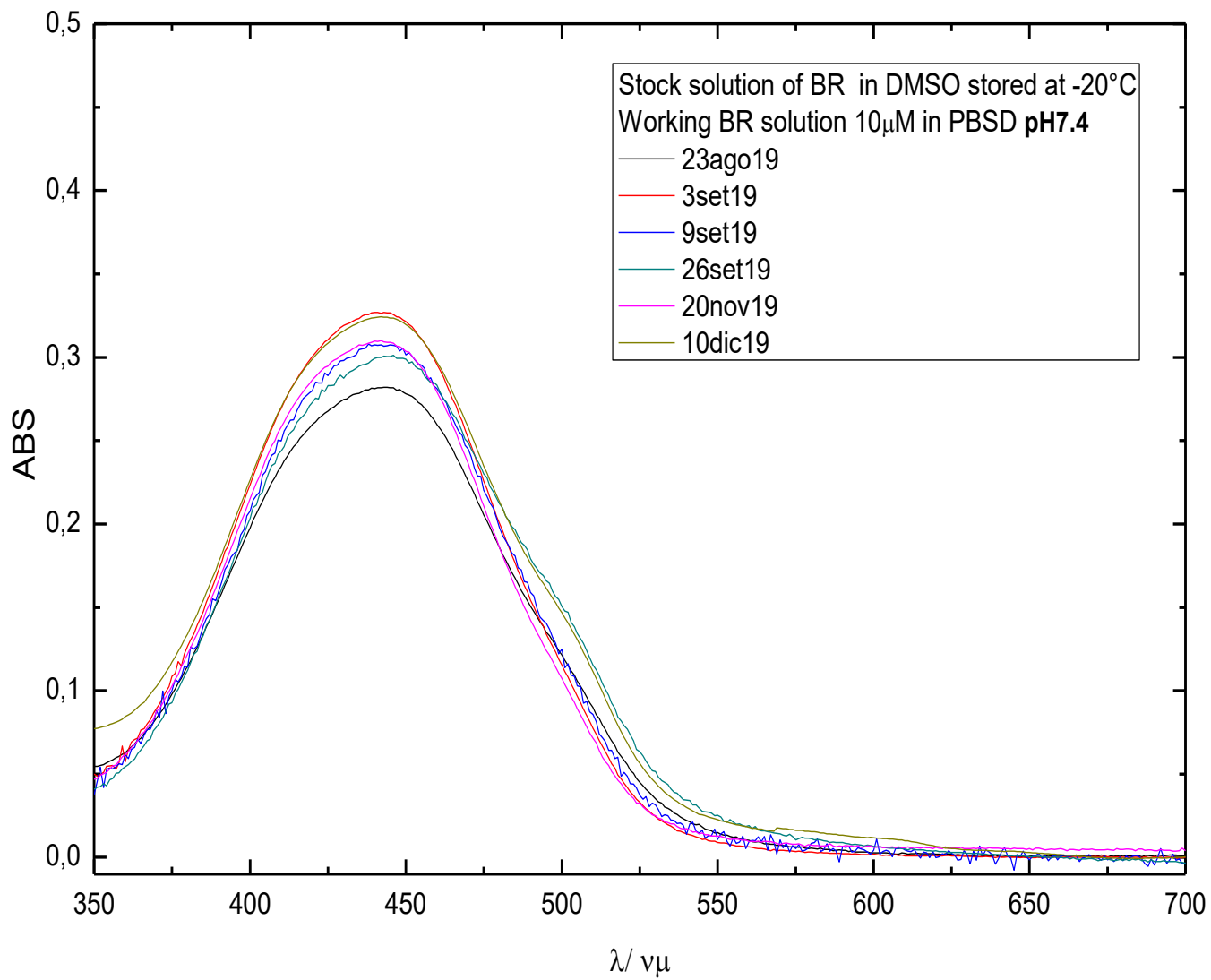
we assessed the bilirubin stability at given experimental conditions:



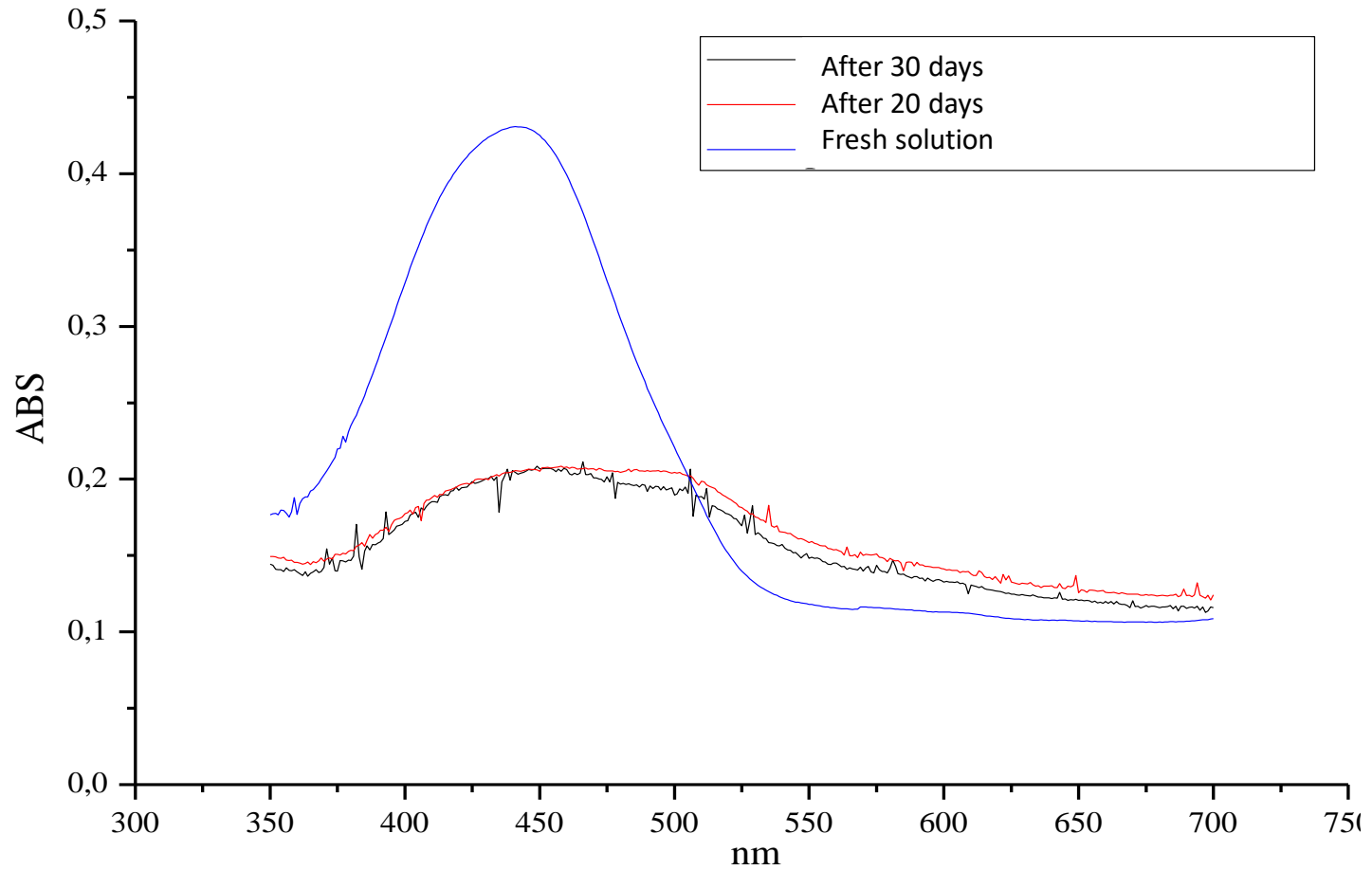
Stock solution of BR is prepared in 100% DMSO and it is stored up to -20°C. Stability assay was done on working bilirubin solution (**BR 10µM in PBS + 0.3%DMSO**). This mother solution is used to prepare the standard solutions.

NB “To prevent the denaturation of UnaG, the final concentrations of DMSO were at most 0.2% (vol/vol) in all experiments.” Shitashima 2017

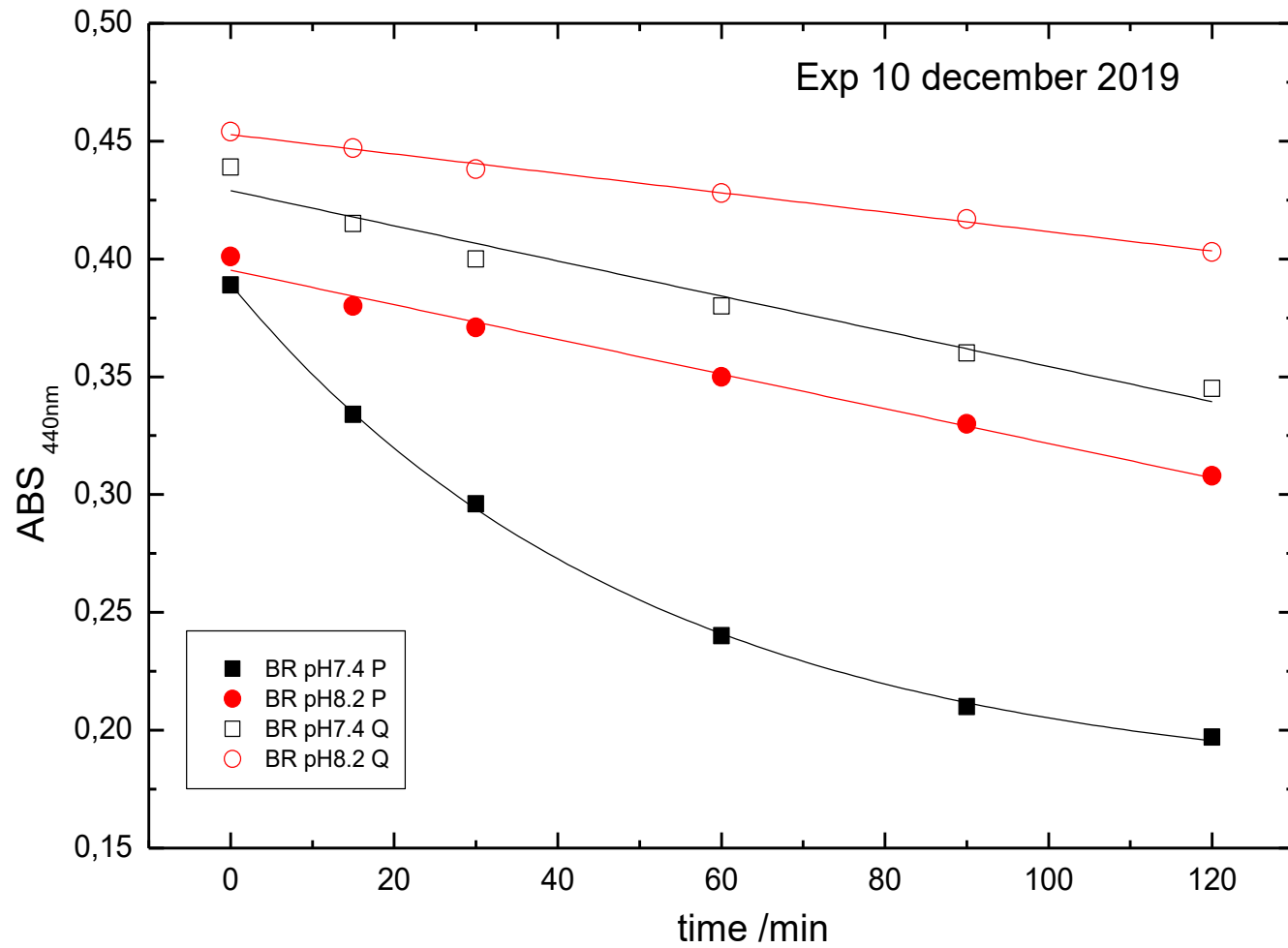
***BR is stable when the solution is stored at -20°C***



**Working solution (PBSD pH 7.4) is not stable if it is stored at -20°C**

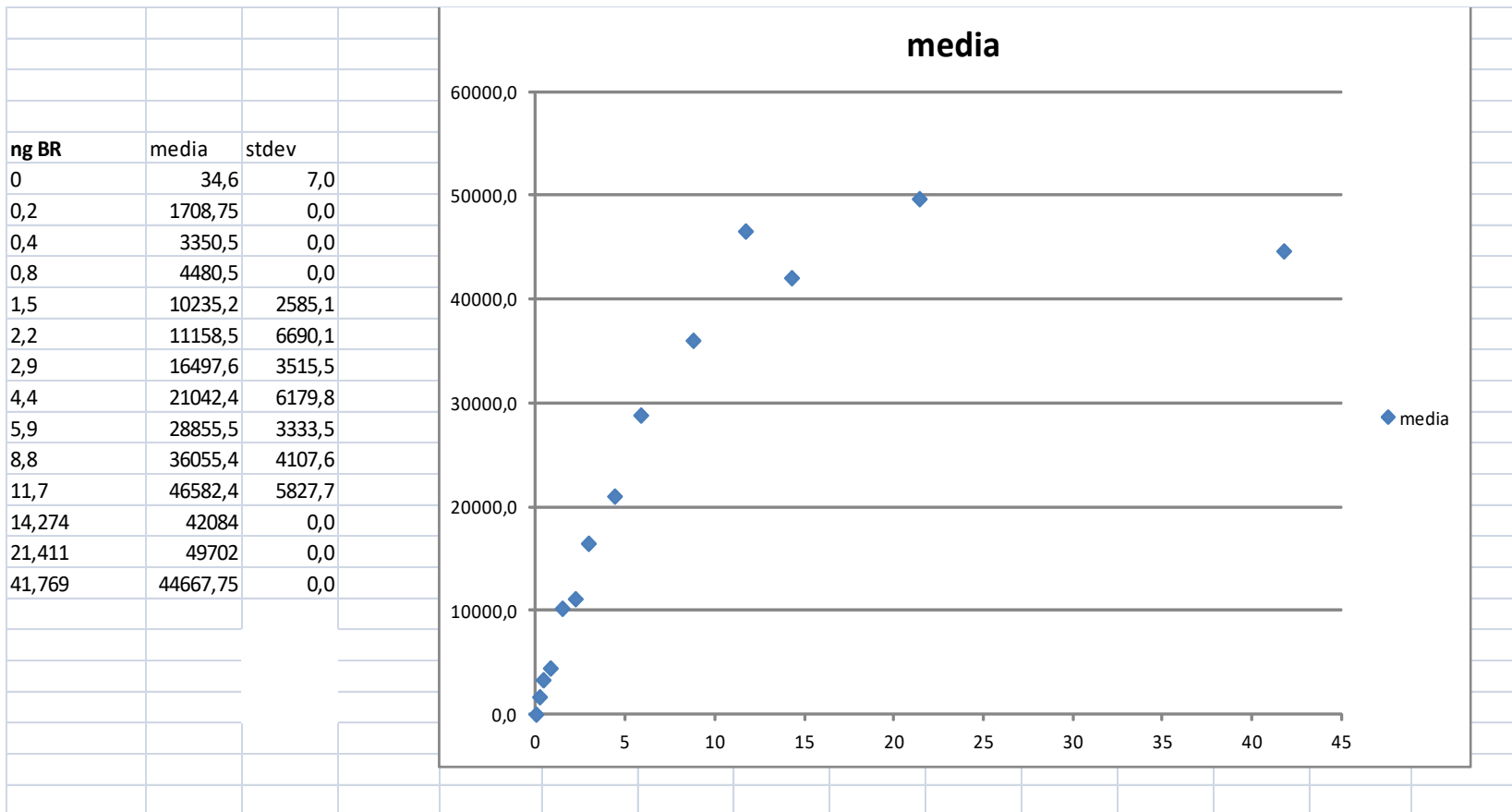


# BR stability is influenced by surface effects (plastic or quartz cuvettes)



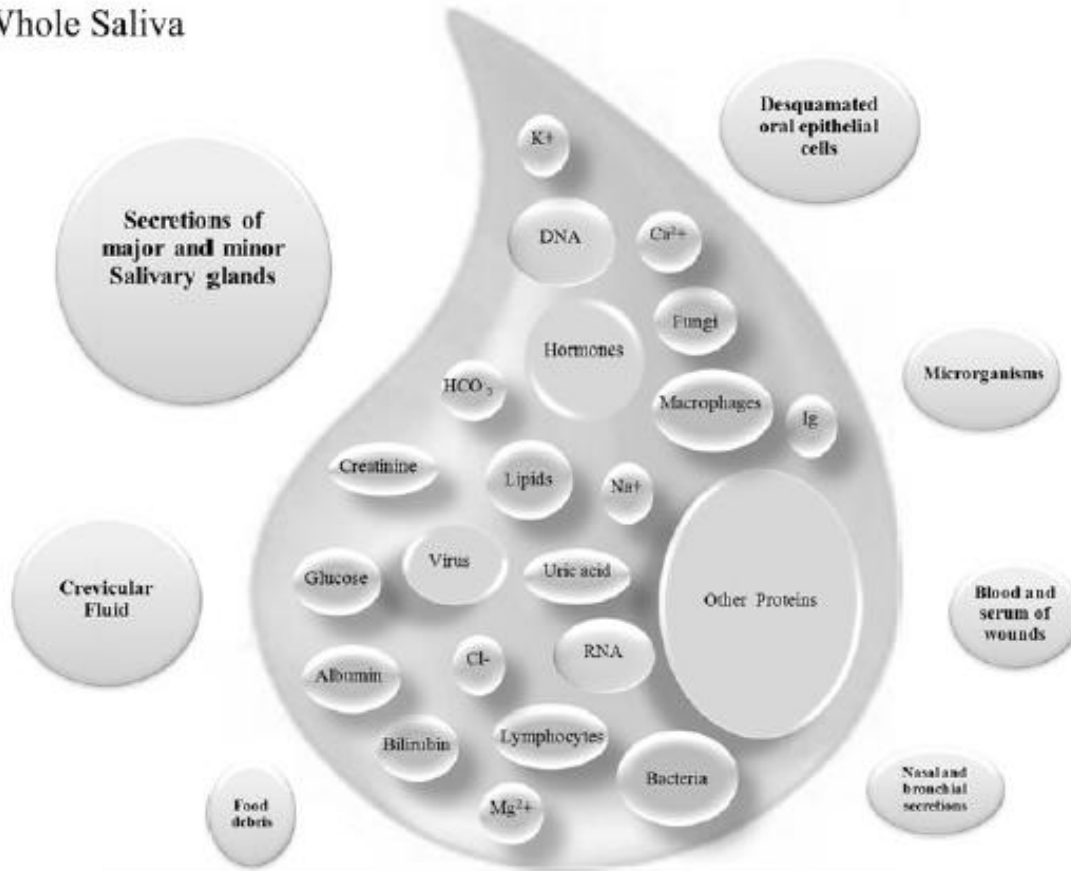
# HUG-BR interaction

The estimate of unconjugated bilirubin concentration has been performed on 96-wells plate in liquid or solid phase (PBSD pH 7.4)

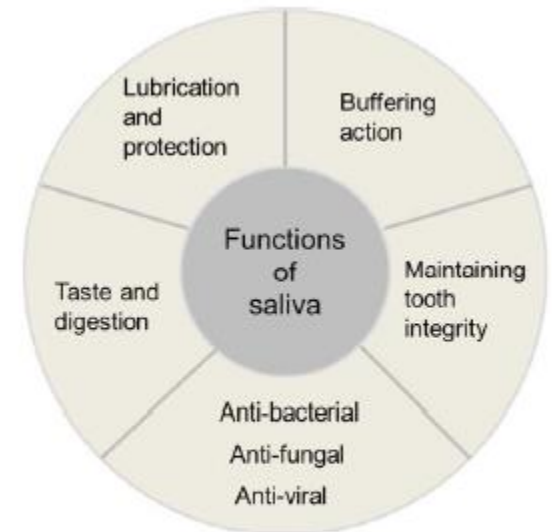




Whole Saliva



**FIG. 1.** Schematic representation of the components of whole saliva. The size of each individual droplet is an approximate representation of its concentration in whole saliva.



**Figure 1: Main functions of saliva**

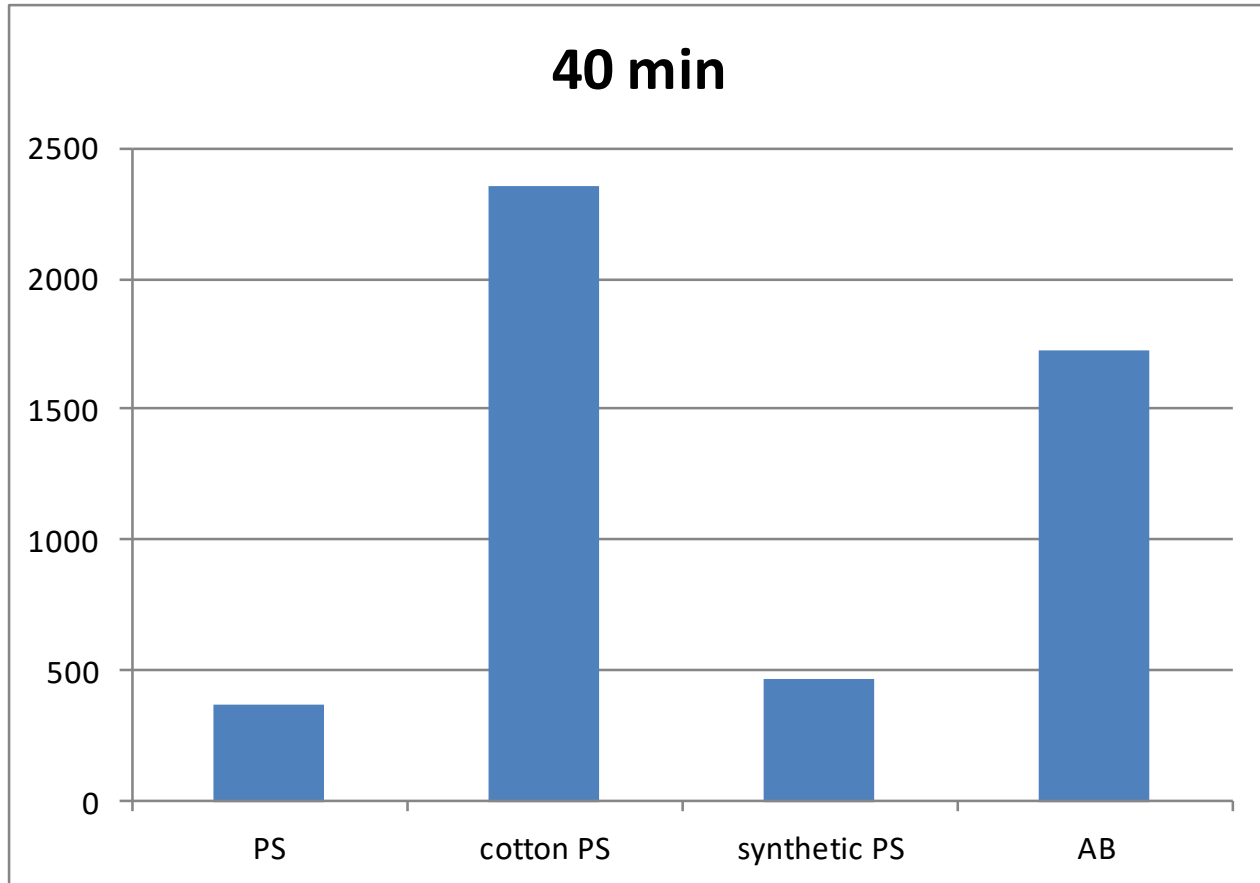
**Table 1** Components of authentic human saliva and a comparison of the normal range of the concentrations between saliva and other biological fluids

Real saliva compositions		Normal range		Ref			
		Saliva	Other biological fluids:				
1. Inorganic compounds	Na <sup>+</sup>	20-80 mmol/L	Plasma 145 mmol/L	11			
	K <sup>+</sup>	20 mmol/L	4 mmol/L				
	Ca <sup>2+</sup>	1-4 mmol/L	2.2 mmol/L				
	Cl <sup>-</sup>	30-100 mmol/L	120 mmol/L				
	HCO <sub>3</sub> <sup>-</sup>	15-80 mmol/L	25 mmol/L				
	Phosphate	4 mmol/L	1.2 mmol/L				
	Mg <sup>2+</sup>	0.2 mmol/L	1.2 mmol/L				
	SCN <sup>-</sup>	2 mmol/L	< 0.2 mmol/L				
	NH <sub>3</sub>	3 mmol/L	0.05 mmol/L				
2. Organic compounds (non-protein and lipids)	Uric acid	3.38 ± 0.21 mg/dL 217.2 ± 110.3 mol/L 0.1-7.5 mg/dL	Serum 6.31 ± 0.24 mg/dL	21-23			
	Bilirubin	0.5-5.0 μmol/L	Serum 0.2-1.2 mg/dL		24		
	Creatinine	0.13 - 0.06 mg/dL	Serum 0.09 - 0.17 mg/dL		25		
		0.05-0.2 mg/dL	Serum 0.6-1.5 mg/dL				
	Glucose	91.3 ± 10.1 mg/dL 4-13 mg/dL	Plasma 80-120 mg/dL		22, 26		
	Cholesterol	0.02-5.46 μmol/L	Serum <5 mmol/L		27		
	Lactate	0.3-1.8 mM 0.1 to 2.5 mmol/L	Serum 0.5-1.0 mM		28, 29		
	3. Protein/Polypeptide compounds	a-Amylase	19-308 U/mL* 93 ± 62 U/L* 2.64 ± 1.8 mg/mL		Serum 0.05-0.125 U/mL*	25, 30	
		Albumin	0.2 ± 0.1 mg/mL		Serum 3.5-5.5 g/dL		31
		Secretory-IgA	80-717 mg/dL 124.3-333.5 μg/mL		Serum 70-400 mg/dL		25, 32
Mucins group		MUC5B: 2.4 ± 1.7 U/mL 1.19 ± 0.17 mg/mL	Serum 9.9 ± 0.8 ng/ml	31, 33			
Lysozyme		3-50 μg/mL 59.7 to 1062.3 μg/ml	Serum 7.4 ± 1.8 mg/mL Serum 4-9 μg/mL	33, 34 34			
Total proteins		7.1-223.2 mg/dL 0.9 ± 0.2 mg/mL	Serum 6-8 g/dL	25, 31			
4. Hormones		Cortisol	3.5-27.0 mg/dL	Serum 2-25 mg/dL	35		
		Testosterone	32-55 pg/mL	Serum 320-600 ng/dL	36		
		Progesterone	Luteal phase 436 ± 34 pmol/L	Serum Male: < 1 ng/mL	37		
			Follicular phase 22.1 ± 2.7 pmol/L	Serum Female: 0.1-20 ng/mL			
	Estrogen(Estradiol)	Luteal phase 20.6 ± 0.4 pmol/L	Serum Male: 15-60 pg/mL	37			
		Serum Female: 15-370 pg/mL					

\*U/mL: enzymatic activity per unit (mL) of saliva

## Preliminary tests on saliva

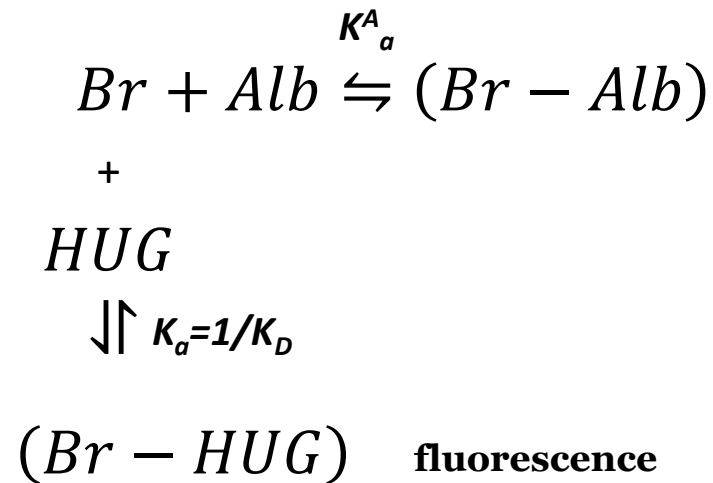
Test with 10 uL of HUG and 150 uL of saliva

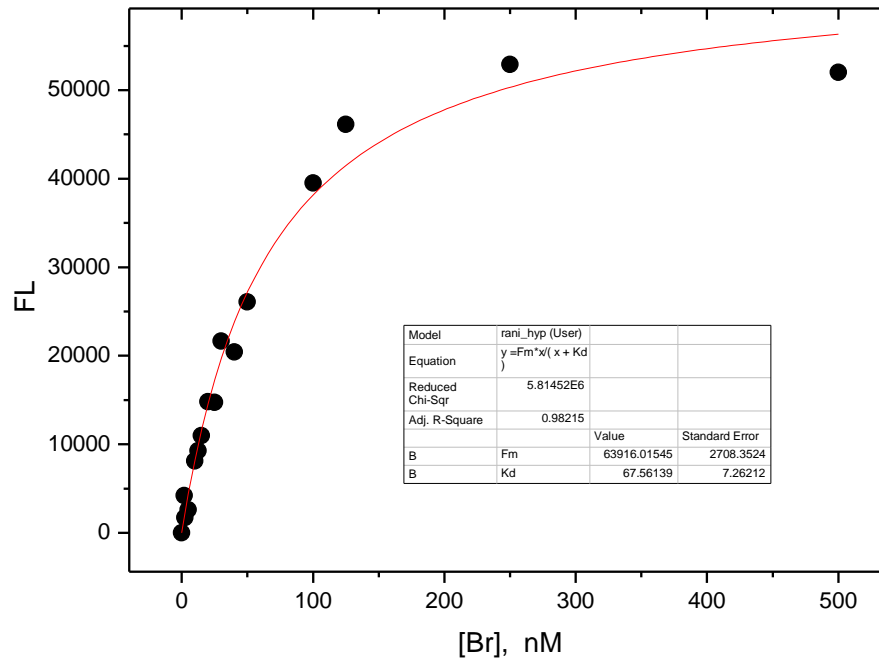


**ATTENZIONE:** non e' stata considerata la fluorescenza spontanea della saliva!!!

## Binding of bilirubin to HUG protein

The Br-HUG interaction was studied by mean of the fluorescence titration technique assessing the intensity enhancement during ligand addition





For each fluorescence value ( $F$ ), the fractional enhancement ( $Y$ ) was computed by the equation:

$$Y = \frac{F}{F_0}$$

where  $Y$  is the fractional saturation related to the extent of binding and  $F_0$  the fluorescence intensity at the Br/HUG ratio greater than 1 (the asymptotic value).

$$K_D = \frac{[HUG][Br]}{[Br - HUG]}$$

During the binding titration the Br concentration is increased so that saturation  $Y$  is expressed in terms of the Br-HUG complex concentration,  $[Br-HUG]$  as:

$$Y = \frac{[Br - HUG]}{[HUG]_T} \quad (1)$$

where  $[HUG]_T$  is the total HUG concentration used for the measurement. Then:

$$[Br - HUG]_{\square} = \frac{[HUG][Br]}{K_D}$$

$$[HUG]_T = [HUG] + [Br - HUG] = P_T$$

$$Y = \frac{[HUG][Br]/K_D}{[HUG] + [HUG][Br]/K_D}$$

$$Y = \frac{[Br]}{K_D + [Br]}$$

Since in the binding measurement instead of free concentrations the total protein and ligand concentrations are known, an expression of  $Y$  as a function of total quantities is derived as follows:

$$[Br] = [Br]_T - [Br - HUG] = L_T - [Br - HUG]$$

$$[HUG] = P_T - [Br - HUG]$$

then:

$$K_D = \frac{(L_T - [Br - HUG])(P_T - [Br - HUG])}{[Br - HUG]}$$

$$K_D[Br - HUG] = P_T L_T - (P_T + L_T)[Br - HUG] + [Br - HUG]^2$$

$$[Br - HUG]^2 - (P_T + L_T + K_D)[Br - HUG] + P_T L_T = 0$$

The root of the equation is:

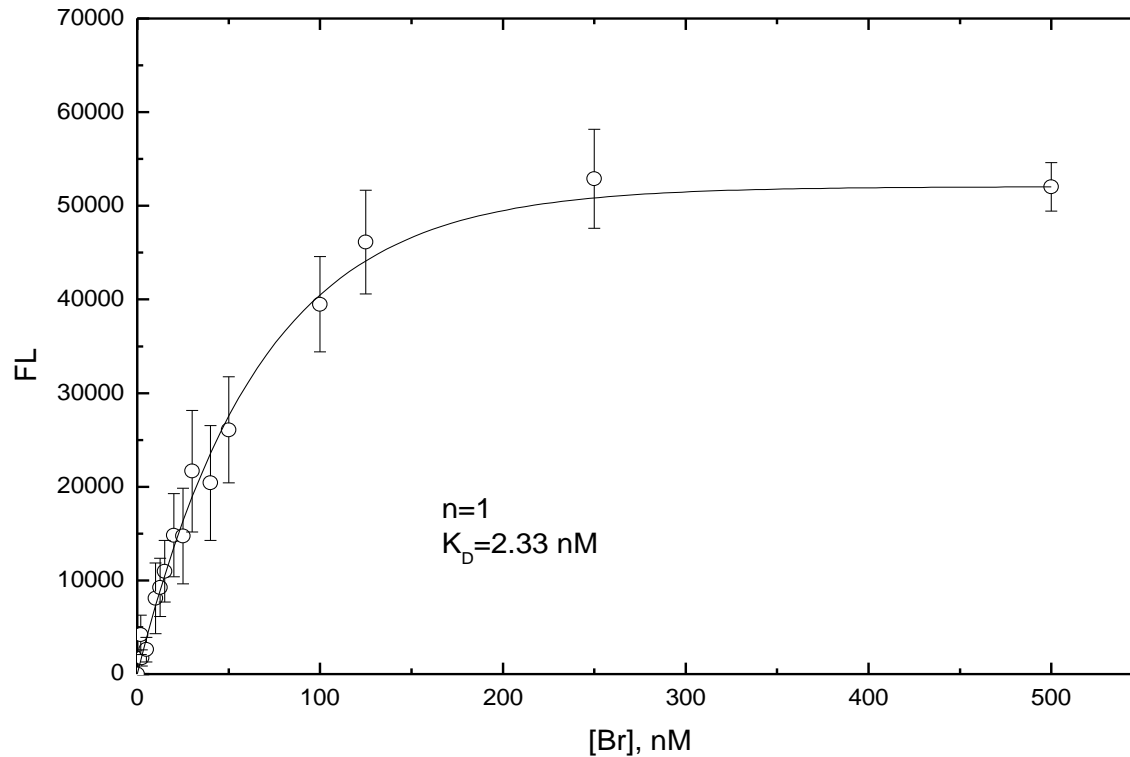
$$[Br - HUG] = \frac{(P_T + L_T + K_D) - \sqrt{\{(-(P_T + L_T + K_D))^2 - 4P_T L_T\}}}{2}$$

By substitution in the equation (1):

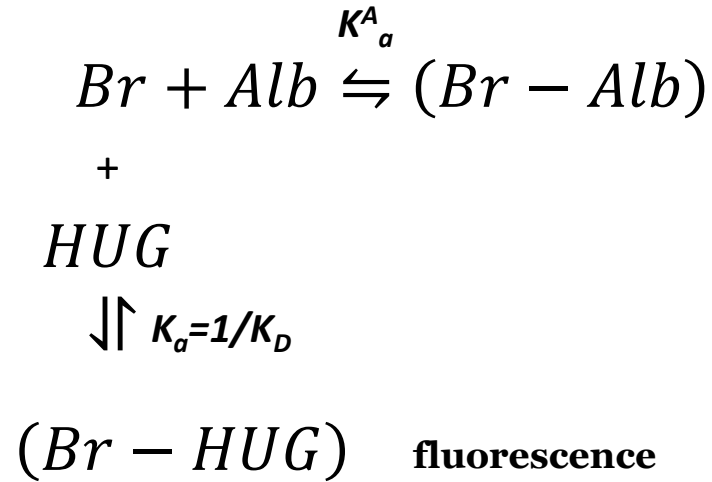
$$Y = \frac{(P_T + L_T + K_D) - \sqrt{\{(-(P_T + L_T + K_D))^2 - 4P_T L_T\}}}{2P_T} \quad (2)$$



The plot of  $F=F_oY$  versus  $L_T = [Br]$  is shown in Figure where average values of all the results obtained by several experiments are reported.



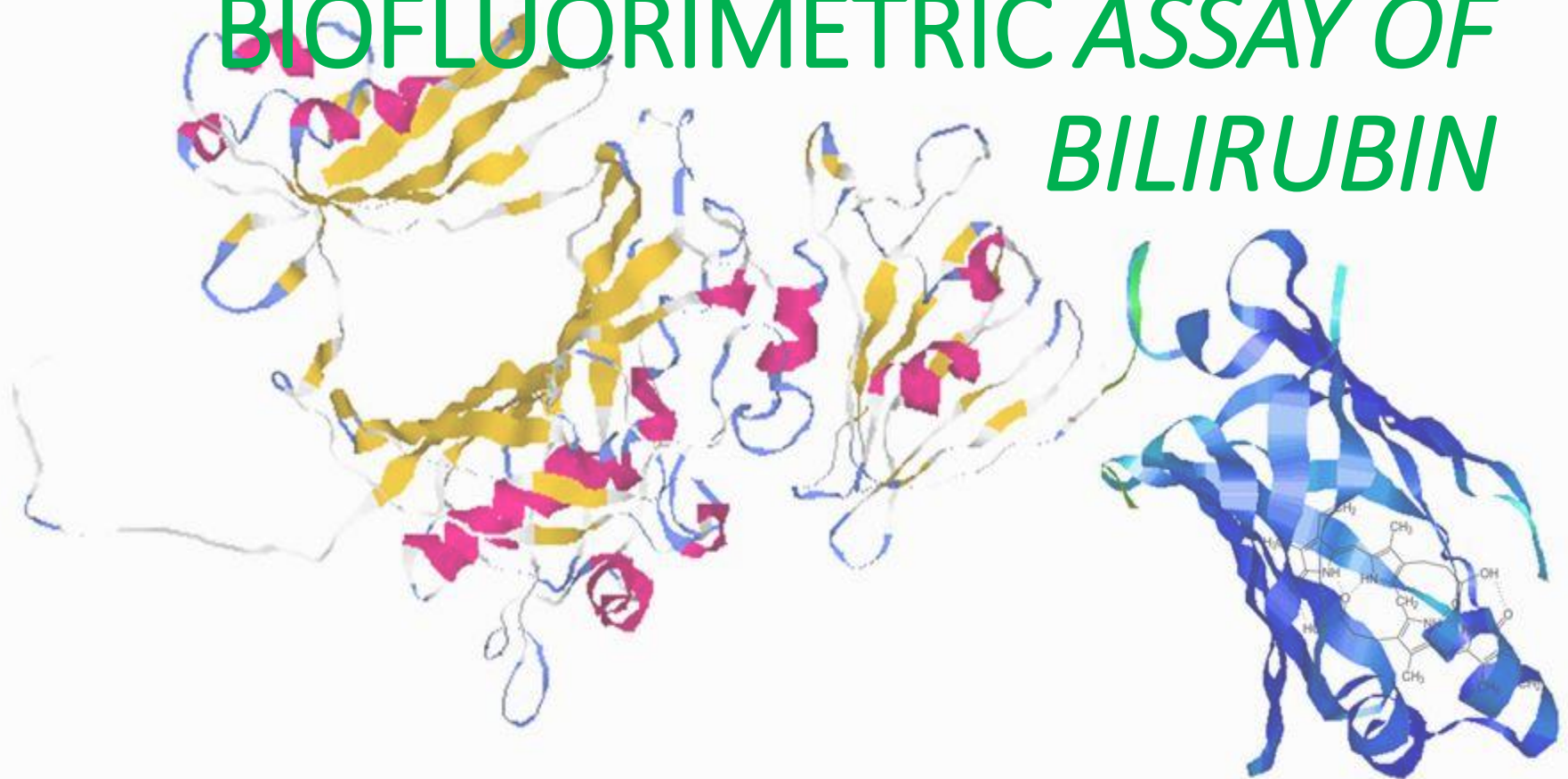
By nonlinear least-squares fitting of the hyperbolic curve the best evaluation of equilibrium dissociation constant ( $K_D=2.33$  nM) and of the maximal fluorescence value of the ligand-bound protein ( $F_o=56300$ ) was obtained



$$K_a=1/K_D = 4.3 \times 10^8 \text{ M}^{-1} \text{ per } Br-HUG$$

$$K_a^A=1/K_D = \text{ca } 10^6 \text{ M}^{-1} \text{ per } Br-Alb$$

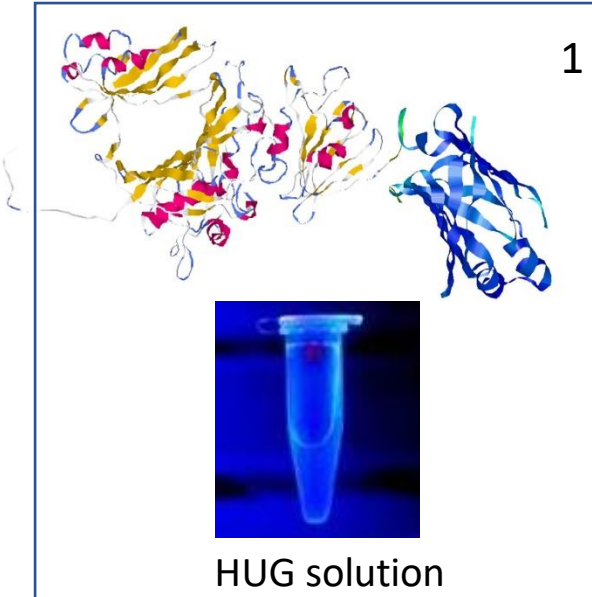
# BIOFLUORIMETRIC ASSAY OF *BILIRUBIN*



# BIOFLUORIMETRIC ASSAY OF SERUM BILIRUBIN

- Direct analysis of indirect bilirubin by HUG

1 HUG is a synthetic protein with a bilirubin-binding domain (UnaG) fused with Human Elastin-like Polypeptide

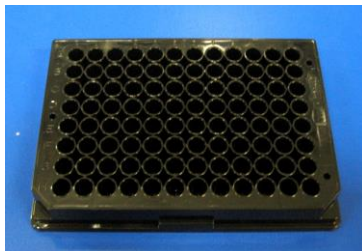


Add sample

2. HUG binds bilirubin in a sample (< 30  $\mu$ L) and emits fluorescence



3. Samples are placed in multiwell plates



4. Fluorescence is measured by reader

The emission fluorescence of the complex was detected at  $\lambda = 528$  nm following excitation at  $\lambda = 485$  nm



## Future goals:

1. Molecular Dynamics simulation of the single protein (HUG and HELP),
2. Molecular Dynamics of binding process (HUG-BR),
3. A thorough study of the reverse thermal transition of HUG in comparison to HELP (which is a critical property for a future upscale of biopolymer production),
4. Bilirubin assay on fish serum samples.

