

Distinctive patterns of transcription and RNA processing for human lincRNAs

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History

1989: RNA world concept 2007: ENCODE Projed 1955: first ncRNA housekeeping: rRNA 1958: Central dogma of molecular bilolog **1998: RNAi** 2005: Antisense transcription ● 1983: first sncRNA: micF 2002: Pervasive transcription **2012: ENCODE** (Chromosome 21, 22) ~80% of the human DNA-based discoveries 2008 genome is transcribed ● 1989: first lncRNA: H19 + RNA world 1993: lin-4, first miRNA **PROMPTs mRNA** 2007: HOTAIRPRC2 2000: let-7 miRNA 1991: Xist concept ncRNA discoveries 2009: LincRNAs/PRC2 1989: H19, first eukaryote ncRNA ● 1991: XIST Housekeeping ncRNA 2009: unstable ncFNAs (yeast) 1960 1983 micF: bacterial mRNA **ncRNA 2006: NRON, BIC** 1998: 2012: circRNAs ● 2001: HGP completed Tsix. 1957 1980: Ribozvmes 2005: PCGEM1 Air, Konglott tRNA 2020 ● 2004: Only 1,2% of the human genome 1976: 2003: MAL AT1/cancer نتسط eRNAs snoRNAs and rRNA processing 1955 1968: TERRA 2002: MEG3, DISC **rRNA** snRNAs and splicing T-UCRs codes for proteins1960 1970 1980 1990 1995 2000 2005 2010 2015 1869: << Nudein >> 1953: Double-helix DNA 2001 : Human gen ome sequence 1939: Link between proteins and FNA 2002: Mouse genome sequence 1944: DNA carries 1968: Genetic code 2004: 98.8% of the human genome is non-coding Ash of the Book of the genetic material Important March 2006 EC DEFENSE DEFENSE AND DEFENSE OF STRAIGHT **ABUSTAL** echnological achive ments

non-coding RNA

Chromatin topology: gene transcription regulations

Scaffolding and modulating the activity of proteins and RNAs

Protein and RNA decoy

Encoding functional micropeptides

Chromatin topology: gene transcription regulations

Epigenetic markers derive from the recruitment of methyl-transferase binded to lincRNA such HOTTIP pr HOTAIR

Scaffolding and modulating the activity of proteins and RNAs (RepA-EZH2)

Interaction between RepA and PRC2 demonstrated by an Electophoretic Mobility Supershift Assay

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G ā G c C A Mut

Protein and RNA decoy

Some lncRNA can act as ceRNA

linc- RoR maintains stem cell pluripotency.

In pluripotent stem cells, linc-RoR sequesters miR-145, thereby promoting the accumulation of OCT4, the transcription factor SOX2 and the homeobox protein Nanog, which are miR-145 targets.

The levels of linc-RoR decrease during differentiation, and miR-145 is released and promotes the degradation of SOX2, Nanog and OCT4 mRNAs

Encoding functional micropeptides

lincRNAs can contain little ORFs in their sequence. They are called smORFs.

smORFs are translated in micropeptides that can be functional for another structure (SERCA).

Another meaning for the existence of these smORFs is the presence of an early stop codon that activate the Nonsense-Mediated Decay

Family of SERCA-inhibiting micropeptides A

> Contraction Sarcomere

Relaxation

Sarcomere

Invertebrate

000

Vertebrate

Characterization of lincRNAs

>200 nt

- 5' capping (CAGE)
- 3' polyadenylation (3P-seq)
- Different splicing from mRNA
- Epigenetic markers similar to mRNA
- Inefficiently polyadenylated

Human

- lack of primary structure conservation despite protein-coding gene
- average of 40 Kb compared to other genes
- *NOT* coding for proteins
- *NOT* overlapping with other transcripts
- Degraded by exosomes in the nucleus

lincRNA candidates (no overlap with known ncRNA can have ORFs **imperfect** protein-coding genes) **criteria:** ● Coding regions tend to be much longer than expected by chanceLong ORF UGA **AUG** 234 aa

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Comparison between mRNA and lincRNA: genes

mRNA

About 20.000 genes. Average of 11 exons of 3 Kb each. Higher gene density than lincRNA

lincRNA

About 13.000 genes. Average of 3 exons of 1 Kb each.

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Comparison between mRNA and lincRNA: localization

Comparison between mRNA and lincRNA: phosphorylation of RNA Pol II CTD and RNA maturation

mRNA

They have a peculiar pattern of phosphorylation of RNA Polymerase II's CTD:

Ser5 phosphorylated during early elongation Ser2 phosphorylated during later elongation

And what about the phosphorylation of RNA Polymerase II's CTD in lincRNA?

The paper tries to answer the following questions:

- How does Pol II CTD phosphorylation differ between protein-coding and lincRNA genes?
- Are there any differences between splicing of protein coding and splicing of lincRNA?
- Are lincRNA and protein-coding genes differentially polyadenylated?
- Why are lincRNA levels substantially reduced in the nucleoplasm?
- Are lincRNAs co-transcriptionally cleaved?
- Could lincRNA endonucleolytic cleavage be mediated by the microprocessor?

How does Pol II CTD phosphorylation differ between protein-coding and lincRNA genes?

Specific **Pol II CTD phosphorylation states** are associated with **different stages of transcription**:

- **Ser5P** > early elongation, 5' capping and active splicing
- **Ser2P** > later elongation and 3' end processing
- Y1P, T4P, S7P, unph > additional phosphorylation states

mNET-seq can be used to sequence nascent RNA by employing Pol II antibodies against specific CTD phosphorylation states in order to isolate RNA from immunoprecipitated Pol II

mNET-seq (Mammalian Native Elongating Transcript sequencing)

Key steps:

- RNAPII complexes are isolated through chromatin fractionation
- MNase is used to digest all exposed DNA while leaving RNA strands protected by RNAPII or spliceosomes intact
- RNAPII complexes are immunoprecipitated using RNAPII antibodies and 5' phosphorylated by T4 PNK
- 3' linkers are ligated to the 3' hydroxyl end of the RNA strand
- Nascent RNAs are isolated, size-selected for 35–100 nt, processed into cDNA sequencing libraries, and sequenced

mNET-seq analysis:

- *lincRNA genes* show less pronounced **unph** and **Y1P TSS** peaks and a generally **more even distribution** of mNET-seq reads across the gene body
- *protein-coding genes* show a higher **T4P** signal in the **TES** region compared to lincRNA genes, where the **T4P** signal is more evenly distributed

Conclusion > Pol II termination probably occurs at multiple positions across lincRNA genes

Splicing differences between lincRNA and protein-coding genes

Analysis of specific lincRNAs using splicing specific mNET-seq/S5P profiles:

- When HeLa cells are treated with Pla-B, most **S5P CTD-specific 5'ss peaks** on *protein-coding genes*, such as PTCD3, **are lost**
- *lincRNA genes* are **less sensitive** to Pla-B treatment
- 55-70% of protein-coding introns are associated with 5'ss peaks. In contrast, only 20-30% of lincRNA exons show 5'ss peaks

Conclusion > lincRNAs are inefficiently spliced compared to protein-coding genes

Duplicate HeLa cell transcript libraries from either **pA+** or **pA**nuclear RNA were prepared to measure splicing efficiency directly:

- **pA+ reads** across the *protein-coding gene* **WDR13** are **exon restricted** (> efficient co-transcriptional splicing), with little signal detected in the pA- NpRNA-seq profile
- the *lincRNA* **TUG-1 pA+** profile shows **significant levels of intron reads** over its annotated intron regions, whereas the pA- profile revealed a higher level of intron signal

Comparison of splicing events between these two transcript classes shows a consistently lower splicing rate for lincRNAs

Are lincRNA and protein-coding genes differentially polyadenylated?

Analysis of mNET-seq/T4P datasets shows a close correlation between the CTD **T4P mark** and **protein-coding gene termination**, whereas lincRNAs show a more widespread T4P mNET-profile across the whole transcription unit (TU) Protein-coding

- **Depletion of CPSF73** (cleavage and polyadenylation factor) causes a **substantial decrease in T4P mNET-seq reads over the termination region** of the protein-coding gene **GAPDH**
- The lincRNA **TUG1** mNET-seq/T4P profile is not affected by CPSF73 depletion > **TUG1 termination is CPSFindependent**

Are lincRNA and protein-coding genes differentially polyadenylated?

Meta-analysis of the termination region associated with mNET-seq/T4P profiles shows that protein coding, but not lincRNA gene termination, is strongly affected by CPSF73 depletion

Are lincRNA and protein-coding genes differentially polyadenylated?

pA+ and pA- NpRNA-seq libraries were employed to examine the degree of 3' polyadenylation in lincRNAs:

- *protein-coding transcripts* are predominantly **pA+**
- histone RNAs are exclusively in the pA- fraction because histone mRNA is maturated by a PAS-independent mechanism
- *lincRNAs*, such as LINC01021, are **more pA- than pA+**

Conclusion > lincRNAs are inefficiently polyadenylated compared to protein-coding transcripts

Why are lincRNA levels substantially reduced in the nucleoplasm?

lincRNA and protein-coding gene transcripts are often similar in abundance in the chromatin fraction

Transcription profiles for a tandem lincRNA and protein-coding gene **LBR** show lower levels of lincRNA in the nucleoplasm compared to chromatin-associated lincRNA

RNA-seq data were analyzed for lincRNA expression in the cytoplasm to exclude the possibility of rapid nuclear export > **less cytoplasmic lincRNA is detected compared to chromatin-associated lincRNA**

median (Mean 90%, excluding top 5% and bottom 5%) [stddev 90%]

lincRNAs are substrates of the RNA exosome (shown in mESCs)

ChrRNA-seq and NpRNA-seq following depletion of the RNA exosome component **EXOSC3** > **lincRNAs were all significantly increased in the nucleoplasm**

Comparison of the ratio of chromatin to nucleoplasm RNA levels between protein-coding and definable classes of lncRNAs following exosome depletion:

- protein-coding RNA levels are slightly stabilized
- **all categories of lncRNAs show significant nucleoplasmic stabilization**
- tRNAs, structural ncRNAs and small nuclear RNAs were significantly destabilized > known role of the exosome in tRNA and snRNA maturation

Conclusion > lincRNAs are post-transcriptionally degraded by the nuclear exosome

- The mNET-seq technique involves the ligation of a linker oligonucleotide onto any RNA 3' end protected from micrococcal nuclease digestion
- RNA 3' ends principally derive from the Pol II active site, reflecting nascent transcription
- Co-precipitated RNA processing complexes can generate RNA 3' ends (detected by mNET-seq): e.g. splicing intermediates or microRNA precursors

Empigen is employed to separate mNET-seq reads derived from Pol II active site RNA 3′ ends and those derived from co-precipitated RNA processing complexes

mNET-seq after Empigen treatment:

● *MYC* gene: S5P-specific 5′splicing sites peaks are specifically lost

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mNET-seq after Empigen treatment:

- *MYC* gene: S5P-specific 5′splicing sites peaks are specifically lost
- lincRNA *MIR17HG:* S5P-/S2P-specific microprocessor-mediated RNA cleavage intermediate is lost
- *MALAT1* and *LINC01021*: lots of S5P and S2P peaks are reduced

Conclusion > lincRNAs are co-transcriptionally cleaved at multiple positions across their TUs and most Empigen-sensitive lincRNA peaks are insensitive to Pla-B treatment, indicating that they are distinct from splicing intermediates

mNET/S5P datasets using chromatin from HeLa cells depleted for either DGCR8 (a double-stranded RNA binding protein) or Dicer. DGCR8 depletion also inactivates Drosha as an integral part of the microprocessor.

● protein-coding gene *CCND1*: neither DGCR8 nor Dicer depletion affected mNET-seq/S5P profiles

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- protein-coding gene *CCND1*: neither DGCR8 nor Dicer depletion affected mNETseq/S5P profiles
- *MIR17HG*, which encodes the miR17-92a cluster: DGCR8 depletion affected mNETseq peaks corresponding to release of these pre-miRNAs.
- lincRNA: neither loss of DGCR8 nor Dicer caused a general loss of mNET-seq/S5P peaks

Seems that these endonucleases have not a role in lincRNA cleavage

- DGCR8 interacts with nuclear RNA exosome components, independently of the endonuclease Drosha, facilitating exosome recruitment to degrade abundant lncRNAs.
- DGCR8, but not Dicer, depletion acted to selectively increase Empigen-sensitive mNET-seq/S5P peaks on lincRNA genes (*MALAT1* and *LINC01021*).

Conclusion > DGCR8 also acts to recruit the exosome to co-transcriptionally cleaved lincRNA, independently of miRNA

PCA reveals lincRNAs are generally distinct from protein-coding genes

Principal-component analysis (PCA) compare protein-coding versus lincRNA TUs based on multiple parameters.

Main features:

- lincRNA TUs \rightarrow upregulation upon exosome knockdown and general lack of polyA
- protein-coding $TUs \rightarrow$ stability within the nucleoplasm and cytoplasm

CONCLUSION:

STILL TO DISCUSS:

LincRNAs appear unlikely to possess sequence-specific functions. Possibly, the act of transcription rather than the nature of the transcript underlies their biological purpose. However, it remains an attractive possibility that tissue-specific RNA-binding proteins (possibly absent in HeLa cells) may selectively restrict lincRNA turnover and so allow their sufficient accumulation to promote functional roles at least for some of these RNAs.

Thanks for your attention!

