

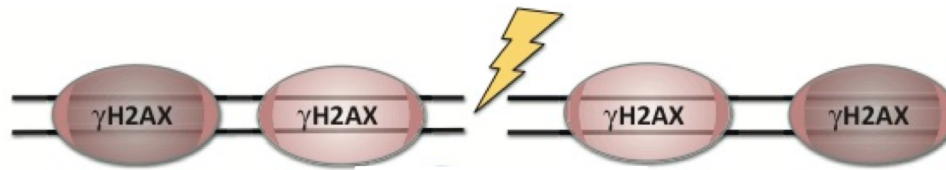
**DDRNAS**

**DNA DAMAGE RESPONSE RNAs**

## **Site-specific DICER and DROSHA RNA products control the DNA-damage response**

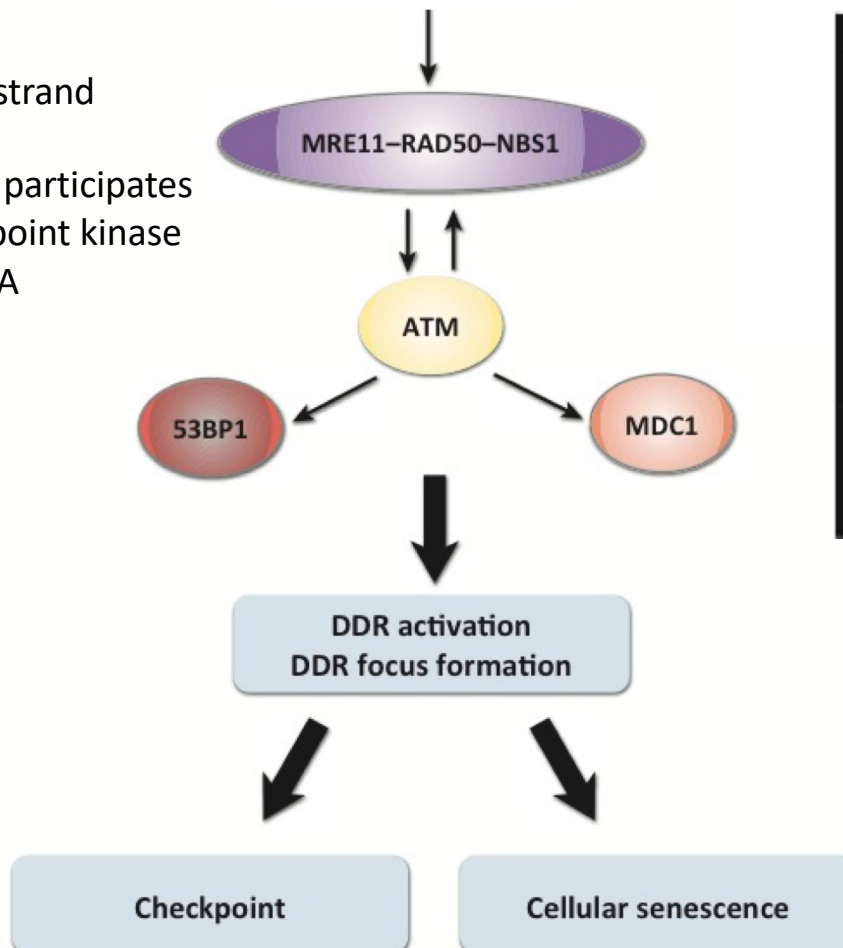
Sofia Francia<sup>1,2</sup>, Flavia Michelini<sup>1</sup>, Alka Saxena<sup>3</sup>, Dave Tang<sup>3</sup>, Michiel de Hoon<sup>3</sup>, Viviana Anelli<sup>1†</sup>, Marina Mione<sup>1†</sup>, Piero Carninci<sup>3</sup> & Fabrizio d'Adda di Fagagna<sup>1,4</sup>

# The DNA damage response revisited

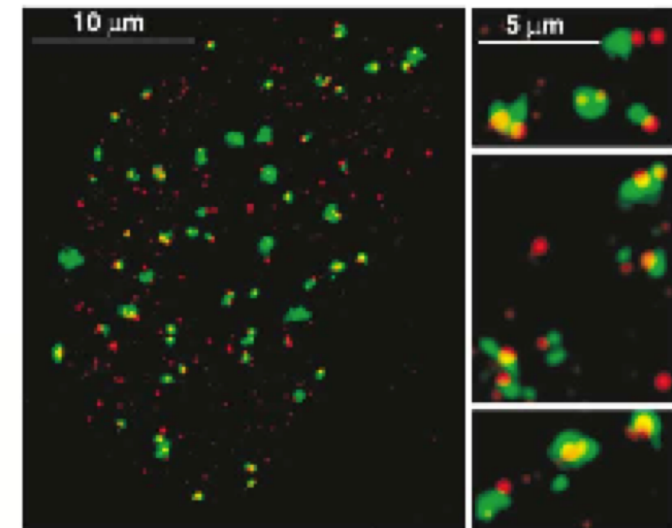


## The MRN complex

- binds avidly to double-strand breaks
- The MRN complex also participates in activating the checkpoint kinase ATM in response to DNA

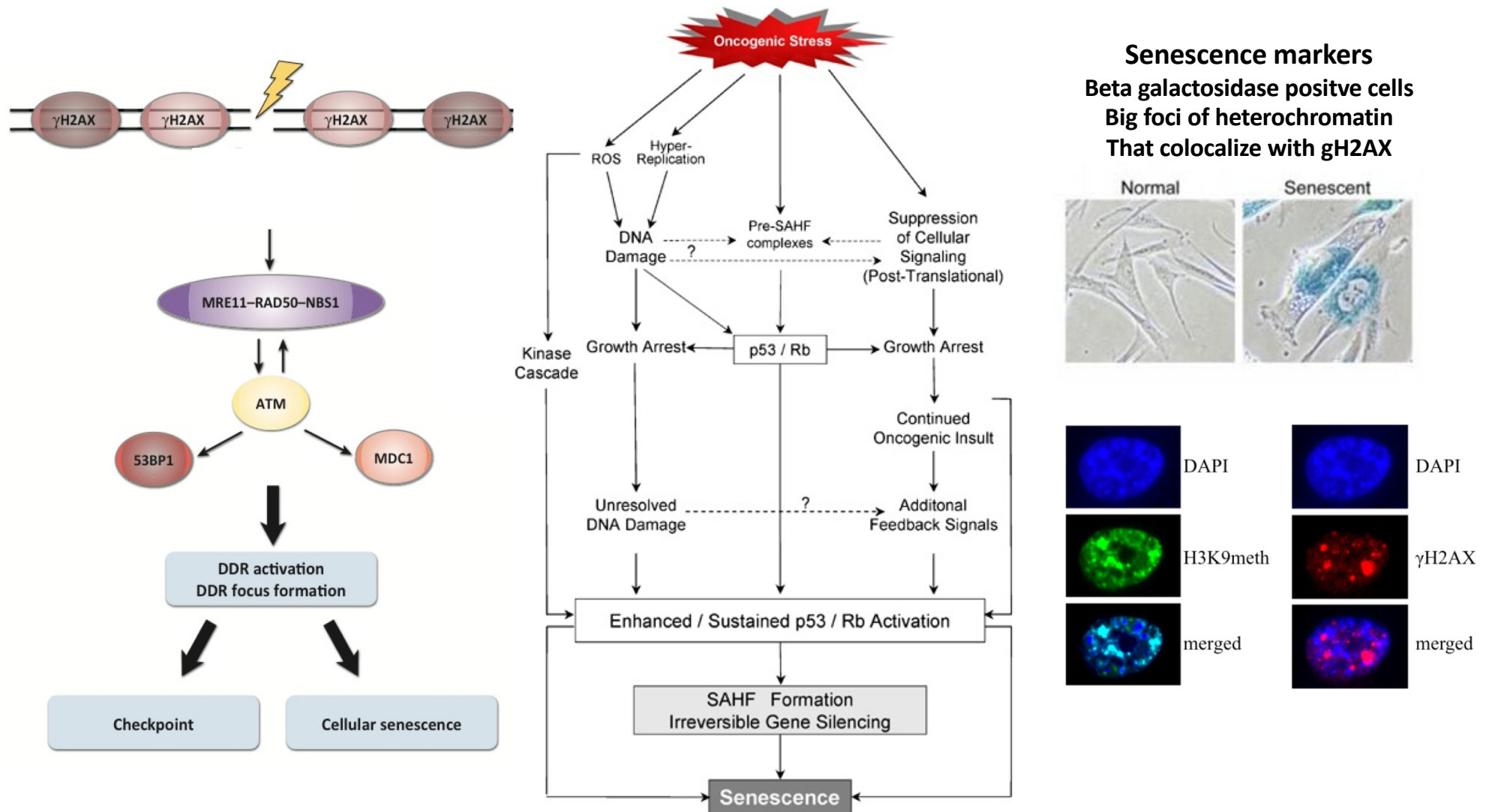


## DNA Damage Foci



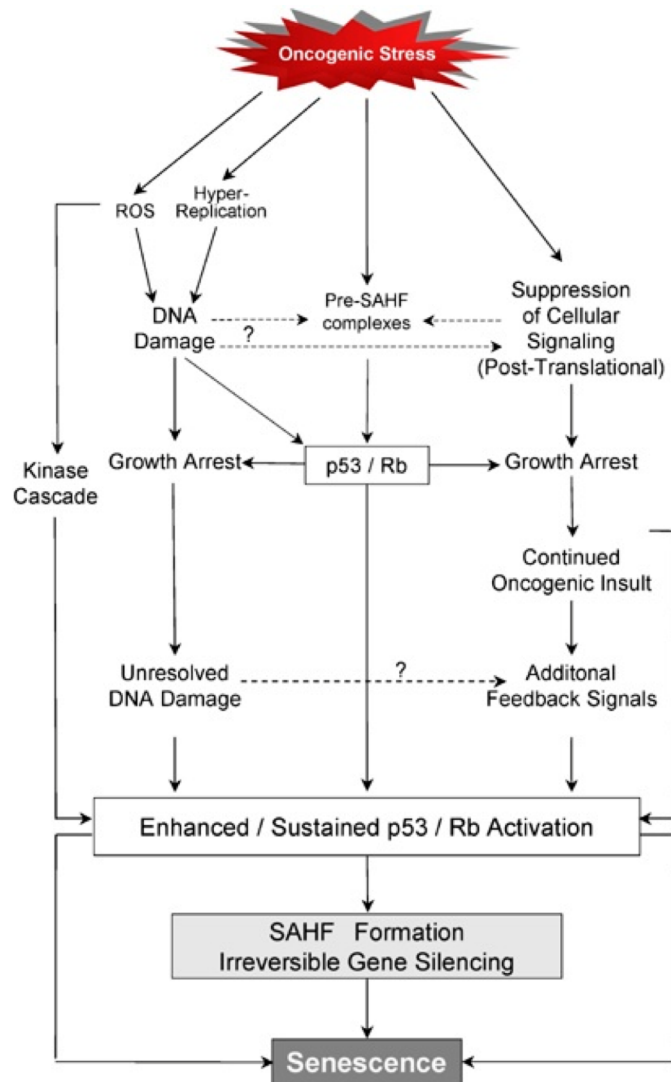
$\gamma$ H2AX  
53BP1  
MRE11  
P-ATM

# Model system for persistent DNA damage: ONCOGENE INDUCED SENEESCENCE



Expression of oncogenes mediates increased DNA damage load  
= tumorsuppressor mechanism  
→ Additional mutations required to escape from tumorsuppression  
→ Cancer formation

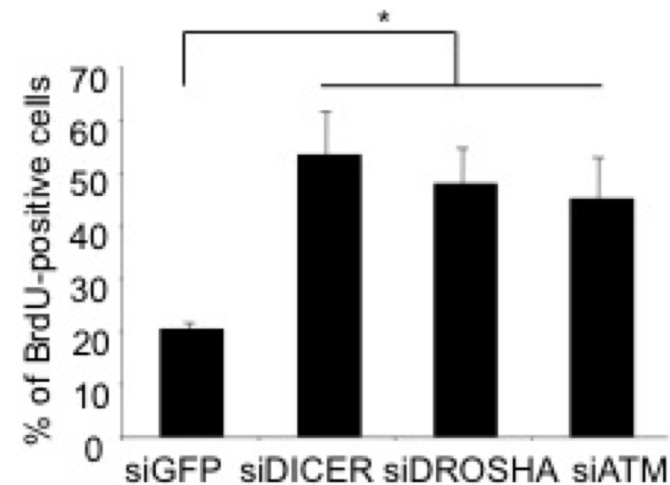
# LOSS OF DICER/DROSHA REDUCES ONCOGENE INDUCES SENESCENCE



Human foreskin fibroblasts (BJ cells) retrovirally transduced with a vector encoding a Ras cDNA containing an oncogenic mutation = **H-RasV12**  
=Oncogene induced senescent cells (“OIS cells”)

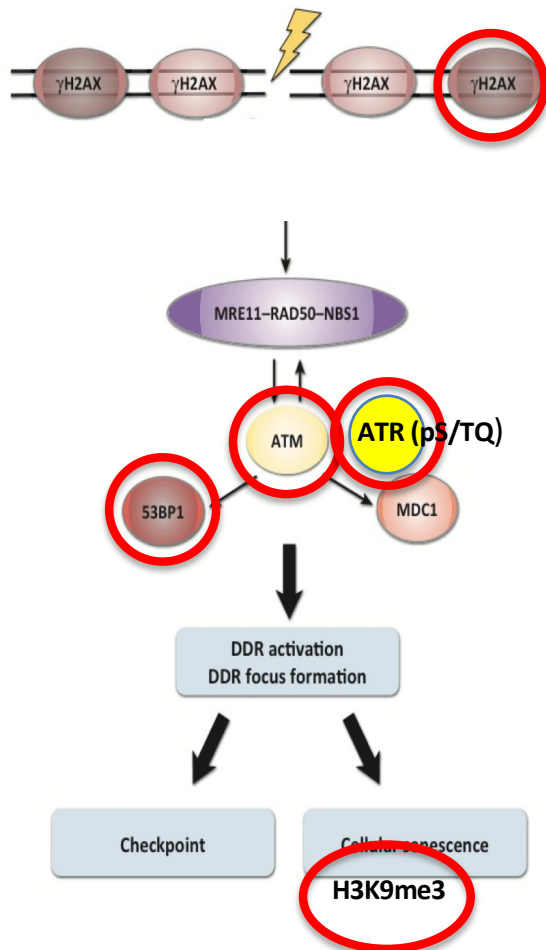
**H-RasV12 drives excessive proliferation →  
Accumulation of DNA damage → Senescence  
→ SAHF**

**OBSERVATION IN THE LABORATORY:**  
Knock-down of DICER and DROSHA  
or ATM increases cell proliferation markers

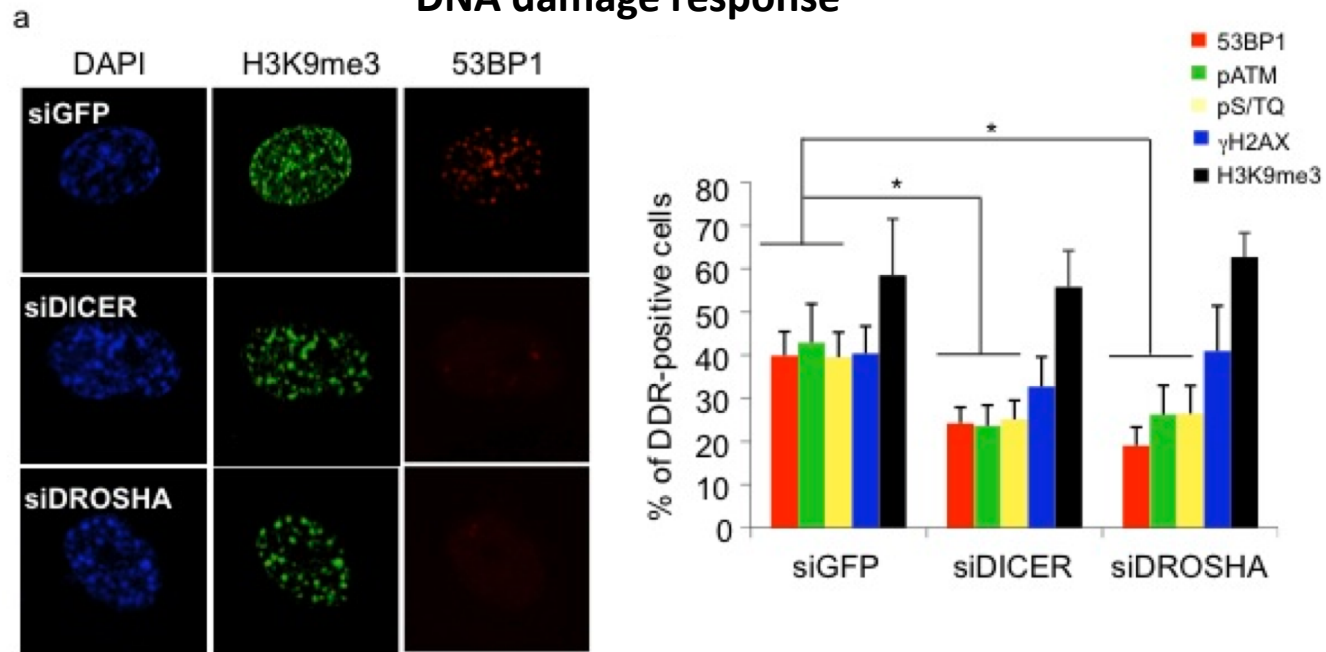


BrdU is incorporated in S-Phase and can be detected  
Using an antibody (IF); more BrdU+ cells = more proliferation

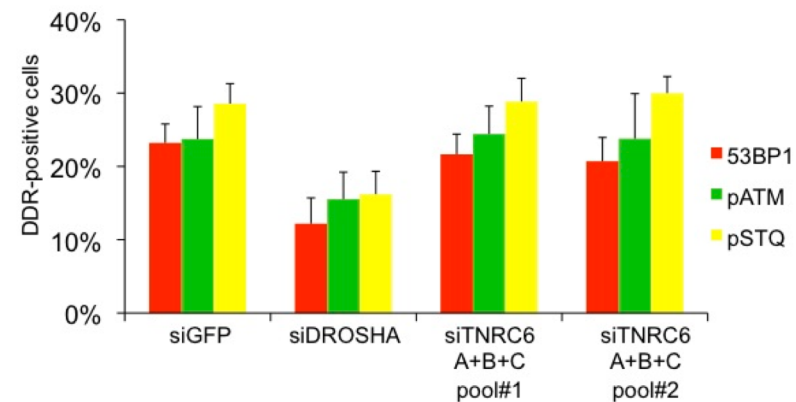
# LOSS OF DICER/DROSHA REDUCES DNA DAMAGE SIGNALLING IN OIS CELLS



## siRNA/miRNA pathways are important for DNA damage response

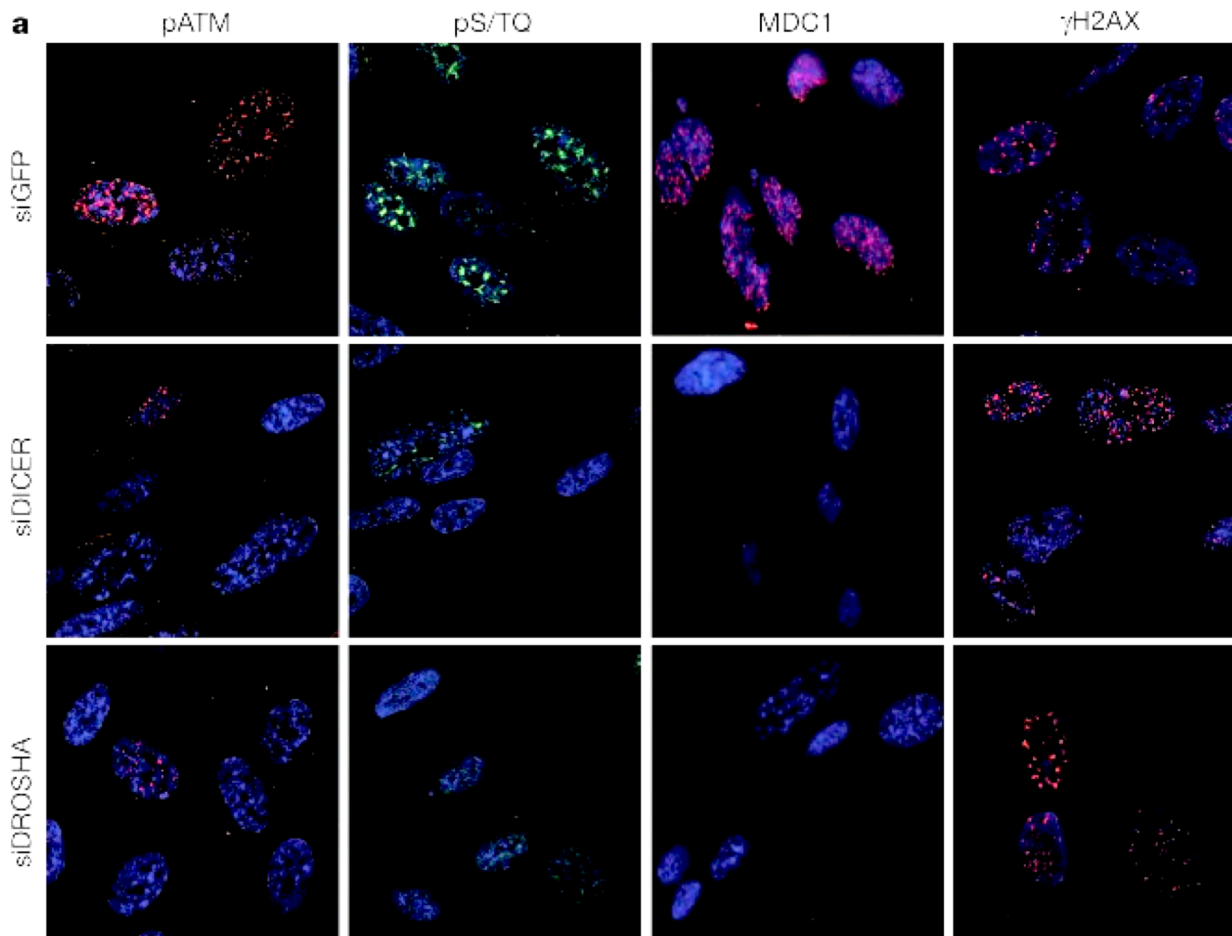


**KNOCK DOWN  
OF GENES IMPORTANT  
FOR miRNA FUNCTION  
(TNRC6A,B,C; translational  
repression)  
do not impact on DNA  
damage response**

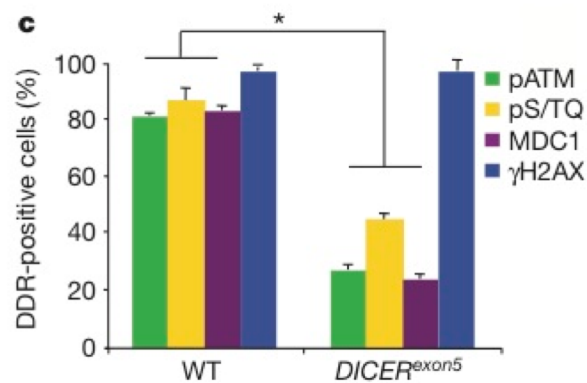
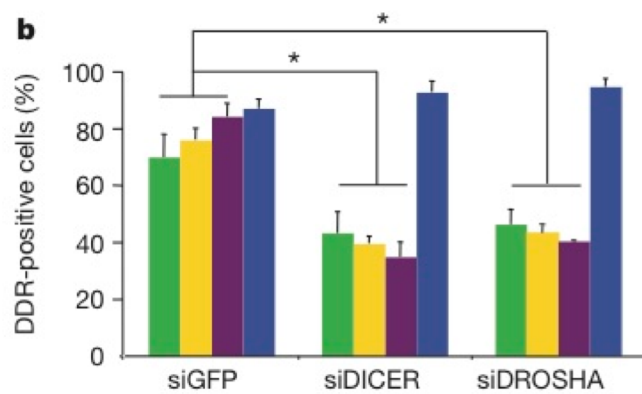
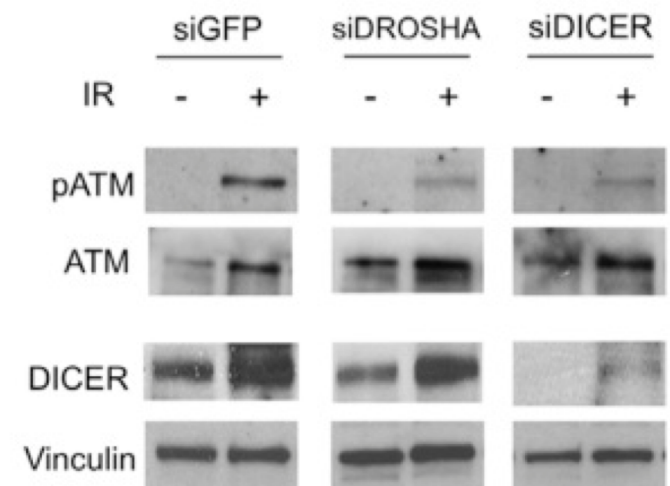


Note: multisite phosphorylation of SQ/TQ motifs is required for normal DNA-damage responses  
**siRNA PATHWAYS ARE INVOLVED IN THE CONTROL OF DNA DAMAGE RESPONSE**

# ANOTHER MODEL: GAMMA IRRADIATION OF NORMAL FIBROBLASTS



**Knock-down of  
DICER and DROSHA  
Impairs the activation  
of a DNA damage response  
In gamma irradiated cells**



# IS RNA REQUIRED TO TRIGGER AN EFFICIENT DNA DAMAGE RESPONSE?

**Model  
Heterochromatin**

Permeabilize  
Cells (detergent)



Treat with  
RNase



Heterochromatin  
Proteins are lost

**Model  
DNA damage**

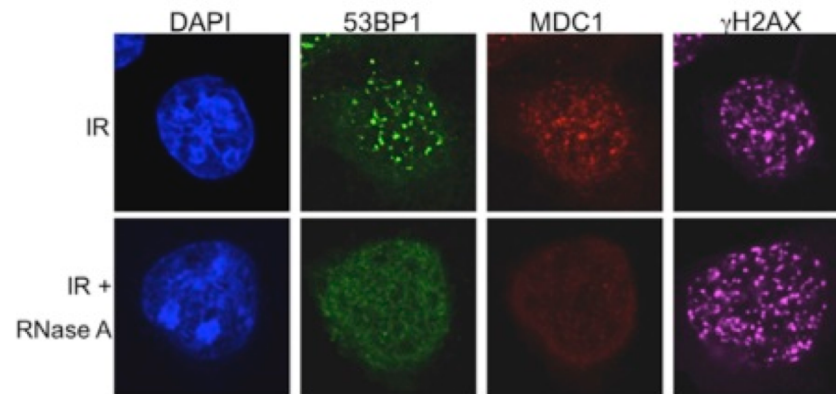
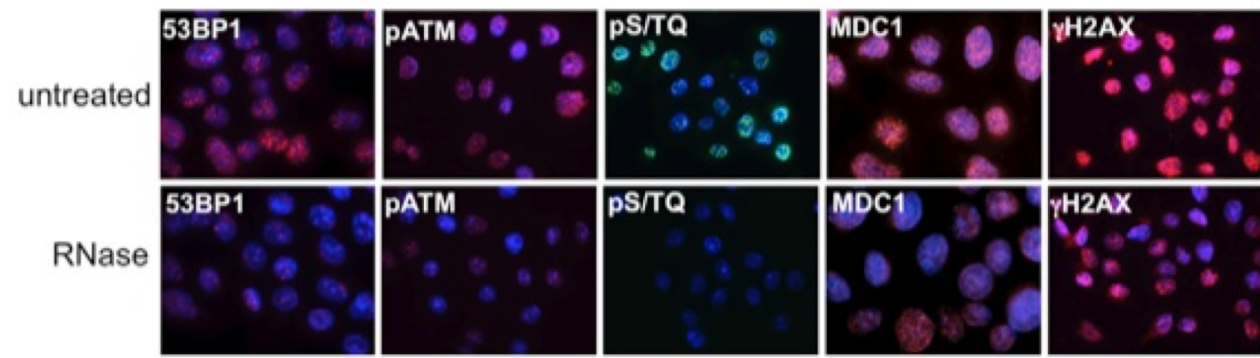
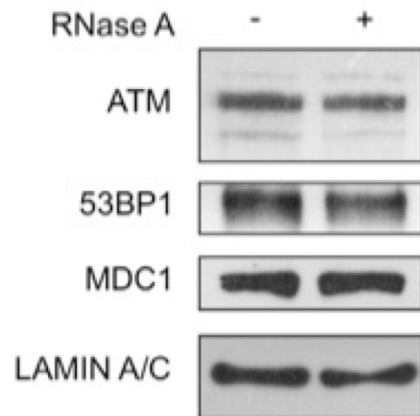
HeLa cells +  
Gamma irradiation +  
permeabilize



Treat with  
RNase



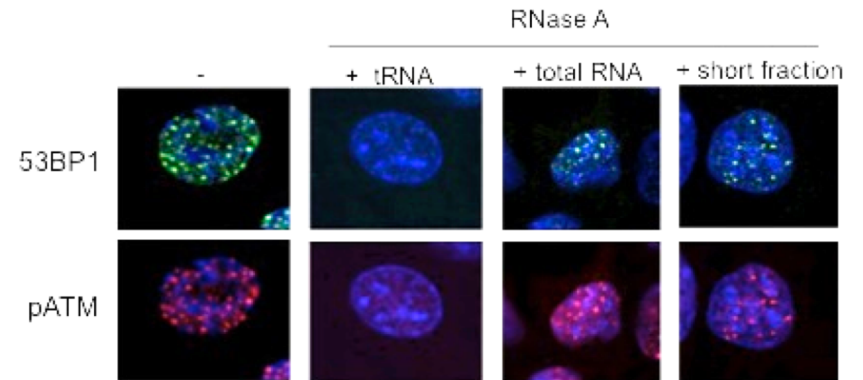
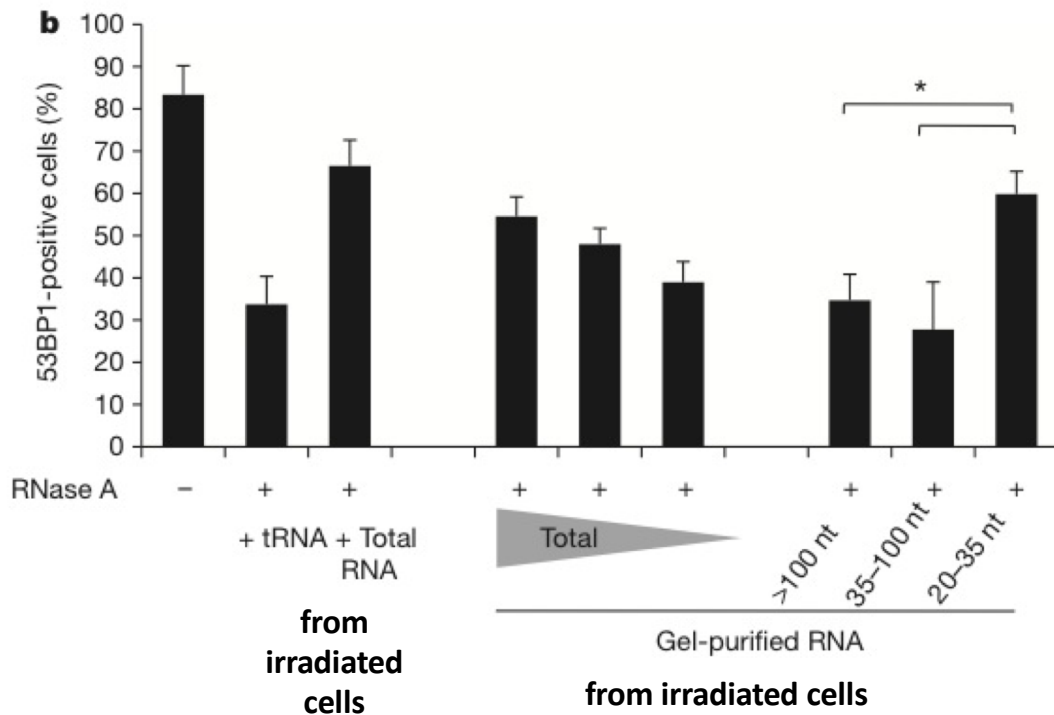
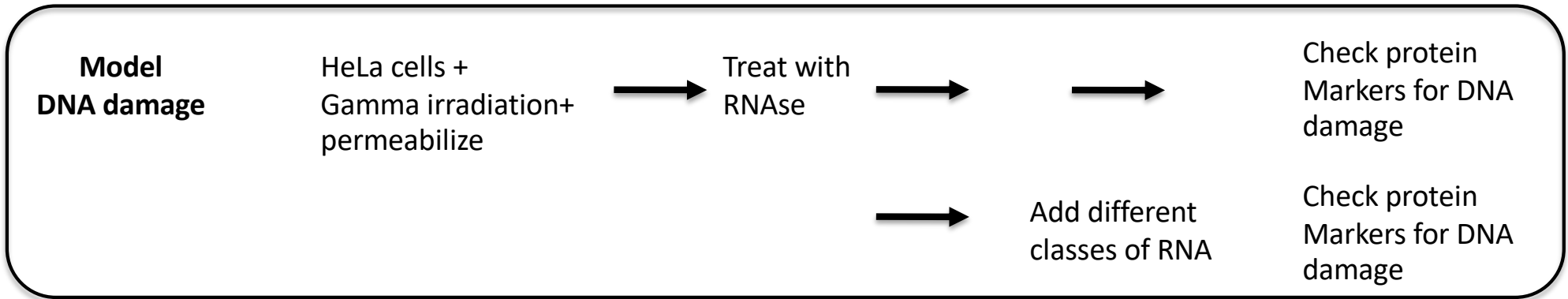
Check protein  
Markers for DNA damage



**RNase treatments reduces the amount of DNA damage markers  
after gamma irradiation**

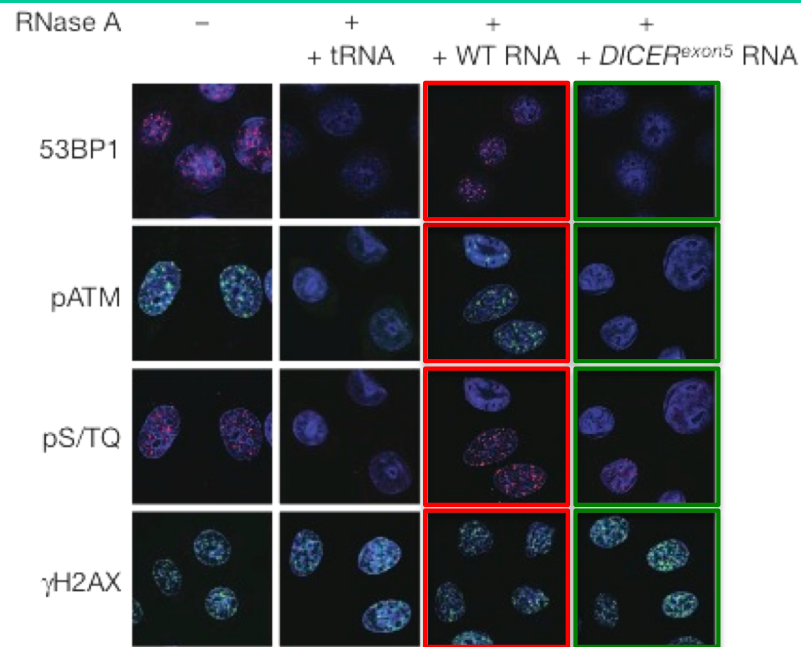


# WHAT KIND OF RNA IS REQUIRED TO TRIGGER AN EFFICIENT DNA DAMAGE RESPONSE?

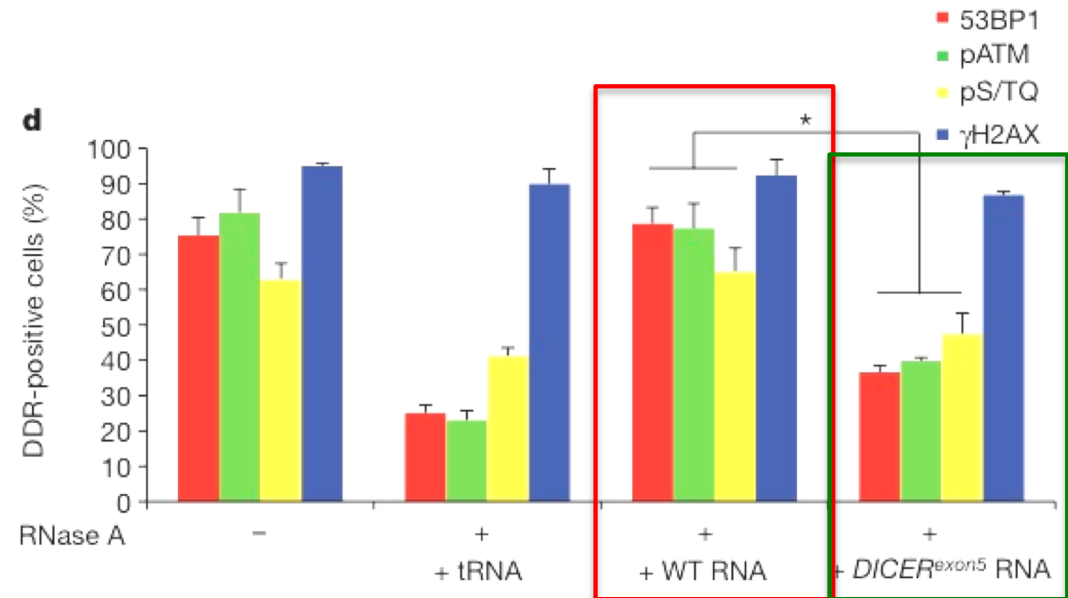


**A short RNA fraction (20-35 nt) rescues DNA damage response after RNase treatment = POTENTIAL DICER/DROSHA PRODUCTS**

# WHAT KIND OF RNA IS REQUIRED TO TRIGGER AN EFFICIENT DNA DAMAGE RESPONSE?



RNA From Irradiated Dicer wt Cells = Small RNAs present	RNA From Irradiated Dicer KO Cells = NO small RNAs
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Total RNA from Dicer null cells cannot rescue defects of DNA damage foci formation after RNase treatment

→ **Dicer has a critical role in DNA damage response**

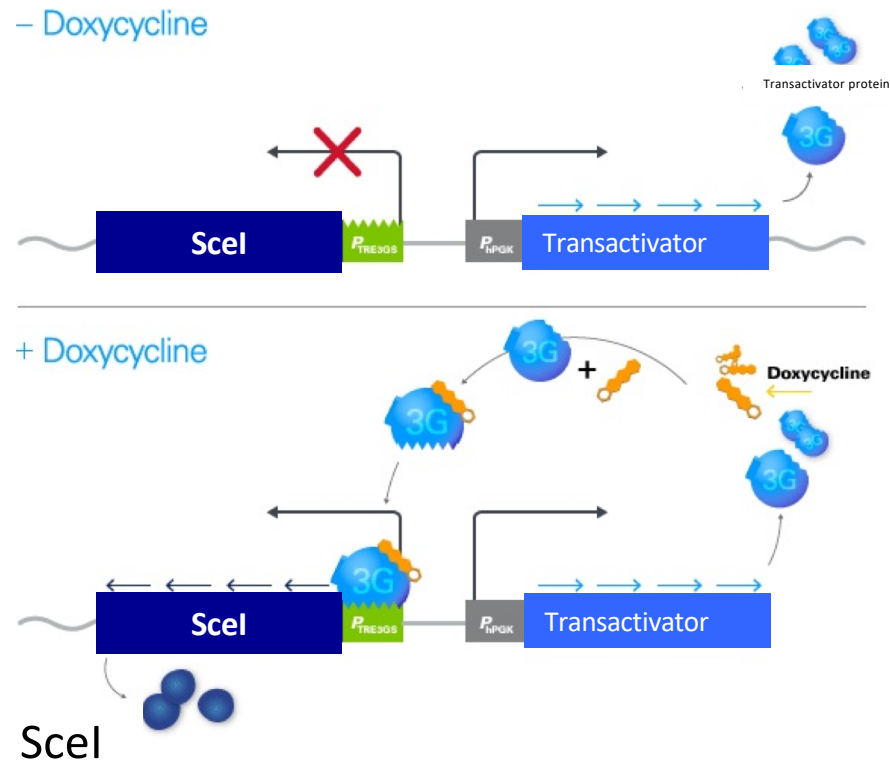
**CAN SMALL RNAs = DDRNAs (DNA DAMAGE RESPONSE RNAs) ACT DURING DNA DAMAGE RESPONSE AT A DEFINED SITE IN THE GENOME**

# A MODEL SYSTEM TO STUDY THE KINETICS OF DNA DAMAGE

## Cell line:

Contains

1. An inducible transactivator
2. the restriction enzyme SclI under the control of a inducible promoter
3. A SclI site between Lac Repressor DNA sequences
4. The Lac Repressor that binds DNA sequences around the SclI sites



SclI

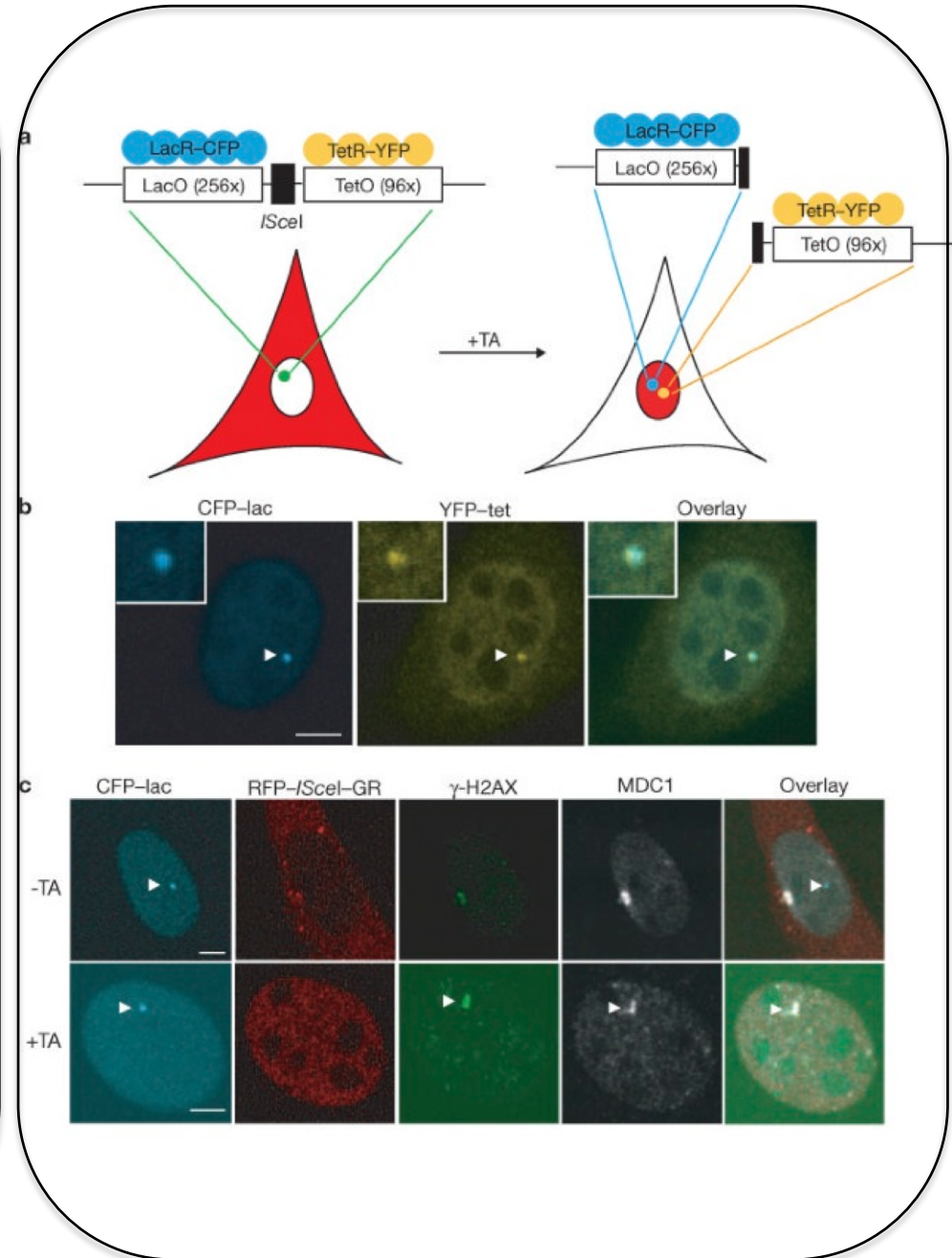
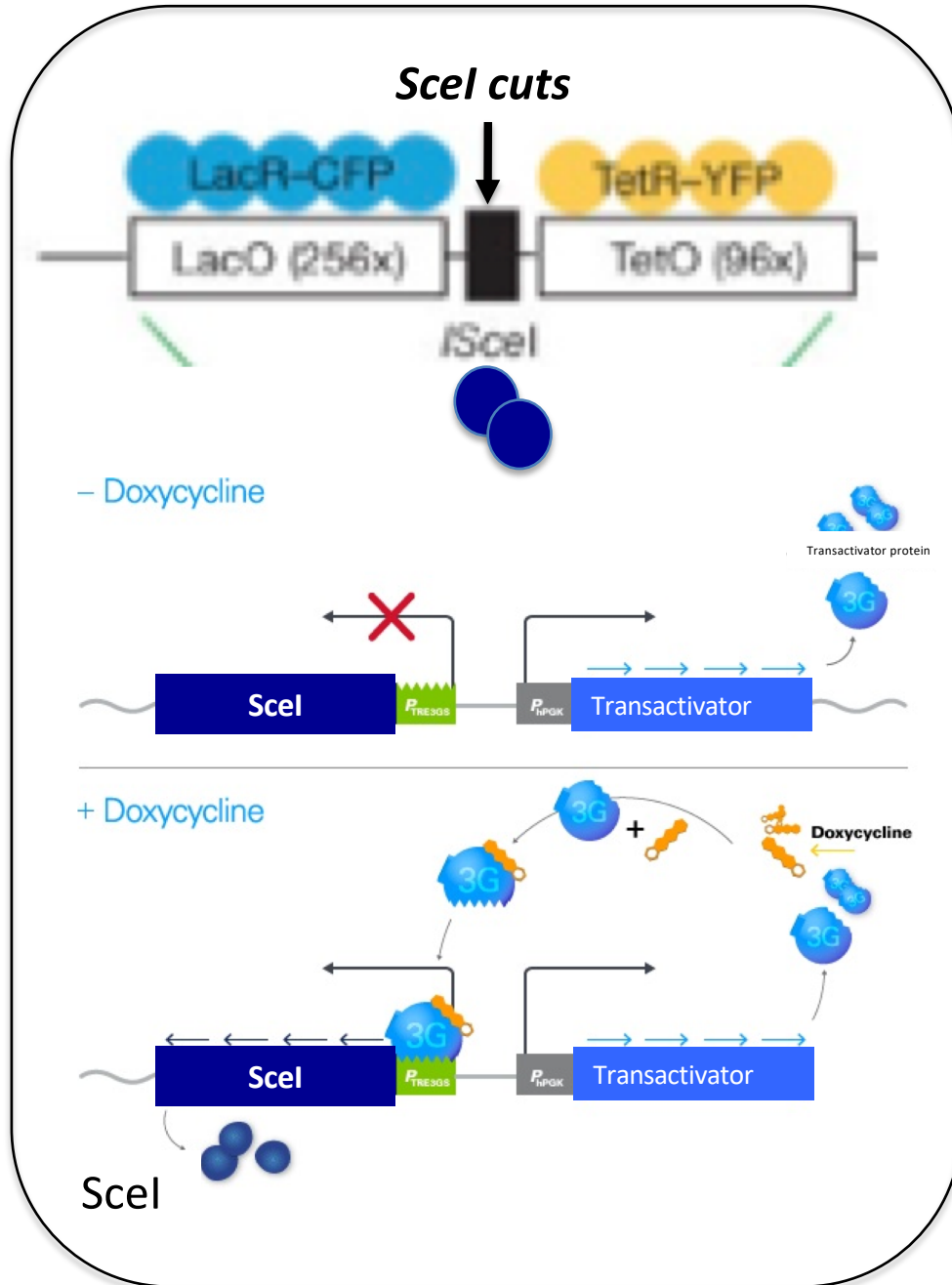
5'...TAGGGATAACAGGGTAAT...3'  
3'...ATCCCTATTGTCCCATTA...5'

SclI is a restriction enzyme  
That does not cut in the human  
genome

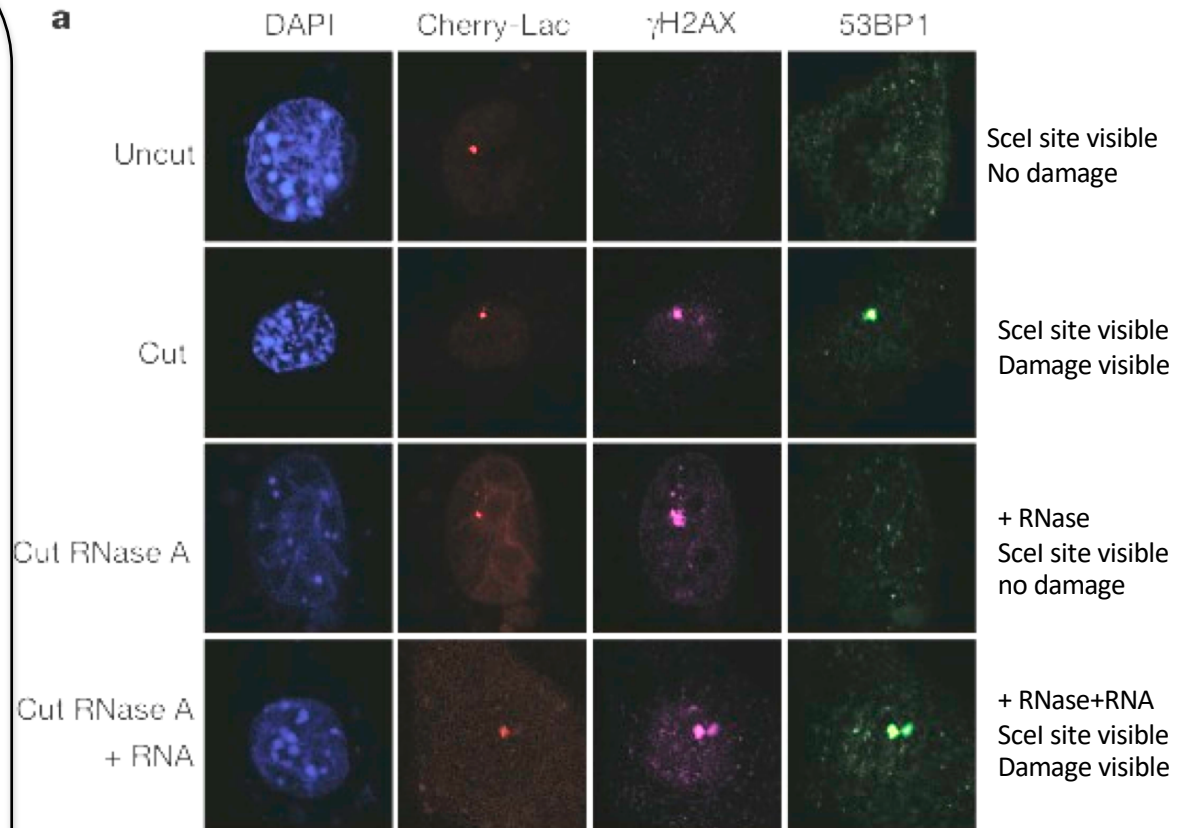
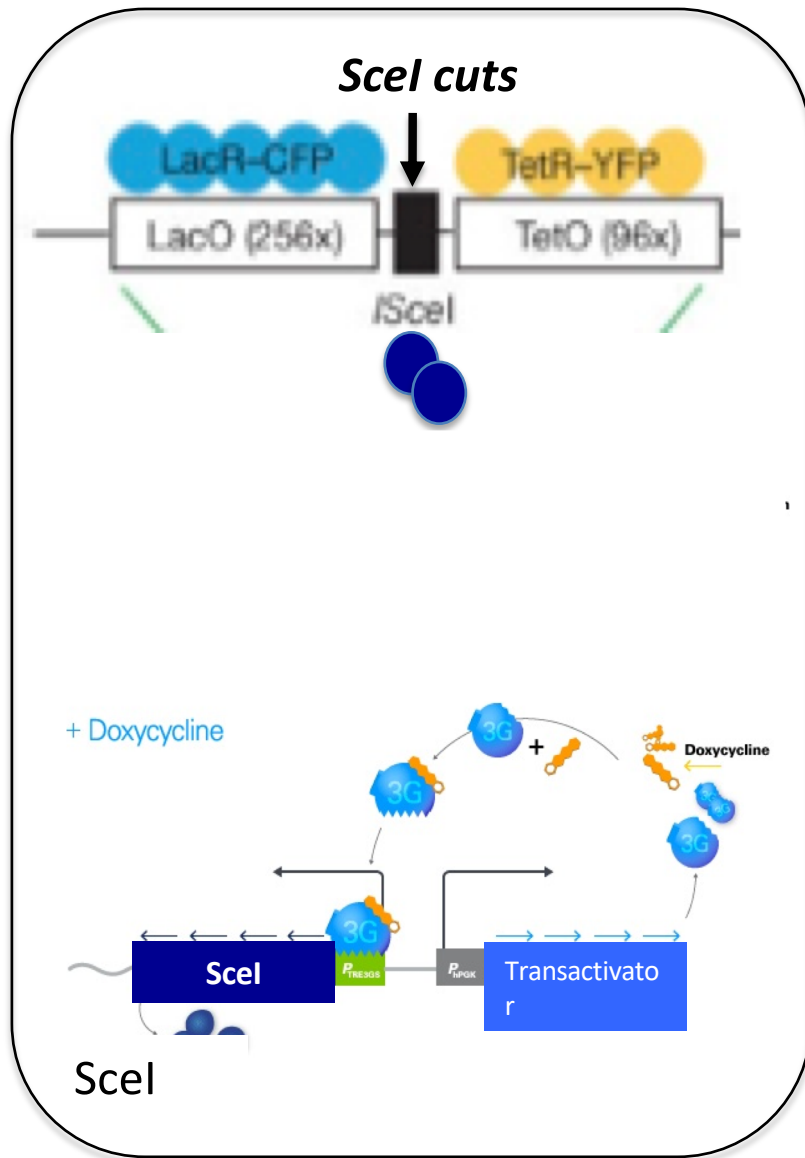
**Inducing SclI expression does not  
cut genomic DNA in human cells!**

**LET'S INTRODUCE A SclI SITE  
AND MARK THE SclI SITE USING  
SEQUENCES BOUND BY THE  
Lac REPRESSOR**

# A MODEL SYSTEM TO STUDY THE KINETICS OF DNA DAMAGE



# DEFINED RNAs FROM DNA DAMAGE SITES ARE IMPORTANT FOR DNA DAMAGE RESPONSE



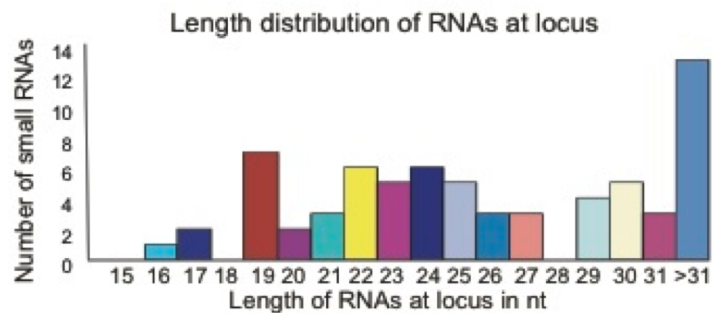
→ RNA was prepared from cells that have cut the *Scel* site  
 → RNA from cells that do not have the *Scel* site but express *Scel* cannot rescue!!!!

# DEFINED RNAs FROM DNA DAMAGE SITES ARE IMPORTANT FOR DNA DAMAGE RESPONSE - EVIDENCE 1

Take Scel cells:

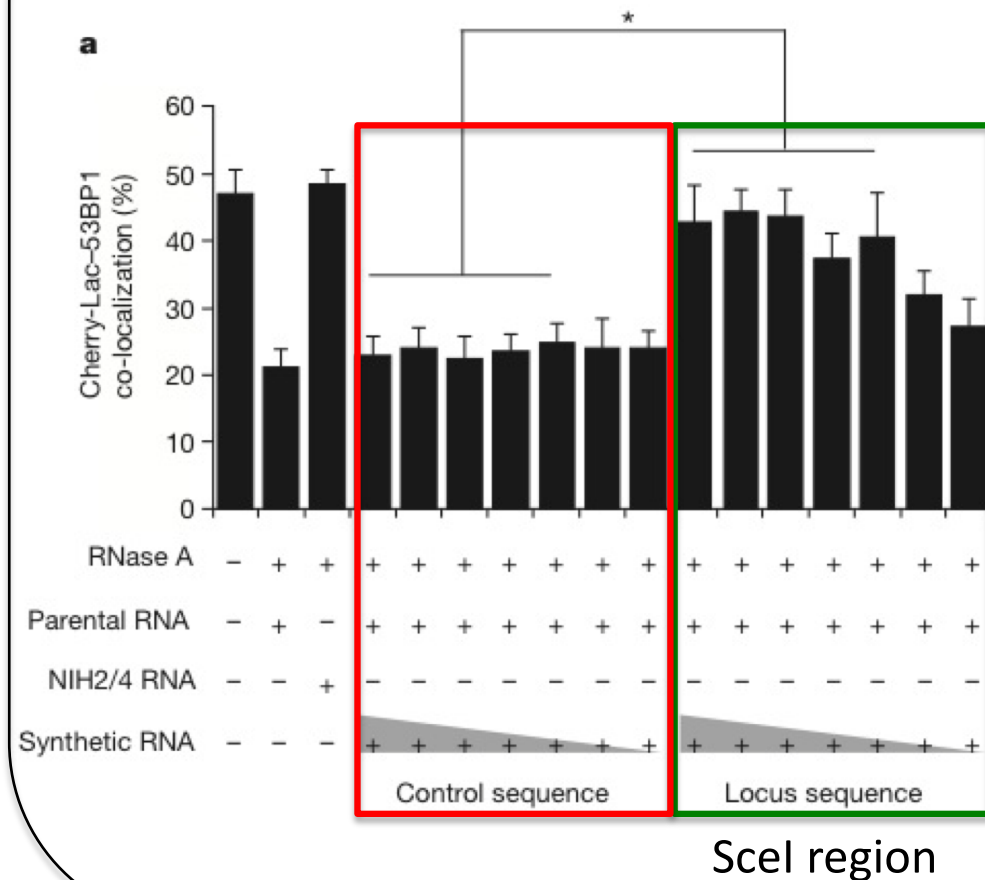
- induce cutting
- prepare smallRNA fraction
- make RNASeq

detect small RNAs from locations around the Scel sites

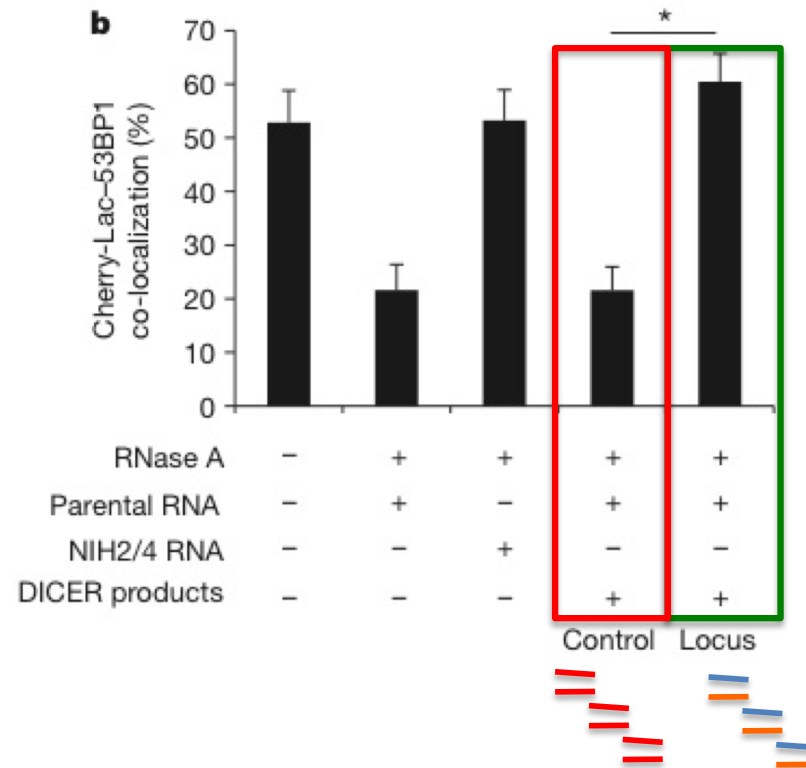
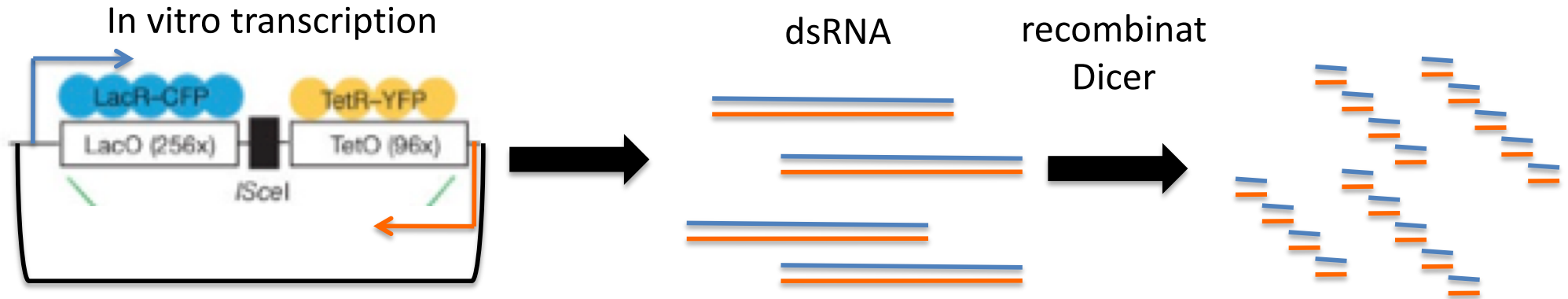


!!!!Small RNAs (16-31) arising from regions around the Scel site can be identified !!!!

Small RNAs with sequences that resemble the region around the Scel site can rescue the formation of DNA damage foci at the Scel site



# DEFINED RNAs FROM DNA DAMAGE SITES ARE IMPORTANT FOR DNA DAMAGE RESPONSE - EVIDENCE 2



# DEFINED RNAs FROM DNA DAMAGE SITES ARE IMPORTANT FOR DNA DAMAGE RESPONSE - EVIDENCE 2

Take Scel cells

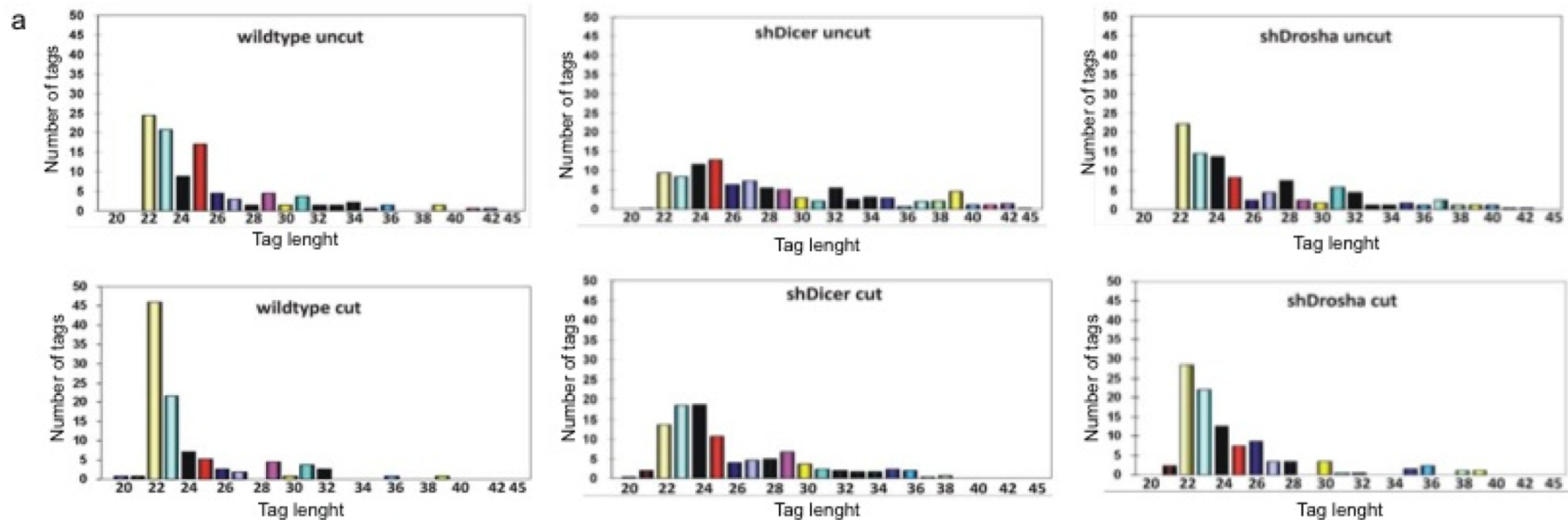
- **Knock down DICER or DROSHA:**

- induce cutting

- prepare RNA

- make RNASeq

detect small RNAs from locations around the Scel sites



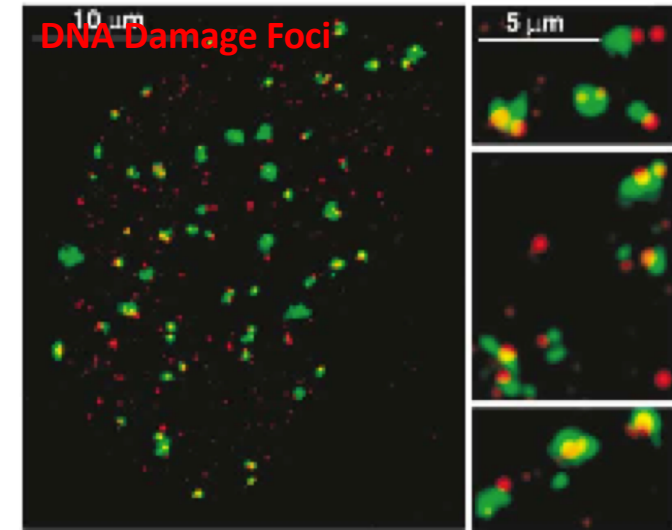
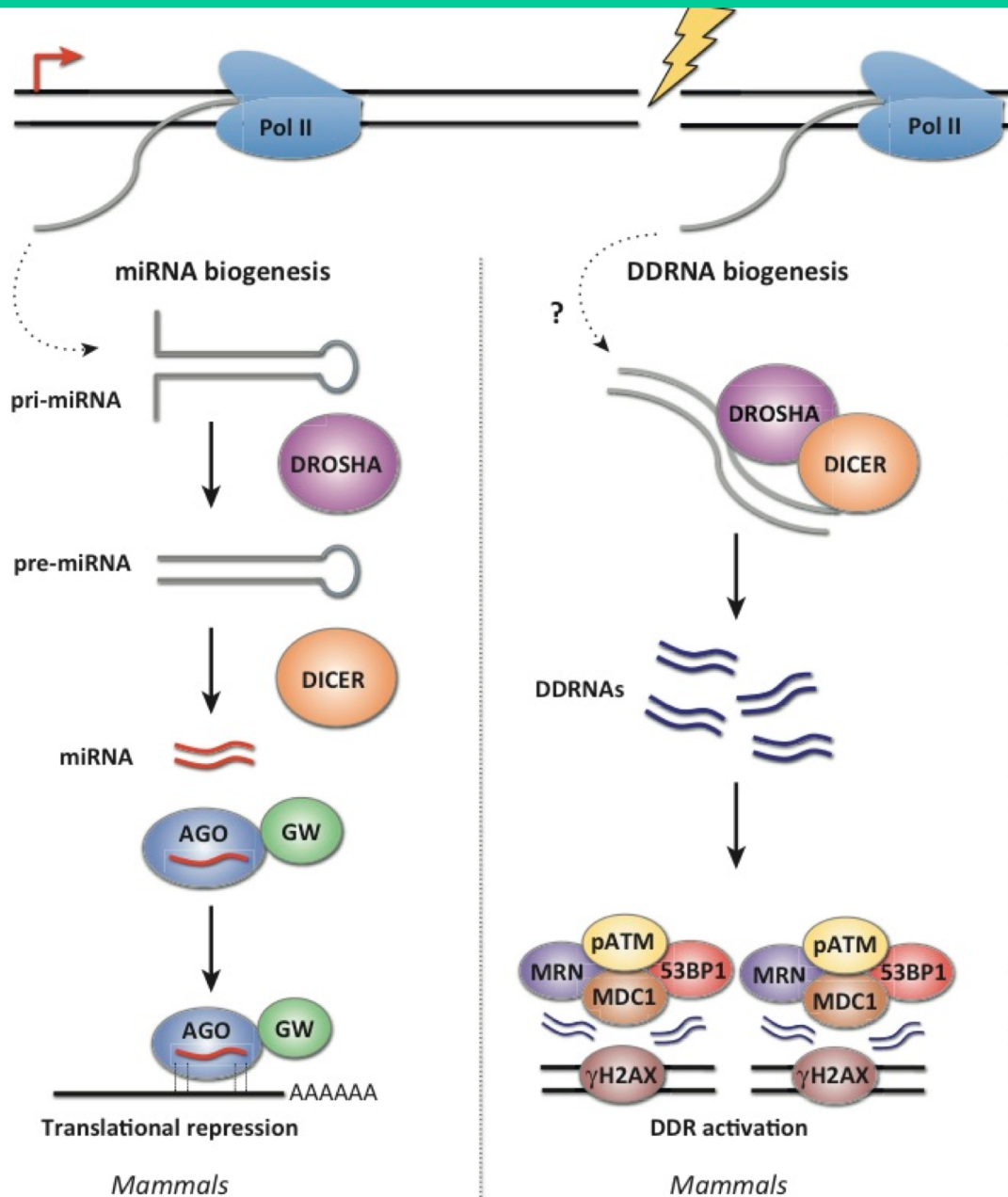
*DICER RNAi*

*DROSHA RNAi*

Small RNAs (16-31) arising from regions around the Scel site is reduced in DICER/DROSHA knock-down cells



# DNA DAMAGE RESPONSE RNAs (DDRNA) CONTROL THE DNA DAMAGE RESPONSE



$\gamma$ H2AX  
 53BP1  
 MRE11  
 P-ATM

In summary, we demonstrate that different sources of DNA damage, including oncogenic stress, ionizing radiation and site-specific endonucleases, activate the DDR in a manner dependent on DDRNAs, which are DICER- and DROSHA-dependent RNA products with the sequence of the damaged site. DDRNAs control DDR foci formation and maintenance, checkpoint enforcement and cellular senescence in cultured human and mouse cells and in different cell types in living zebrafish larvae. They act differently from canonical miRNAs, as inferred by their demonstrated biological activity independent of other RNAs and of GW182-like proteins.