

Course:

Remote Sensing of Global Changes

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Part 3

Methodologies for the interpretation and processing of remotely sensed images

SCIENCE OF REMOTE SENSING



Digital volume – Radiometric resolution

How much digital volume is occupied according to radiometric resolution?

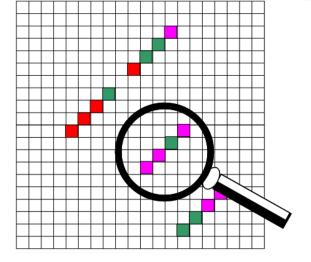
Example:

Image 1024x768 pixel RGB 24-bit, meaning that each colour (RGB) could be represented by 8 bits

How many MB are occupied by the image?

1024x768x3 = 2359296 = 2.25 MB

	1 byte = 8 bits
	1 kilobyte = 1024 bytes
1	. megabyte = 1024 kilobytes
	gigabyte = 1024 megabytes
	1 terabyte = 1024 gigabytes



255	87	0
0	0	255
0	130	0
87	255	255
0	255	255
130	255	255

Digital volume – Radiometric resolution

You try:

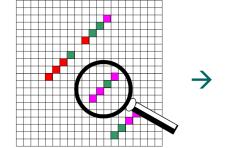
100x100 km tile, how many pixels? 100000x100000 pixels

For each band, how many MB, with 8, 16 and 24 bit?

8bit = 100000x100000x1 = 100000000000 bytes = 9.31 GB

16bit = 100000x100000x2 = 200000000000 bytes = 18.62 GB

24bit = 100000x100000x3 = 3000000000 bytes = 27.93 GB



255	87	0
0	0	255
0	130	0
87	255	255
0	255	255
130	255	255

Digital volume of multiband image = n total pixels + n bit to represent a pixel + palette dimensions (for RGB, $3x2^n$)

RGB 8bit = 3x256 = 768 bytes RGB 16bit = 3x65546 = 196608 bytes = 0.18 MB

Image statistics and analysis

Like any numeric variable, basic statistical parameters:

1. Mean

$$\overline{DN_k} = \frac{\sum_{i=1,n} DN_{i,k}}{n_k}$$
 Target band N pixel in image

2. Standard deviation

$$SD_k = \sqrt{\frac{\sum_{i=1,n} (DL_{i,k} - \overline{DL_k})^2}{(n_k - 1)}}$$

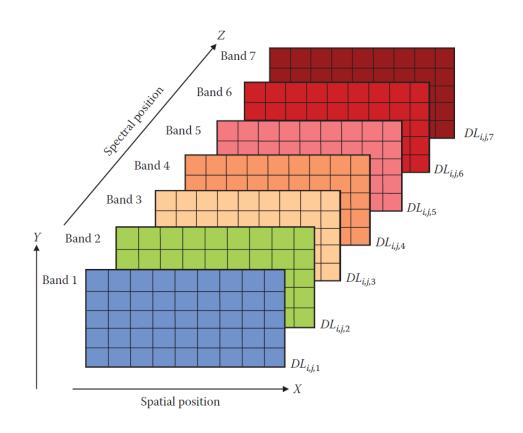
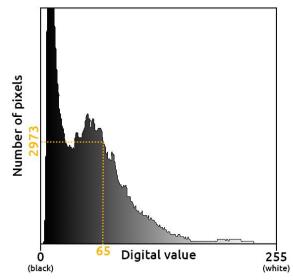
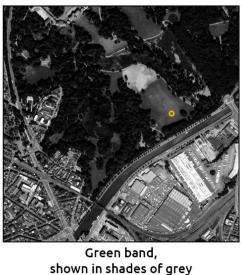


Image statistics and analysis





describes the distribution of DNs in the image.

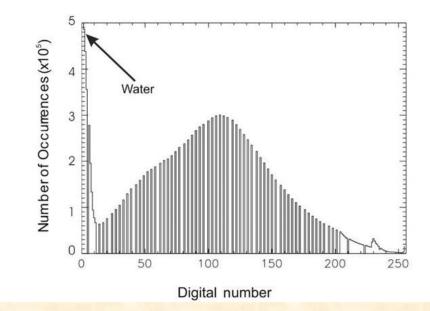
It is possible to compute the **histogram** for each band, which

https://eo.belspo.be/en/remote-sensing-images

$$RF(DN_i) = \frac{F(DN_i)}{\sum_{t=1,m} DN_i}$$

The <u>histogram</u> provides a first evaluation of the data distribution of each band.

The center indicates the dominant brightness of each band (mean), while its width is related to band variability (SD).



The process of image (or contrast) enhancement attempts to **adjust the** radiometric resolution of the image to the capabilities of the display system.

Two possible situations:

1. Range of DNs in the image is lower than the range of brightness display values



2. Image has a higher number of DNs than can be visualized.



Definitions of contrast:

$$C_{1} = \frac{DN_{max}}{DN_{min}}$$

$$C_{2} = DN_{max} - DN_{min}$$

$$C_{3} = SD_{DN}$$

Stretching operations should provide higher contrast, increasing any of the three measures.

Colour compression implies maintaining the visual quality of the colour images with a significant reduction of the file size.

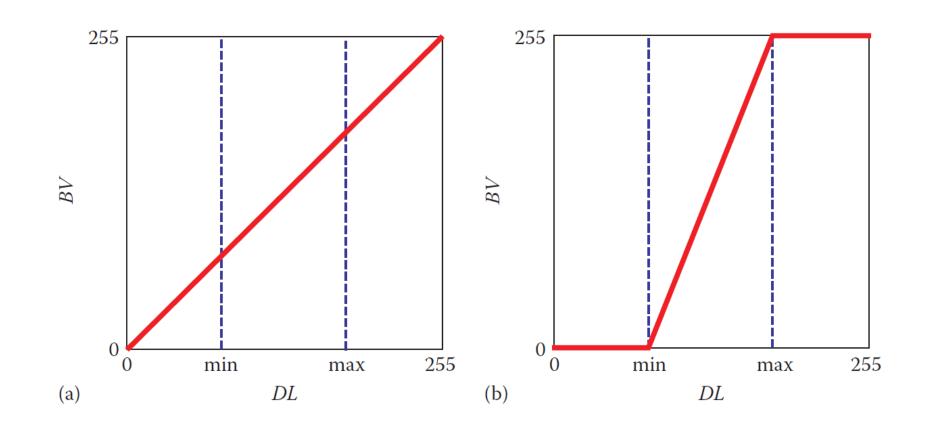
<u>Lookup table (LUT):</u> is a numeric matrix that shoes the brightness values (BVs) on the screen that will be assigned to each DN in the image.

The order of the row indicates the input DN

The value in the table expresses the BV that will display that DN. In this case, the higher the BV is, the more R, G and B the pixel will be.

BV (blue)	BV (green)	$BV({f red})$	
1	1	1	
2	2	2	
3	3	3	
•••	•••	•••	
255	255	255	

Lookup table (LUT) can be also represented as bivariate graph, where the DN of the image are displayed in the x-axis and BV is represented on y-axis.



CONTRAST ENHANCEMENT:

Applied when the range of DNs in the image is lower than the range of brightness display values.

Used adjusting the original DN distribution to the capabilities of the display system, improving image contrast.

Usually, even if a sensor has a 8-bit radiometric resolution, in most images the original data distribution do not cover the entire range of available BVs. In a monoband image, this means that a large range of grey tones are not used, and the image has a low contrast.

Three different ways:

- 1. <u>Linear Contrast Enhancement</u> (or Linear Stretch)
- 2. <u>Histogram Equalization</u>
- 3. <u>Special Contrast Stretch</u>

<u>Linear Contrast Enhancement</u> (or Linear Stretch)

The raw histogram in Fig (b) shows poor contrast.

Fig (d) is the stretched image, and displays higher contrast. Look at the stretched histogram in Fig (c).

In this case, linear transformation:

$$BV = b + g DN$$

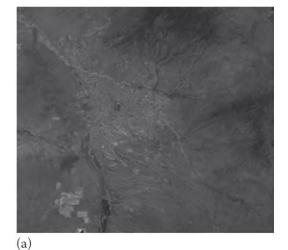
b = bias (intercept on y-axis)

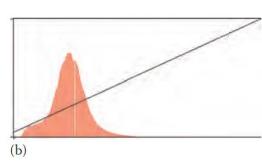
g = gain (slope)

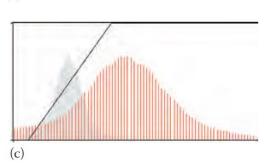
$$0 = b + g DN_{min}$$
$$255 = b + g DN_{max}$$

$$g = \frac{255}{DN_{max} - DN_{min}} \qquad b =$$

$$b = \frac{-255*DN_{min}}{DN_{max}-DN_{min}}$$

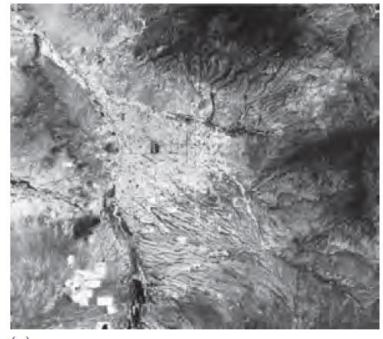




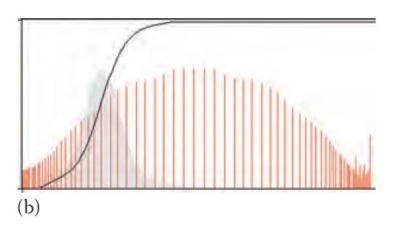




Instead of using DN_{max} and DN_{min} , the user can select different thresholds to fit the equation (e.g. certain percentiles).



(a)



<u>Histogram equalization</u>

The values are stretched according to their actual frequency distribution.

DNs with higher number of pixels will occupy a proportionally larger range in the enhanced image than those with lower frequency.

Fig (b) is the stretched histogram. The graphical profile of the LUT is not a straight line but a curved line, similar to cumulative distribution function of the original DNs.

Special Contrast Stretch

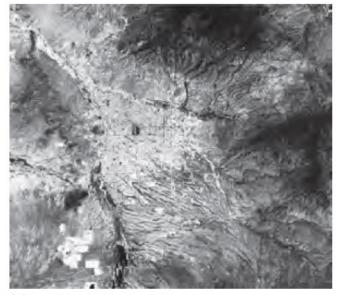
The idea is to restrict the contrast to a specific range of the DN

This could be helpful to enhane a specific feature on Earth's surface.

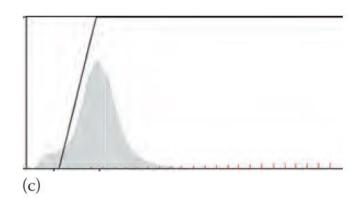
It is based either on:

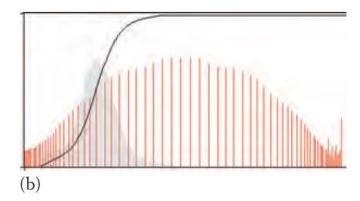
- Setting a min and a max considering the DNs range of a selected feature
- By restricting the histogram equalization into a certain range

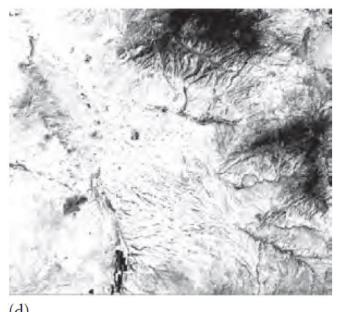
The feature of interest is enhanced at the expense of other areas in the image.











Special case of Special Contrast Stretch is the creation of a binary image (b/w image)

1 = white = pixel related to the feature of interest

0 = black = other pixels

Step 5: Sample the SAR image

Step 6: Construct histogram and calulate threshold by Otsu

Step 7: Otsu threshold binarize

CONTRAST COMPRESSION

CONTRAST COMPRESSION:

Applied when the range of the sensor exceeds the number of available BVs.

It consists in the compression of original DNs range.

Most common approach:

Reducing the DNs range to a number of reduced intervals selected from the histogram.

Might be:

- Intervals of equal width
- Intervals of equal frequency
- Intervals manually selected

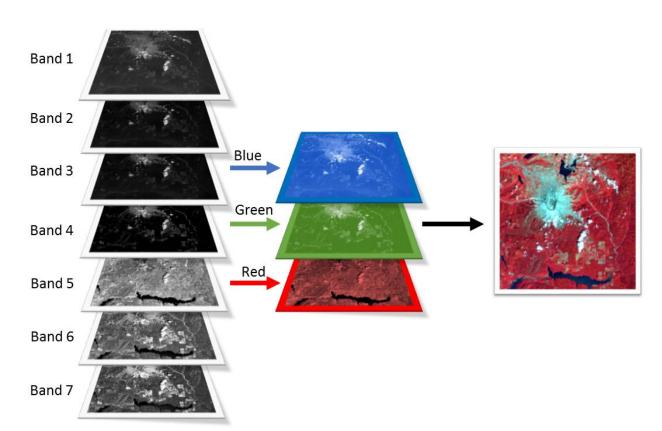
Or more elaborated algorithms exists:

- LZW
- JPEG (accepts 24 bits, 20:1 compression)
- PNG (24 bits but lower compression)





COLOR COMPOSITES



From multispectral data, it is possible to display RGB images, assigning one band to each colour.

In total three bands can be chosen and assigned to R, G and B colour respectively.

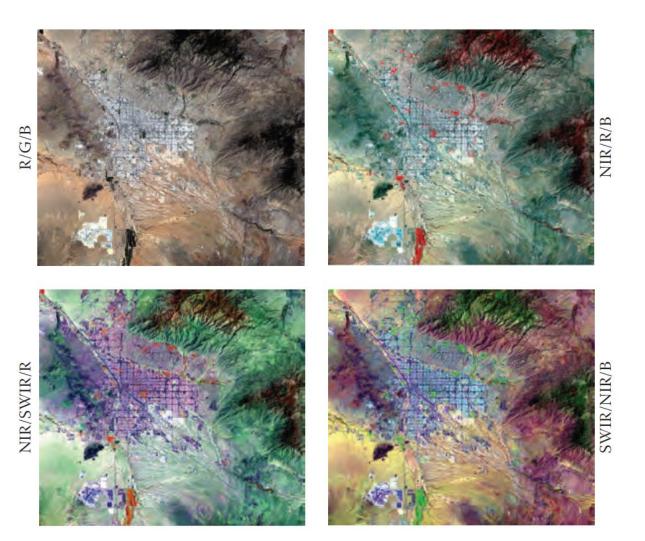
The most common composite is

<u>FALSE COLOR</u> or INFRARED COLOR COMPOSITE: NIR bands are assigned to R, red bands are assigned to G and green bands are assigned to B.

This is used to map vegetation, water bodies, urban areas, ecc.

Of course, any combination is possible.

COLOR COMPOSITES

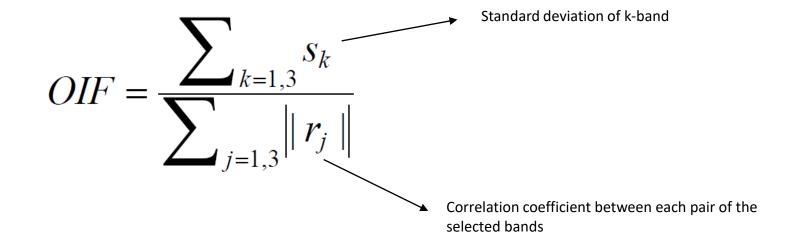


	Landsat 8-9	Sentinel-2
Natural colours	4,3,2	4,3,2
False Colour Infrared	5,4,3	8,4,3
Land/Water	5,6,4	11,8,4
Vegetation analysis	6,5,4	11,8A,5
False colour (urban)	7,6,4	12,11,4
Agriculture	6,5,2	11,8A,2
False c. (TIR, SWIR, Green)	10,7,3	

COLOR COMPOSITES

It is possible to apply quantitative criteria to choose the best band combination for colour composites

OIF = Optimum Index Factor. It evaluates the variances within each bands and the correlations between bands



PSEUDOCOLORS

Use of color even with single band is available.

Since only one band is being used, we use the term pseudocolor.

It is possible to display pseudocolors by creating a LUT where a single DN is represented in different values of RGB.

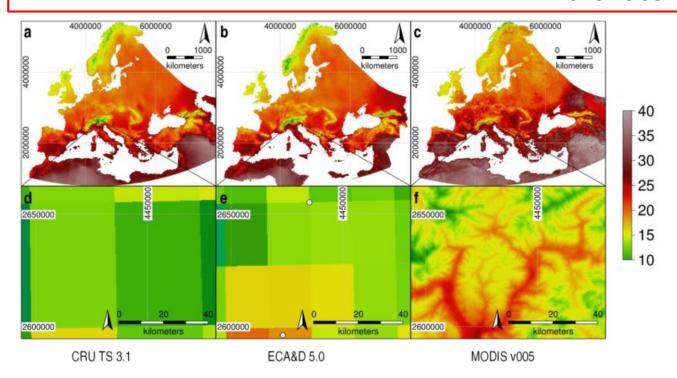
Pseudocolors are generally used:

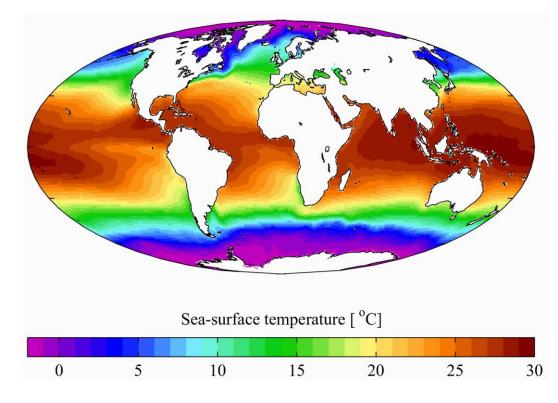
- When a color scheme is needed for a classified image
- To enhance a single-band image, by substituting the gray levels with color tones.

TABLE 6.3
Example of a Lookup Table for a Classified Image

DL	BV Red	BV Green	BV Blue	Resulting Color
0	0	0	0	Black
1	255	0	0	Red
2	0	255	0	Green
3	0	0	255	Blue
4	0	255	255	Cyan
5	255	255	0	Yellow
6	255	127	0	Ochre
7	127	127	127	Gray
8	255	0	127	Pink

PSEUDOCOLORS





Part 3

Analyzing EO Satellites Images

SCIENCE OF REMOTE SENSING

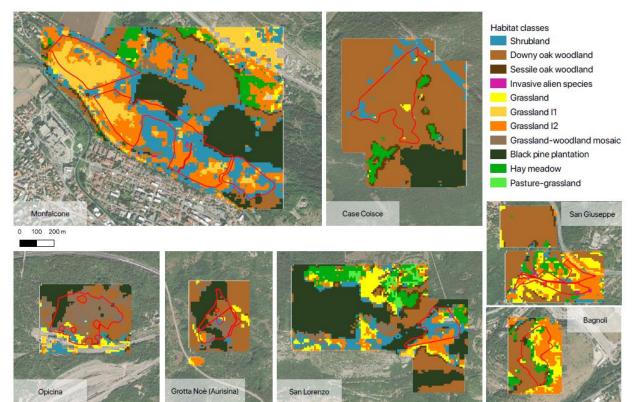


<u>Different approaches to use RS data to study global changes:</u>

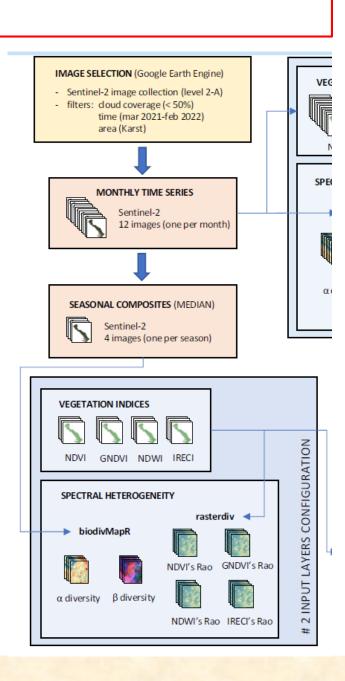
1. CLASSIFICATION

Process that assigns each element of the image to a certain category through digital interpretation.

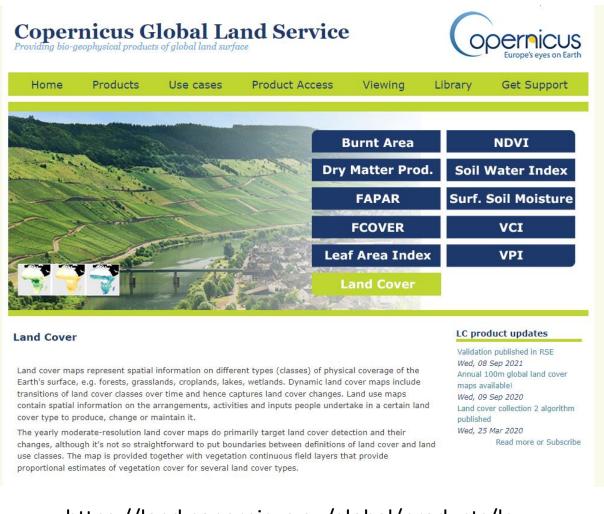
Converting numerical values in categories



Pafumi et al. In Preparation







https://land.copernicus.eu/global/products/lc

h) Landsat-8 LST

40.3° N

40.2° N

2. GENERATION OF CONTINUOUS VARIABLES

Generate spatial distribution of a biophysical variable derived from the sensor though empirical or physical models

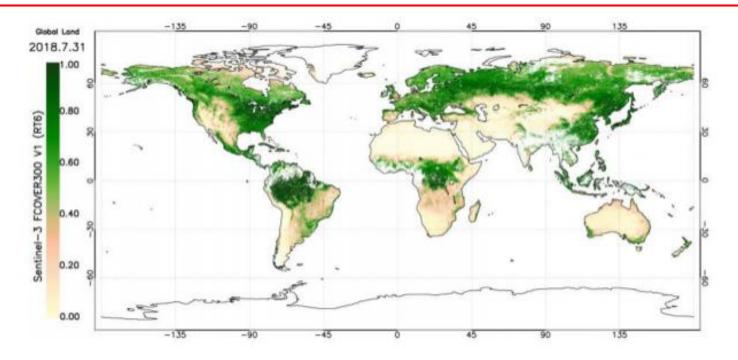
Converting raw digital values into biophysical values

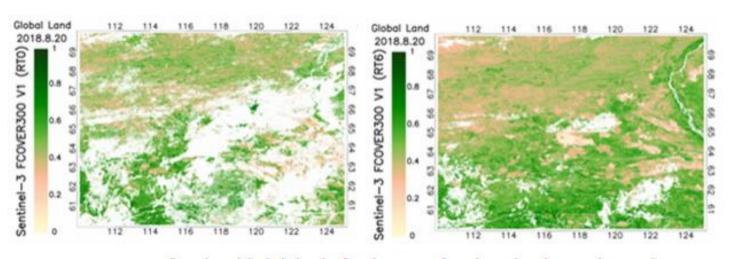
LST = Land Surface Temperature				Thermal Sensor	Operational Land (TIRS) Imager(TM) Landsat-8	
	BAND	SPECTRAL	WAVELEN. [µm]	GEOM. [m]	SENSOR	290 320
	1	aerosols	0.435 - 0.451	30	OLI	40.0° N
	2	blue	0.452 - 0.512	30	OLI	Ermida et al. 2020 Remote Sensing 8.9° W 8.8° W 8.7° W 8.6° W 8.5° W 8.4° W BND DRA
	3	green	0.533 - 0.590	30	OLI	$\mu = 1.2 \ 0.9 \ 1.3$ $\mu = 0.1 \ -0.6 \ -0.4$
	4	red	0.636 - 0.673	30	OLI	330 $\sigma = 1.4 \ 1.3 \ 1.1$ RMSE = 2.5 2.4 2.4 330 $\sigma = 0.9 \ 0.8 \ 1.0$ RMSE = 1.7 1.6 2.0
	5	NIR	0.851 - 0.879	30	OLI	320
	6	SWIR-1	1.566 - 1.651	30	OLI	∑ 310 ∑ 300 ∑ 300
	7	SWIR-2	2.107 - 2.294	30	OLI	∑ 300
	8	pan	0.503 - 0.676	15	OLI	1 290 1 290 1 290 1 290 1 290 1 290 1 290 280 1 290 28
	9	cirrus	1.363 - 1.384	30	OLI	$\frac{-1:1}{-1:1}$ 270 $\frac{-1:1}{-1:1}$ $\frac{-1:1}{-1:1}$
	10	TIR-1	10.600 - 11.190	100	TIRS	260 L5 0 L7 260 0 L7
	11	TIR-2	11.500 - 12.510	100	TIRS	250 260 270 280 290 300 310 320 330 340 250 260 270 280 290 300 310 320 330 340
						Insitu LST [K] 250 260 270 280 290 300 310 320 330 340 250 260 270 280 290 300 310 320 330 340

Vegetation biophysical variables:

FCOVER = Fraction of green vegetation cover

https://land.copernicus.eu/global/products/fcover

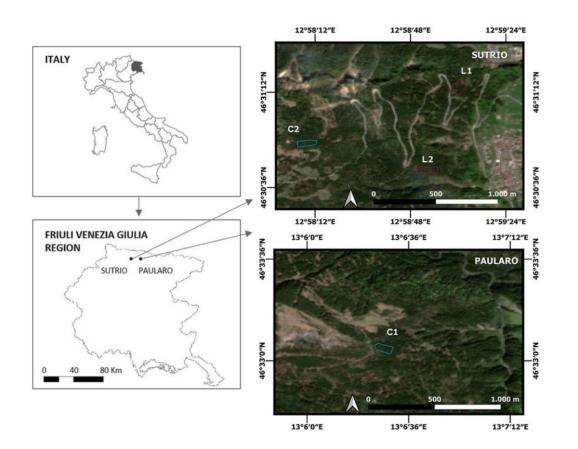


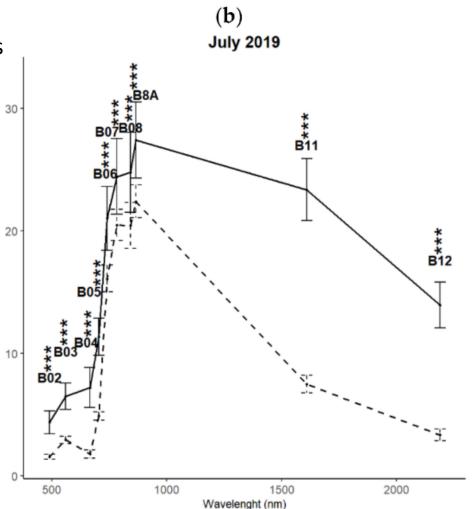


3. CHANGE DETECTION

Quantification of changes occurred between two or more images

Could be performed on both classified images or biophysical variables





remote sensing

Use of Sentinel-2 Satellite Data for Windthrows Monitoring and Delimiting: The Case of "Vaia" Storm in Friuli Venezia Giulia Region (North-Eastern Italy)

4. SPATIAL PATTERNS

Measure spatial properties of an object as: size, connectivity, shape.

ADRIATIC SEA MHW TRENDS

Bevilacqua et al. In Preparation

MHW = Marine Heatwaves





Copernicus Marine Service

Providing free and open marine data and services to enable marine policy

Copernicus Marine Service

opernicus

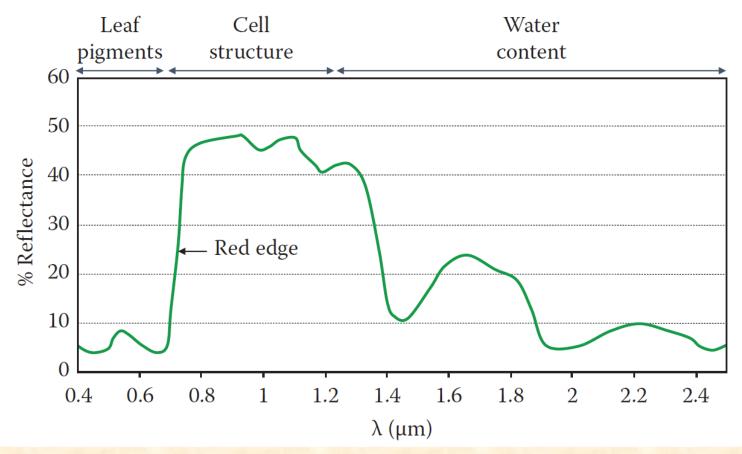
Access Data >

Chl content

- Transforming satellite measurements into biophysical variables
- The estimation of any variables relies on the physical relationship that links the surface properties with the radiance or reflectance measured by the sensor
- Two different variables types:
- **1. PRIMARY VARIABLES**: directly measured by the satellite.
 - Spectral radiances
 - Spectral reflectance
 - Land Surface T° in the TIR region
 - Surface backscatter coefficients in the microwave (MW) region
 - Heights through LiDAR measurements

- **2. SECONDARY VARIABLES**: inferred or derived from models.
 - Chlorophyll content
 - Water content
 - Leaf Area Index (LAI)
 - Fraction of photosynthetically active radiation absorbed by plants (fAPAR)
 - Soil moisture
 - Evapotranspiration (ET)

- For secondary variables, it is necessary to model the relationship between satellite and actual biophysical variables measurements
- Any empirical or theoretical model should be based on theoretical relationship between the parameter being estimated and radiances or reflectance measured by the sensor
- Model must be then validated on the ground



1. INDUCTIVE OR EMPIRICAL MODELS

- These models fit a numerical relationship between the parameter to be estimated and the satellite measurements based on in situ observations
- The satellite image must be calibrated
- Require simultaneous field data collection

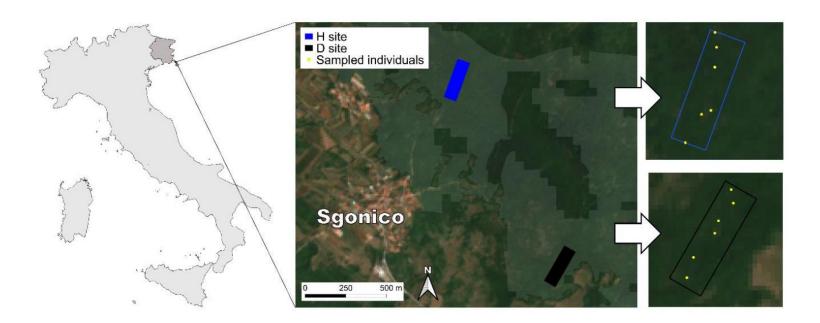




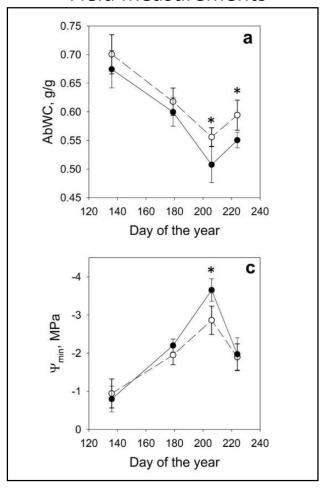
Articl

Correlation of Field-Measured and Remotely Sensed Plant Water Status as a Tool to Monitor the Risk of Drought-Induced Forest Decline

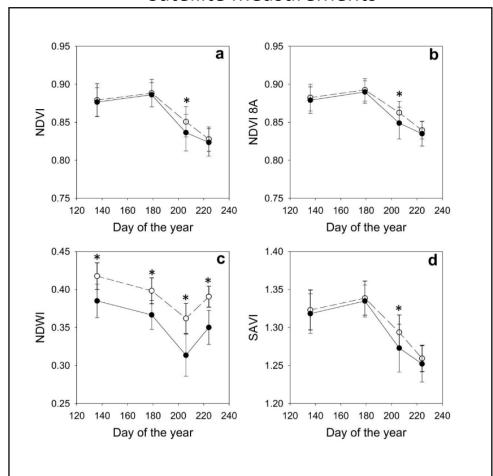
Daniel Marusig, Francesco Petruzzellis[®], Martina Tomasella[®], Rossella Napolitano, Alfredo Altobelli and Andrea Nardini *



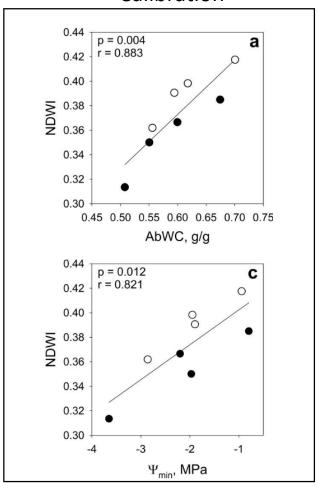
Field measurements



Satellite measurements



Calibration

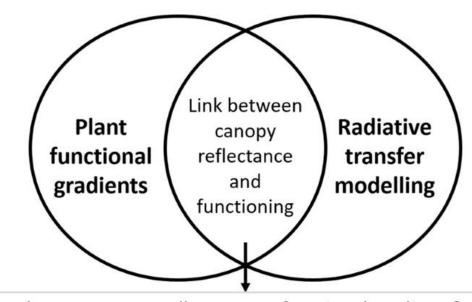


2. DEDUCTIVE OR THEORETICAL MODELS

- These models establish general relationships between the required parameter and satellite measurements
- Example: <u>Radiative Transfer Models</u> (RTMs)

• Relates incident radiation to vegetation canopies trough angular, structural biochemical and biophysics

characteristics



- Why can we spectrally separate functional gradients?
- Can optically relevant plant traits increase our understanding of plant functioning?

Kattenborn and Schmidtlein et al. 2029 Scientific Report



PROSAIL

Combines 4-stream canopy model SAIL

Calculates the diffuse and direct reflectance and transmittance of the whole canopy using:

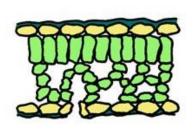
- Solar/viewing angle
- Leaf area index (m²/m²)
- Leaf angle distribution
- · Soil reflectance
- Leaf reflectance/transmittance

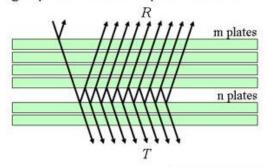
Solar incident downward flux Diffuse upward flux Canopy layer Soil

with leaf optics model PROSPECT

Calculates the reflectance and transmittance of a single leaf using a plate model dependent on:

- Internal leaf mesophyll structure
- Chlorphyll a+b and carotenoid content (µg/cm²)
- Dry matter content (g/cm²)
- Equivalent water thickness (cm)
- Brown pigment





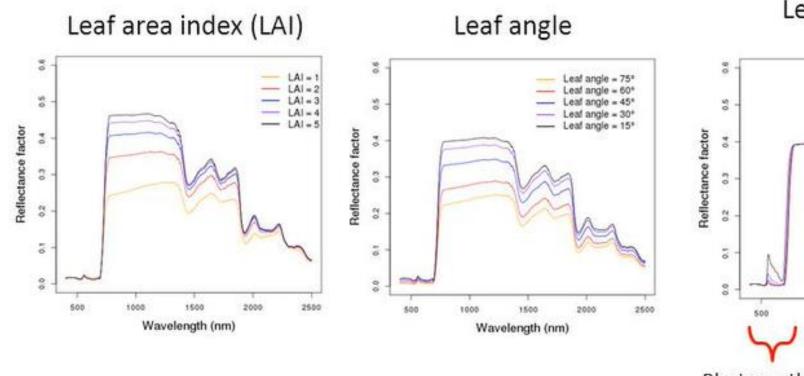
Jacquemoud & Ustin



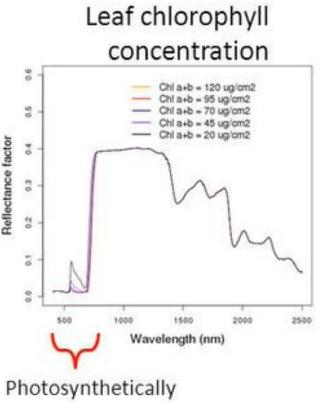




From radiative transfer model INVERSION it is possible to estimate plant traits



Simulations using PROSAIL



Photosynthetically active radiation (PAR) 400-700 nm

Hauser et al. 2021 Remote Sensing of Environment



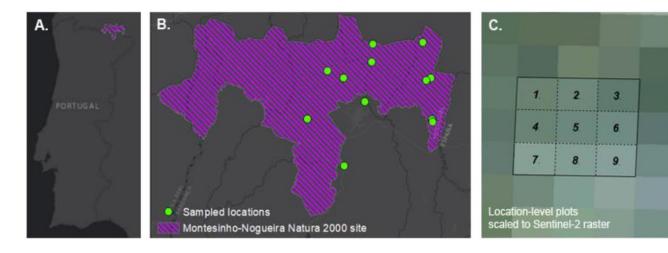
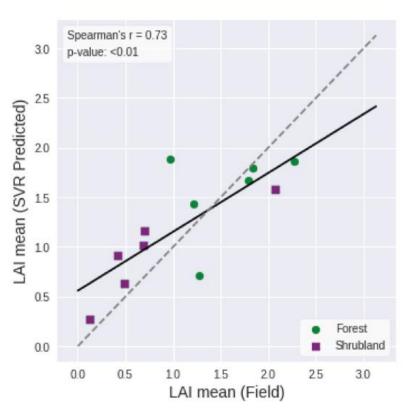


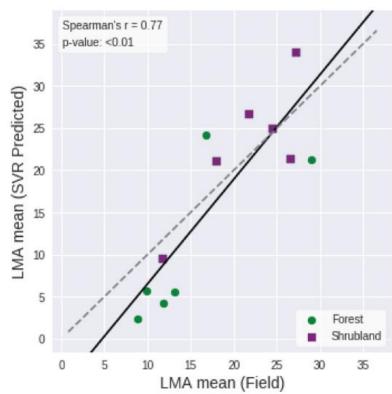
 Table 1

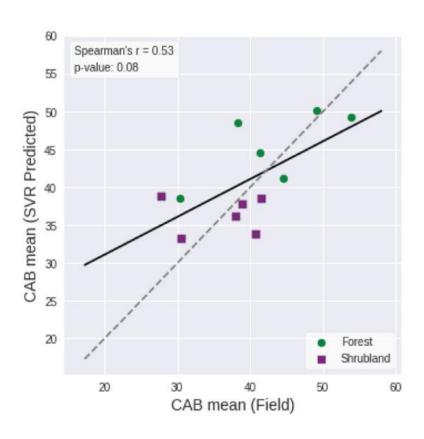
 Ranges of variable input parameters of the PROSAIL model used to generate the LUTs.

Domain	Parameter	Symbol	Unit	Distribution	Range
Leaf	Leaf structural parameter	N	_	Uniform	1.4–1.7
	Chlorophyll $a + b$ content	CAB	μg/cm ²	Gaussian	10–60
	Equivalent water thickness	EWT	g/cm ²	Uniform	0.001-0.045
	Leaf dry mass per area	LMA	g/cm ²	Uniform	0.001-0.040
	Brown pigments content	Cbrown	_	Fixed	0.01
Canopy	Leaf area index	LAI	m^2/m^2	Gaussian	0.01-3.5
	Mean leaf inclination angle	ALA	deg	Uniform	30–70
	Hot spot size parameter	hot	m/m	Fixed	0.01
Abiotic	Ratio of diffuse to total incident radiation	SKYL	_	Fixed	18%
	Soil brightness	psoil	_	Fixed	Spectroradiometer
Positional	Solar zenith	tts	0	Fixed	Sentinel-2 geometry
	Observer zenith	tto	0	Fixed	Sentinel-2 geometry
	Relative azimuth	phi	o	Fixed	Sentinel-2 geometry

Location-level (mean)







GENERATION OF CONTINUOUS VARIABLES

2. DEDUCTIVE OR THEORETICAL MODELS

- They also have limitations, as they need to be tested and constrained by actual data to find uncertainties and improve their performance
- These models are generally preferred to inductive models because of the mechanistic relationships between spectral data and biophysical variables and because they are more extendable in space and time

3. INTERMEDIATE APPROACHES

- Defined as semi-empirical models
- First step: physical modelling
- Second step: empirical adjustement

GENERATION OF CONTINUOUS VARIABLES

Beyond the previous complex approaches, there are simplified methods to generate continuous variables from remotely sensed data.

- 1. PRINCIPAL COMPONENT ANALYSIS
- 2. SPECTRAL VEGETATION INDICES (VIs)
 - 3. SPECTRAL MIXTURE ANALYSIS
 - 4. ADVANCED SATELLITES PRODUCTS

This is a type of analysis used when dealing with multivariate data.

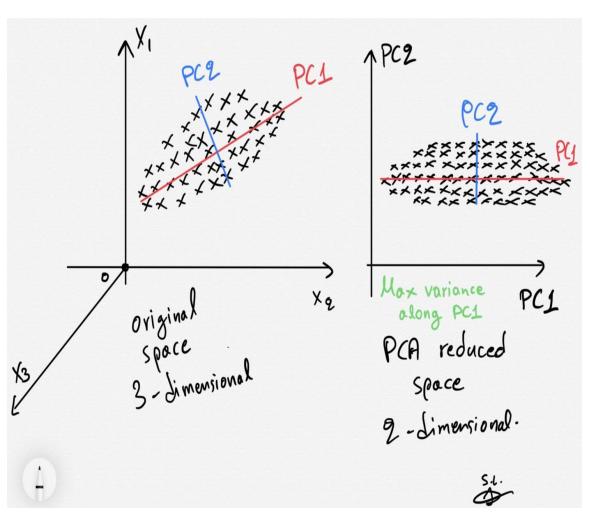


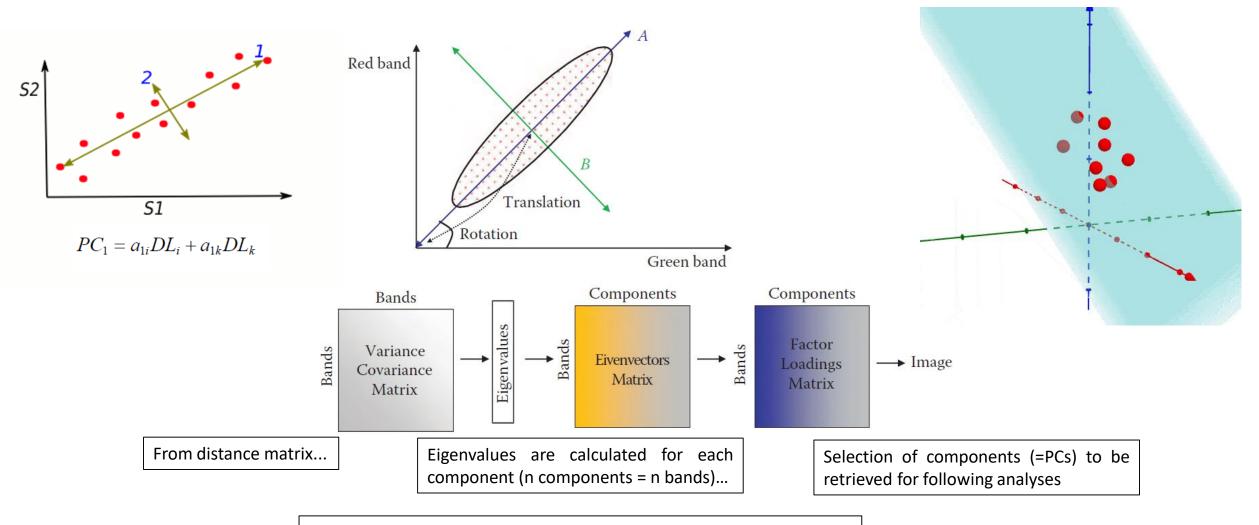
Multispectral data, which usually contains redundant information

This analysis perform a **DIMENSIONALITY REDUCTION**, by removing redundant spectral information present in the multiband images.



without losing a significant amount of original information.





Eigenvalues are proportional to the length of the associated components, which is related to the original information thy retained.

The PCs are obtained in such a way that eigenvalues decreases progressively from the first to the last ones, since the objective is to maximize successively the variance extracted in the analysis.

$$V_j = \frac{\lambda_j}{\sum_{j=1,p} \lambda_j} \longrightarrow \text{ Eigenvalue associated to j-component}$$

We can calculate the relationship between each band with each component, in order to interpret the meaning of the new variables (= the principal components) that have been calculated.

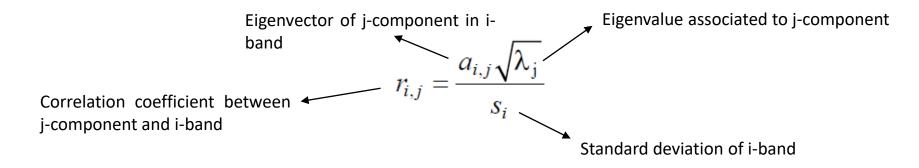
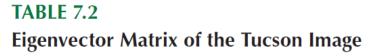


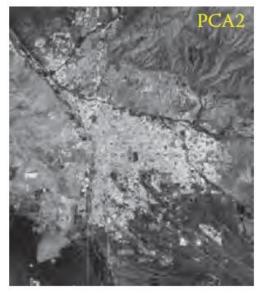
TABLE 7.1 Variance–Covariance Matrix of the Tucson Image

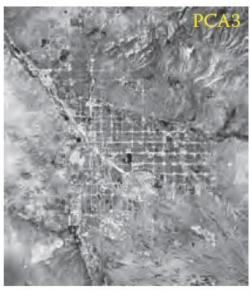
	B1	B2	В3	B4	B5	B 7
B1	1443.30					
B2	443.07	243.85				
B3	1043.43	414.51	1166.48			
B4	859.79	260.55	503.65	630.02		
B 5	999.67	319.26	665.64	664.89	741.61	
B 7	1020.65	342.95	1065.25	492.80	642.28	1039.70



	B1	B2	B 3	B4	B 5	B 7	λ	Variance (%)
CP1	0.552	0.188	0.465	0.320	0.380	0.444	4477.44	85.04
CP2	0.285	-0.017	-0.546	0.503	0.385	-0.468	613.23	11.65
CP3	0.250	-0.847	-0.252	-0.033	-0.063	0.389	112.82	2.14



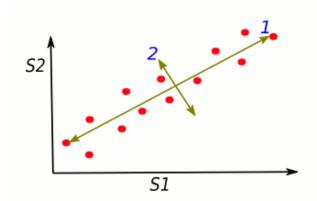


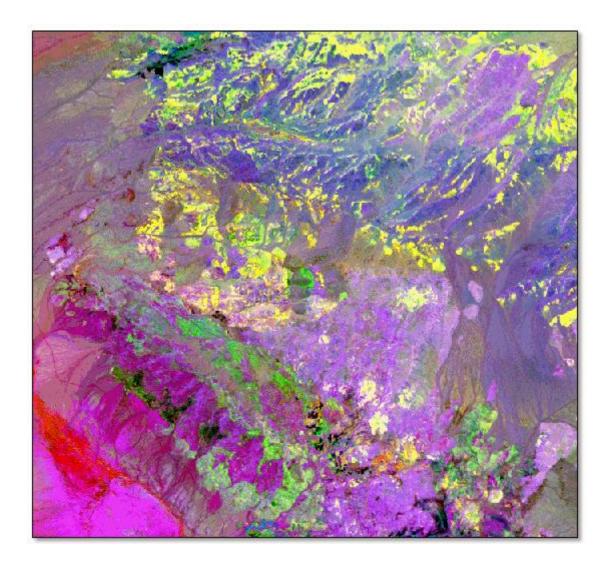


How can these information be used?

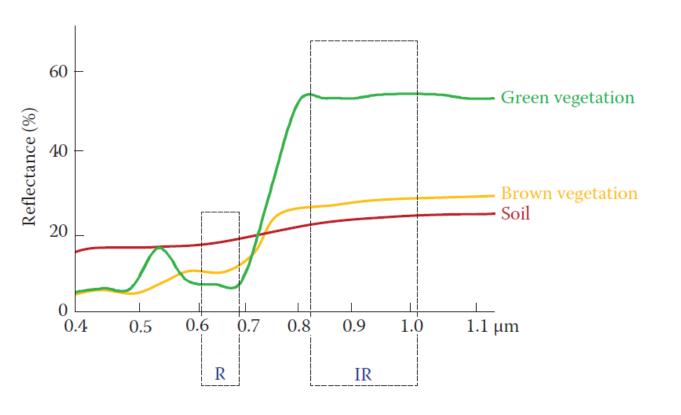
PCA scores = coordinates of each pixel/spatial unit in the space created by the retained PCs)

- <u>Visual inspection:</u> generate images and check which features can be discriminated
- Use PCs with the highest variance explained for <u>multitemporal classification</u>
- Use PCs' scores to <u>detect changes</u> between two or more dates





SPECTRAL VEGETATION INDICES (VIs)



The calculation of VIs is a simple and robust technique to extract quantitative information on the amount of vegetation or greenness.

The most used one involves the use of two bands:

- Red = R
- Near-InfraRed = NIR

Could you tell me why?

The goal is to combine R and NIR to enhance the vegetation signal.

SPECTRAL VEGETATION INDICES (VIs)

VIs can be used as direct measure of greenness, but can also be used as a proxy of biophysical variables:

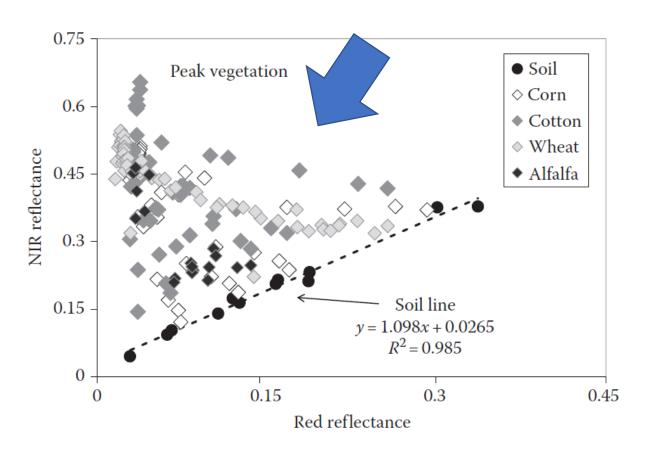
- Leaf Area Index
- fAPAR
- Green Vegetation Fraction (Fv)
- Biomass
- Photosynthesis

Indeed, greenness is a composite signal of leaf chlorophyll content, leaf area, canopy cover, canopy structure



VI = "integrator" of many variables that together determine the photosynthetically active vegetation signal.

SPECTRAL VEGETATION INDICES (VIs)



Starting point = structure of vegetation spectra in R-NIR space

There is a TRIANGULAR PATTERN:

- One side is represented by the "soil line", which indicates the absence of vegetation
- Green apex of maximum NIR and minimum R

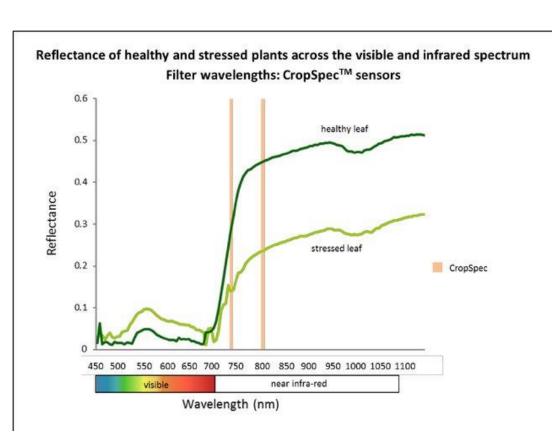
On these basis, the task of a VI is to quantify the amount of vegetation present in a pixel based on its position in the R-NIR space

Different ways to calculate VIs:

- 1. Band ratios
- 2. Normalized differences
- 3. Linear Band Combinations
- 4. Optimized band combinations

SPECTRAL VEGETATION INDICES (VIs) – Band ratios

$$SR = \frac{\rho_{NIR}}{\rho_{R}}$$



SR = simple ratio, the most simple one

It allows separating healthy vegetation from non-vegetation cover types

The main limitation regards the fact that both low and stressed vegetation have reduced reflectance in NIR and higher in R

Low SR values may both indicate low quantity of vegetation and high amount of stressed vegetation.

Ratios are useful in their ability to reduce many forms of multiplicative noise:

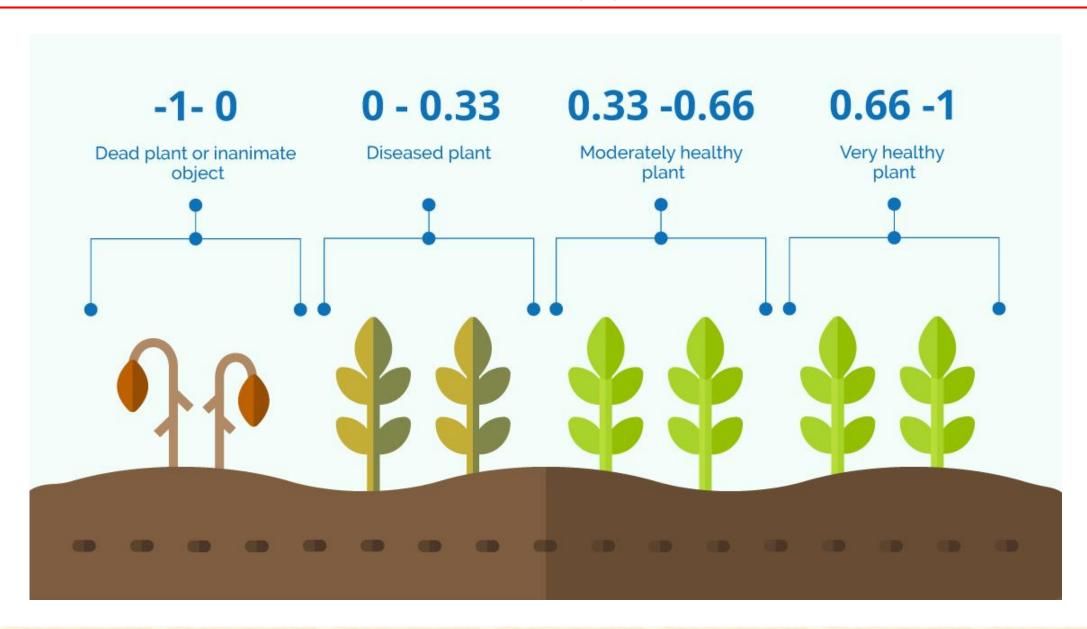
- illumination differences
- atmospheric attenuation
- cloud shadows
- topographic variations (slope, aspect)

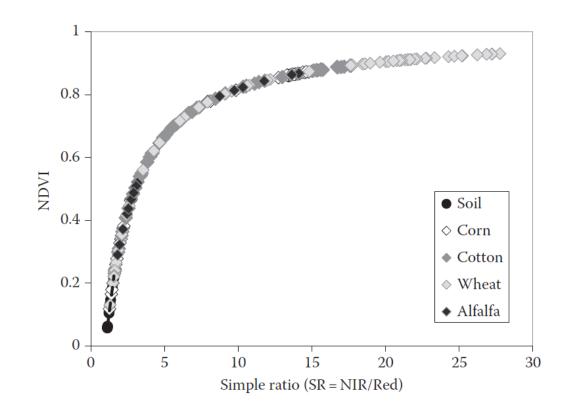
$$NDVI = \frac{\rho_{\text{NIR}} - \rho_{\text{R}}}{\rho_{\text{NIR}} + \rho_{\text{R}}}$$

NDVI = normalized different vegetation index

It ranges from -1 to 1+1

1 aerosols 0.429 – 0.457 60 2 blue 0.451 – 0.539 10 3 green 0.538 – 0.585 10 4 red 0.641 – 0.689 10 5 red edge 0.695 – 0.715 20 6 red edge 0.731 – 0.749 20 7 red edge 0.769 – 0.797 20 8 NIR 0.784 – 0.900 10 8a narrow NIR 0.855 – 0.875 20 9 water vapour 0.935 – 0.955 60 10 SWIR cirrus 1.365 – 1.385 60 11 SWIR 1.565 – 1.655 20 12 SWIR 2.100 – 2.280 20	BAND	SPECTRAL	WAVELEN. [µm]	GEOM. [m]
3 green 0.538 – 0.585 10 4 red 0.641 – 0.689 10 5 red edge 0.695 – 0.715 20 6 red edge 0.731 – 0.749 20 7 red edge 0.769 – 0.797 20 8 NIR 0.784 – 0.900 10 8a narrow NIR 0.855 – 0.875 20 9 water vapour 0.935 – 0.955 60 10 SWIR cirrus 1.365 – 1.385 60 11 SWIR 1.565 – 1.655 20	1	aerosols	0.429 - 0.457	60
4 red 0.641 – 0.689 10 5 red edge 0.695 – 0.715 20 6 red edge 0.731 – 0.749 20 7 red edge 0.769 – 0.797 20 8 NIR 0.784 – 0.900 10 8a narrow NIR 0.855 – 0.875 20 9 water vapour 0.935 – 0.955 60 10 SWIR cirrus 1.365 – 1.385 60 11 SWIR 1.565 – 1.655 20	2	blue	0.451 - 0.539	10
5 red edge 0.695 – 0.715 20 6 red edge 0.731 – 0.749 20 7 red edge 0.769 – 0.797 20 8 NIR 0.784 – 0.900 10 8a narrow NIR 0.855 – 0.875 20 9 water vapour 0.935 – 0.955 60 10 SWIR cirrus 1.365 – 1.385 60 11 SWIR 1.565 – 1.655 20	3	green	0.538 - 0.585	10
6 red edge 0.731 – 0.749 20 7 red edge 0.769 – 0.797 20 8 NIR 0.784 – 0.900 10 8a narrow NIR 0.855 – 0.875 20 9 water vapour 0.935 – 0.955 60 10 SWIR cirrus 1.365 – 1.385 60 11 SWIR 1.565 – 1.655 20	4	red	0.641 - 0.689	10
7 red edge 0.769 – 0.797 20 8 NIR 0.784 – 0.900 10 8a narrow NIR 0.855 – 0.875 20 9 water vapour 0.935 – 0.955 60 10 SWIR cirrus 1.365 – 1.385 60 11 SWIR 1.565 – 1.655 20	5	red edge	0.695 - 0.715	20
8 NIR 0.784 – 0.900 10 8a narrow NIR 0.855 – 0.875 20 9 water vapour 0.935 – 0.955 60 10 SWIR cirrus 1.365 – 1.385 60 11 SWIR 1.565 – 1.655 20	6	red edge	0.731 - 0.749	20
8a narrow NIR 0.855 – 0.875 20 9 water vapour 0.935 – 0.955 60 10 SWIR cirrus 1.365 – 1.385 60 11 SWIR 1.565 – 1.655 20	7	red edge	0.769 - 0.797	20
9 water vapour 0.935 – 0.955 60 10 SWIR cirrus 1.365 – 1.385 60 11 SWIR 1.565 – 1.655 20	8	NIR	0.784 - 0.900	10
10 SWIR cirrus 1.365 – 1.385 60 11 SWIR 1.565 – 1.655 20	8a	narrow NIR	0.855 - 0.875	20
11 SWIR 1.565 – 1.655 20	9	water vapour	0.935 - 0.955	60
	10	SWIR cirrus	1.365 - 1.385	60
12 SWIR 2.100 – 2.280 20	11	SWIR	1.565 - 1.655	20
	12	SWIR	2.100 - 2.280	20





Non linear relationship with SR, meaning that NDVI is highly sensitive to low and sparse vegetation.

When 0 < NDVI < 0.5, then 1 < SR < 3

But...

When 0.5 < NDVI < 0.9, then 3 < SR < 19

The advantages of NDVI are

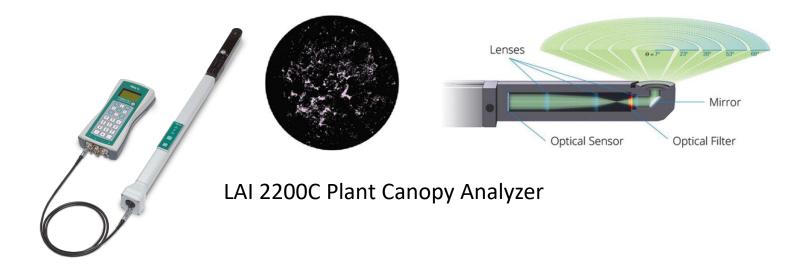
- More stable
- Less sensitive to external variations

NDVI has been used as a proxy of several biophysical variables:

1. Leaf Area Index (LAI): one of the fundamental vegetation biophysical variables. It is defined as the ratio between total canopy leaf area and its projected area on the ground.

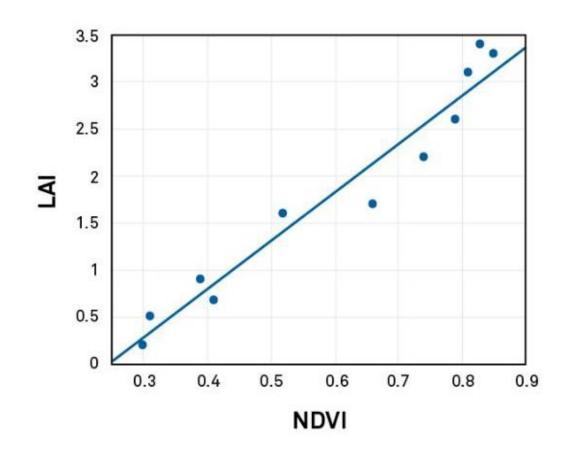


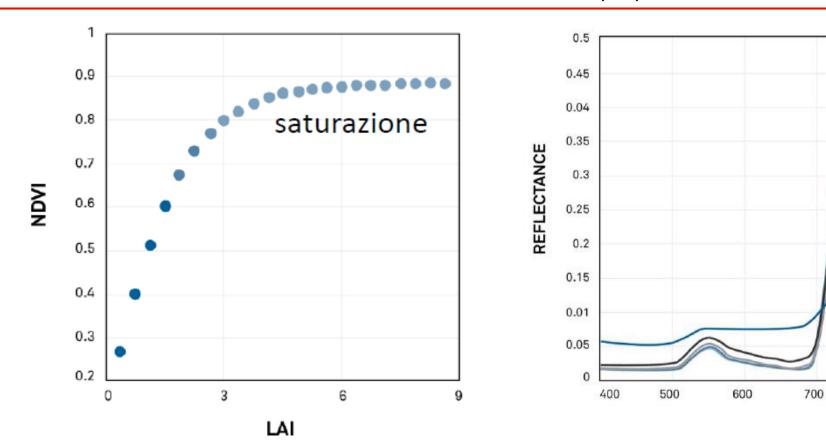
Hemispherical photos

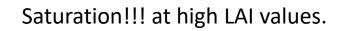


https://www.licor.com/env/products/leaf_area/LAI-2200C/applications

alassa	LAI	dev
classe	medio	std
acero-frassineto	3,56	±1,35
betuleto	4,45	±0,90
carpineto	3,75	±0,69
castagneto	4,09	±0,84
corileto	4,73	±0,57
faggeta	4,01	±1,06
ostrio-querceto	2,84	±0,83
pecceta	3,58	±0,98
piceo-faggeta	3,26	±0,61
rimboschimento	2,84	±1,83
rovereto	2,57	±0,55







NDVI is not a good proxy of LAI when vegetation is dense!

LAI: 5.9

LAI: 4.7

LAI: 3.4

LAI: 1.2

LAI: 0.5

900

1000

800

00

DANGER

NDVI has been used as a proxy of several biophysical variables:

- 1. Leaf Area Index (LAI): one of the fundamental vegetation biophysic as the ratio between total canopy leaf area and its projected area o
- **2.** Fractional vegetation cover (Fv): minimal saturation issue compare
- 3. Fraction of photosynthetically active radiation absorbed by plants
- **Chlorophyll content**
- Water content
- Net CO₂ flux
- 7. Gross and Primary productivity of vegetation
- **Evapotranspiration**



"a jack-of-all-trades"



a jack-of-all-trades is someone who is capable of doing several different jobs/roles

Example:

A: He can pitch, sell, negotiate; he's had input into product development and he optimised the production process - what I want to know is: is there anything this guy can't do?

> B: Yeah, bit of a jack-of-alltrades, isn't he?

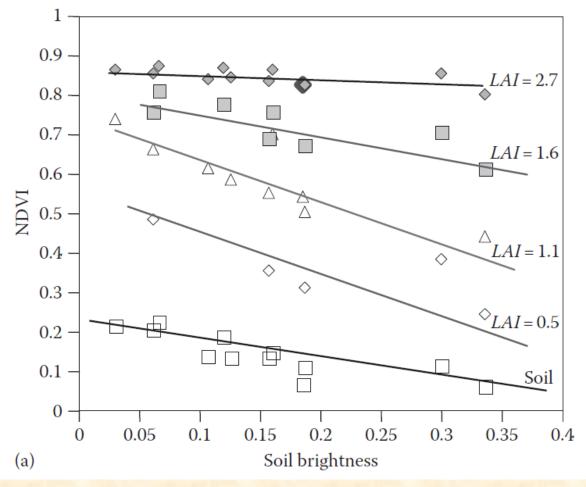




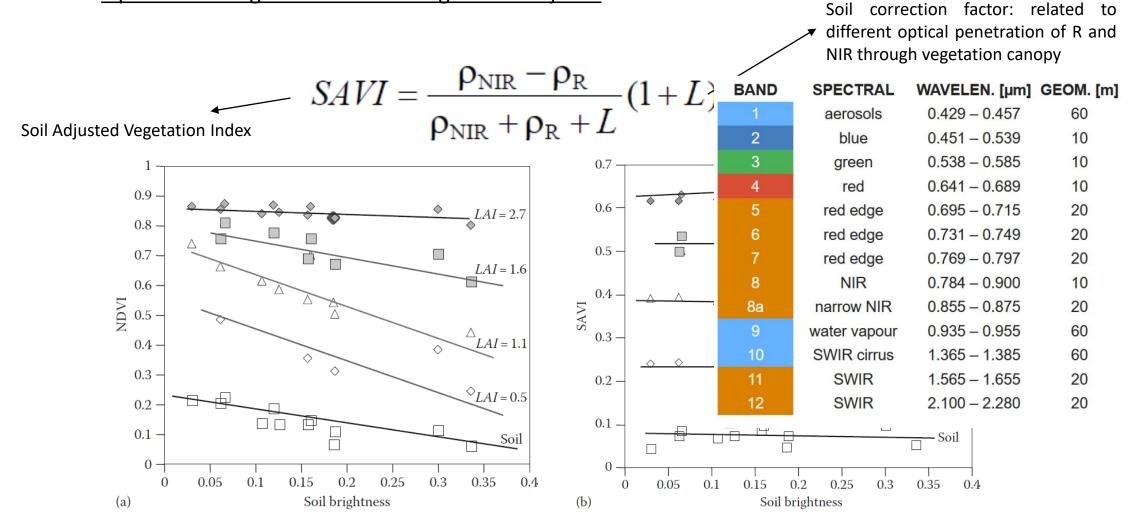
Normalized indices have some disadvantages:

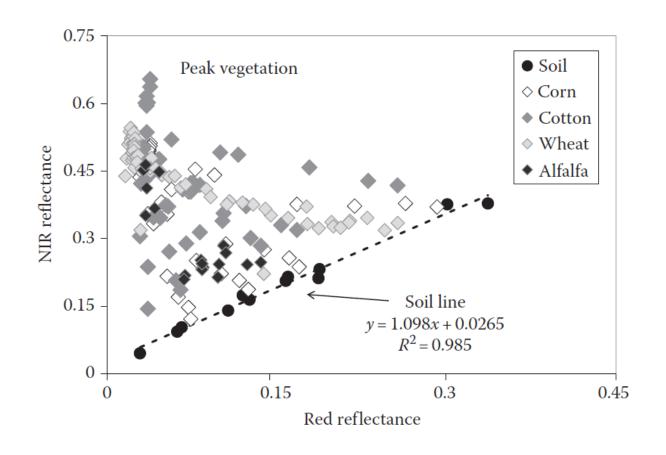
- Non-linear behaviour
- Sensitivity to soil brightness
- NDVI saturation at very high or very vegetation densities





Optimized indices introduced corrections, designed to measure NIR and R contrast of a pixel removing unwanted non-vegetation objects.





$$TSAVI = \frac{a(\rho_{NIR} - a\rho_{R}) - b}{a\rho_{NIR} + \rho_{R} + ab}$$

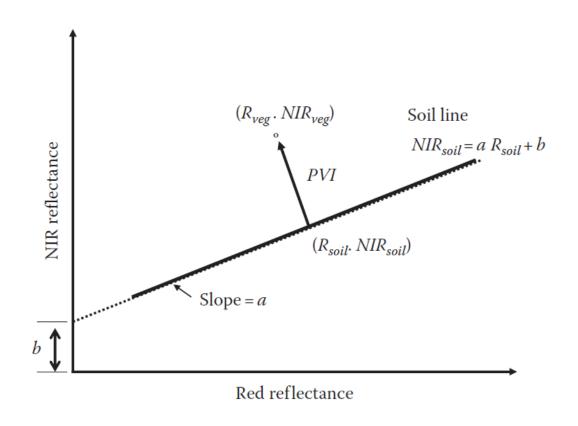
Correction using local-, regional- or global-based soil line. a and b are the coefficient of the linear equation in the figure.

SPECTRAL VEGETATION INDICES (VIs) – Linear Band combinations

In these VI, the contrast between R and NIR is measured by subtracting or linearly combining the R and NIR bands.

$$PVI = \frac{(\rho_{NIR} - a * \rho_{R}) - b)}{(1 + a^{2})^{1/2}}$$

Perpendicular vegetation index

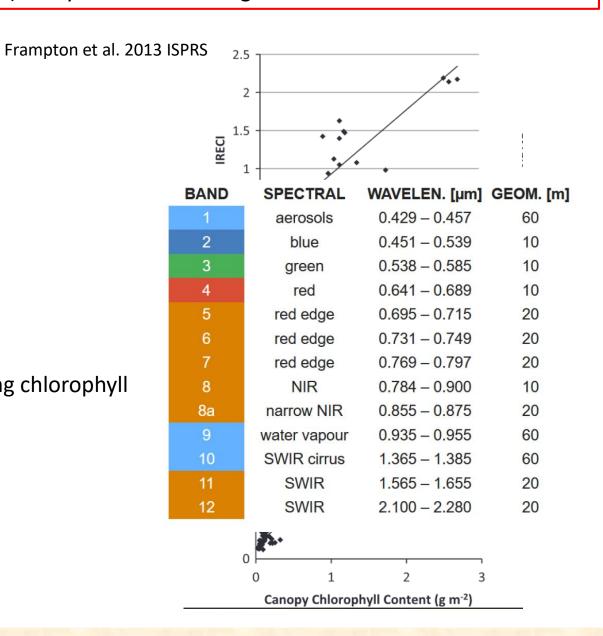


SPECTRAL VEGETATION INDICES (VIs) – Beyond NIR and R region

IRECI =
$$\frac{\left(\rho_{RE/NIR} - \rho_R\right)}{\left(\rho_{RE1}/\rho_{RE2}\right)}$$

IRECI = Inverted Red-Edge Chlorophyll Index

It uses Red Edge bands, which combine the effects of strong chlorophyll absorption and leaf internal scattering



SPECTRAL VEGETATION INDICES (VIs) – Beyond NIR and R region

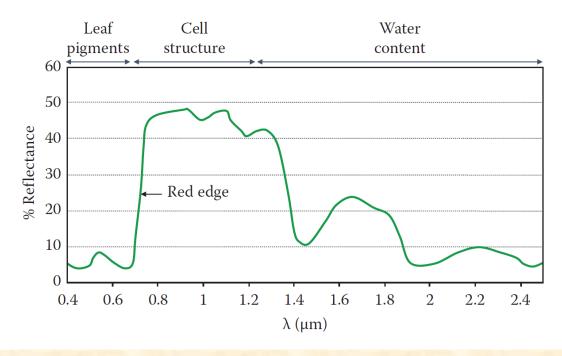
GNDVI = green NDVI

It uses the green band, and it is considered a proxy of the photosynthetic activity of the vegetation

$$GNDVI = \frac{(\rho_{NIR} - \rho_G)}{(\rho_{NIR} + \rho_G)}$$

Often used to asses water and nitrogen content

$$NDWI = \frac{(\rho_{NIR} - \rho_{SWIR})}{(\rho_{NIR} + \rho_{SWIR})}$$



https://www.indexdatabase.de/

https://custom-scripts.sentinel-hub.com/custom-scripts/sentinel-2/indexdb/