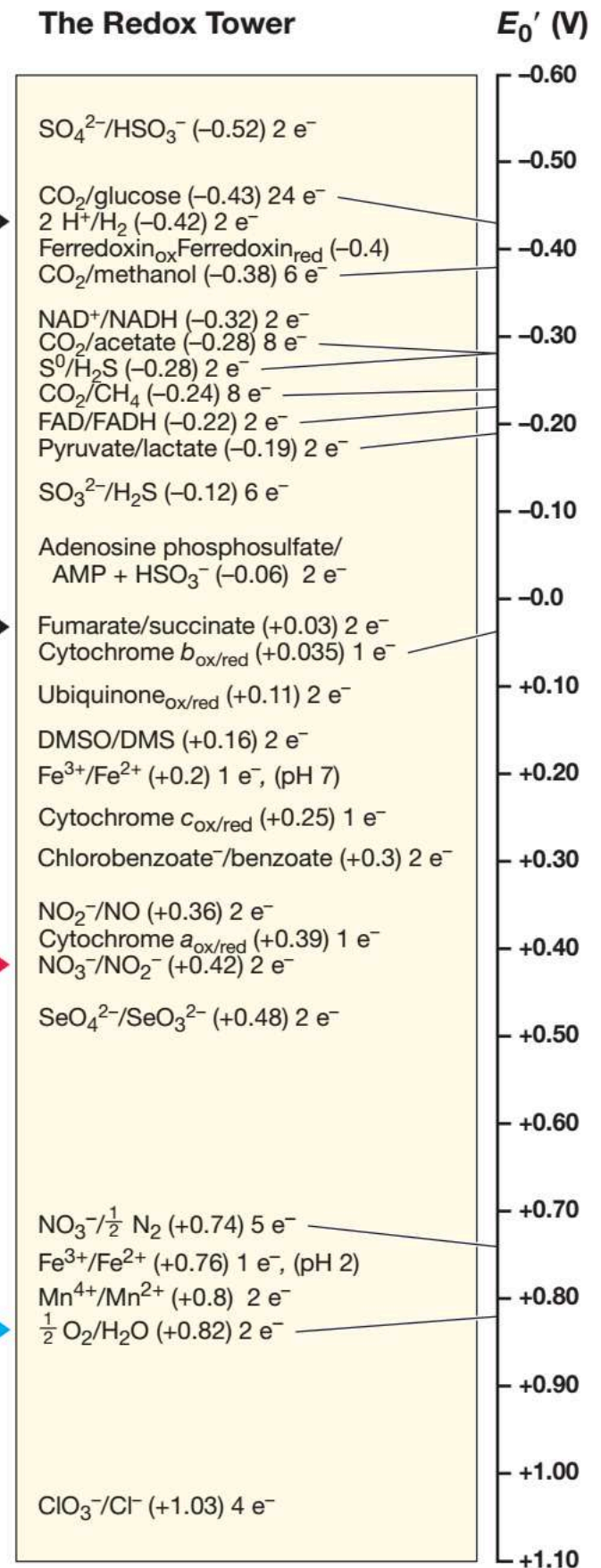
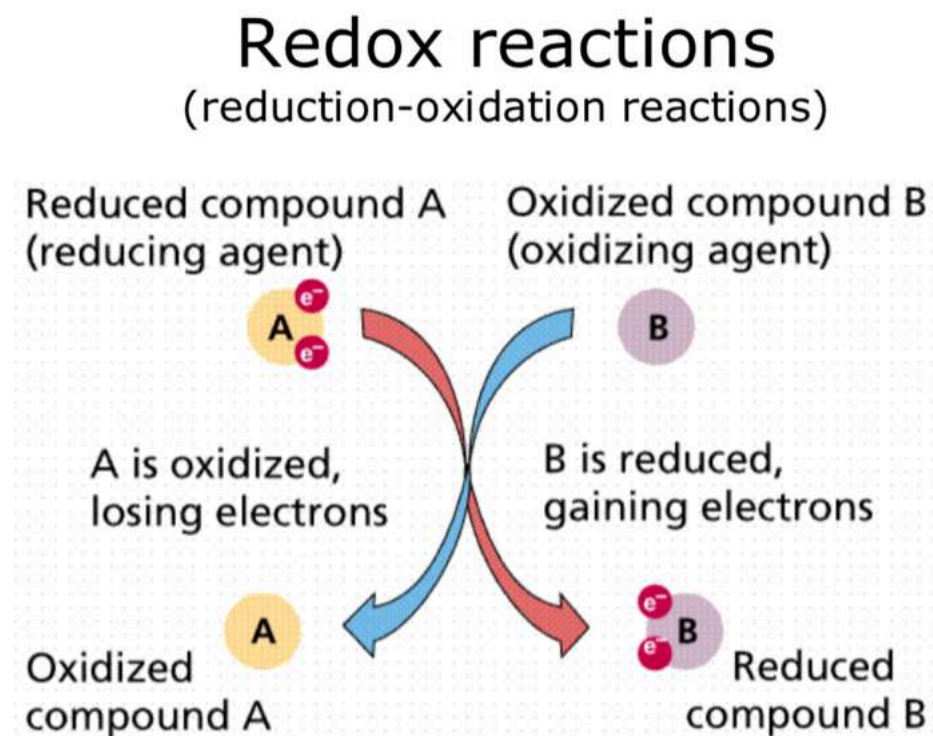


**LO3b**

# Recap L03

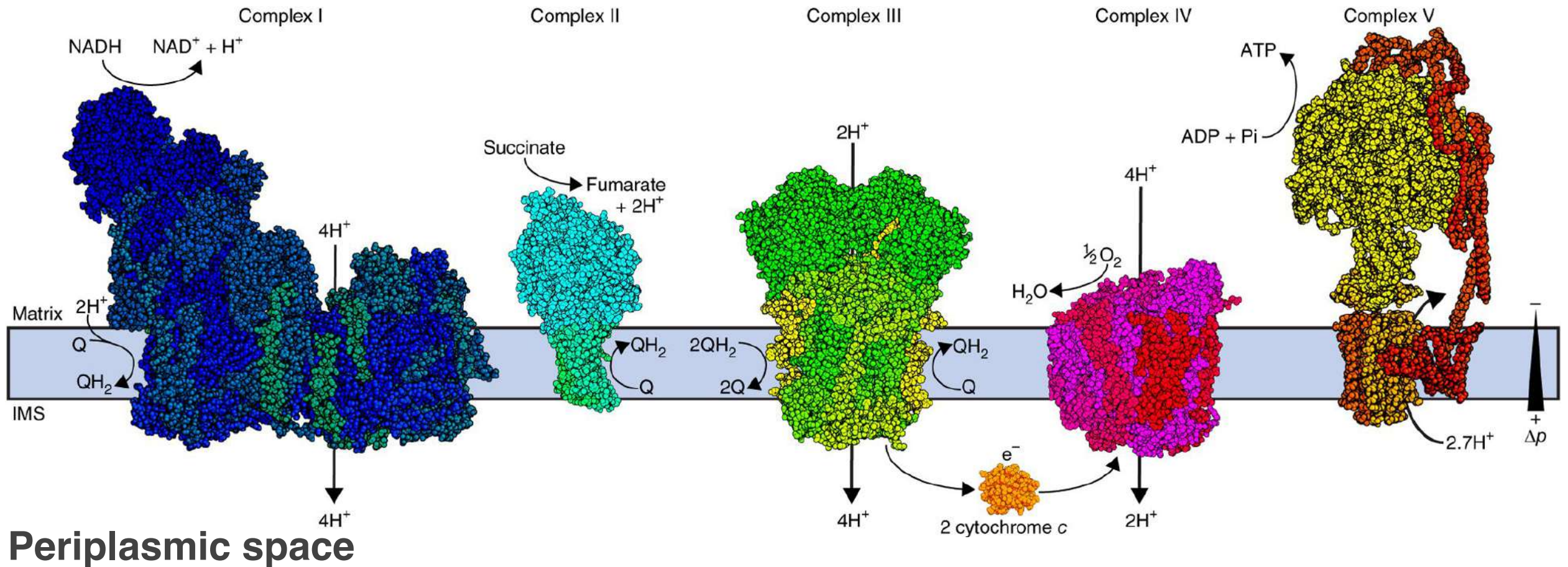
# The Redox Tower

- Redox couples are arranged from the **strongest e<sup>-</sup> donors at the top ( $E_0' < 0$ )** to the strongest e<sup>-</sup> acceptors at the bottom ( $E_0' > 0$ )
- The larger the difference in reduction potential between electron donor and electron acceptor, the more free energy is released ( $\Delta G_0'$  can be computed via Nernst equation from reduction potential)



# Electron transport chain (ETC), I

## Cytoplasm



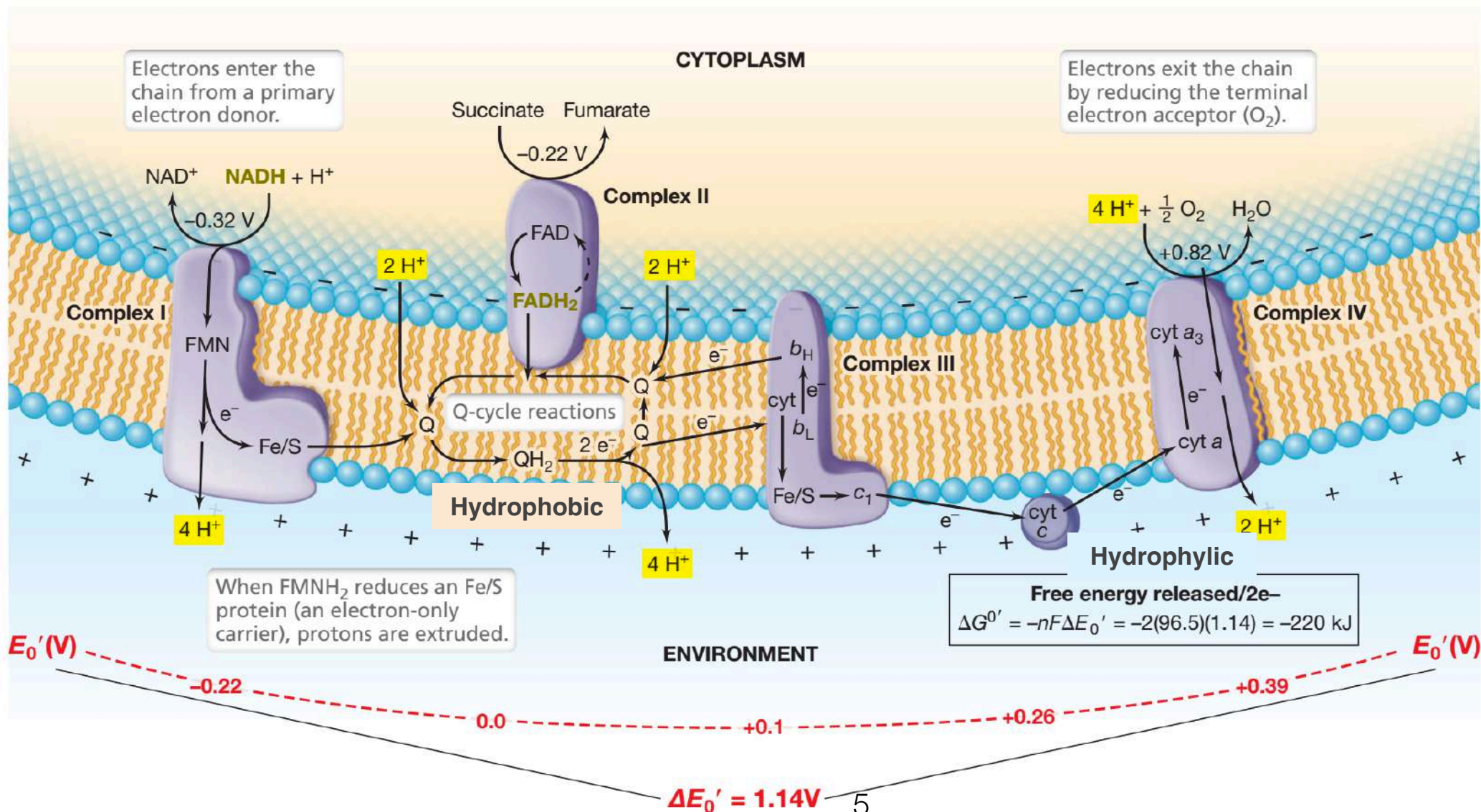
## Periplasmic space

- In the membrane
- Intimate interaction between proteins (dehydrogenase, flavoproteins, iron-sulfur proteins) and diffusible molecules (quinons and cytochromes)
- Electrons are swapped
- Protons are pumped outside the cell (cytoplasm  $\rightarrow$  periplasmic space)



# Electron transport chain, II

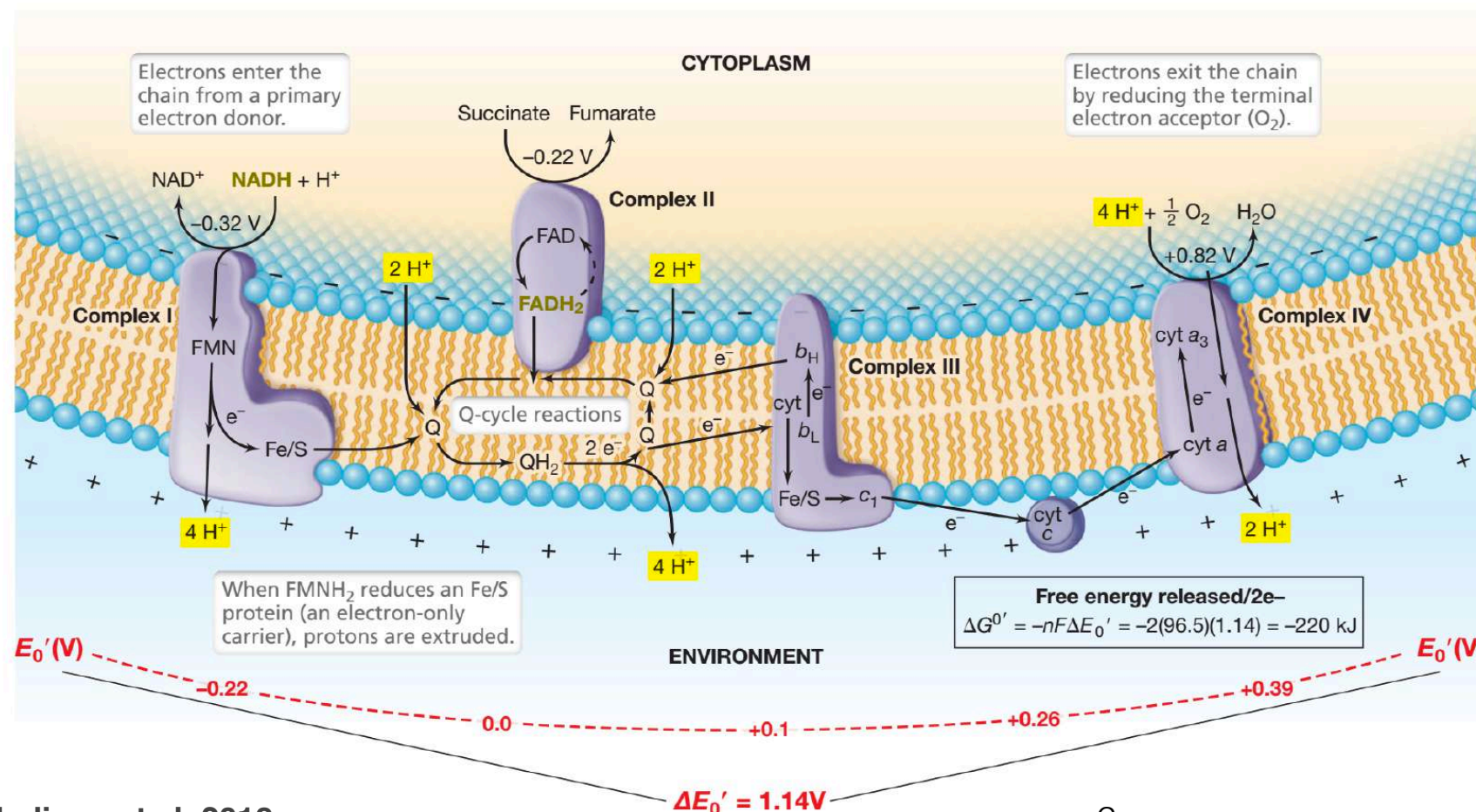
- A set of membrane-bound electron carriers (4) organized from **high to low redox potentials** —> **spontaneous** flow of electrons to the **terminal electron acceptor**
- The **membrane carriers are not structurally linked** so they can **diffuse** laterally in the membrane and collide with one another to promote the rapid exchange of electrons
- *Escherichia coli* uses lipophilic organic molecules called **quinones** to **electronically link a dehydrogenase enzyme complex to a specific terminal reductase**



# Electron transport chain, III

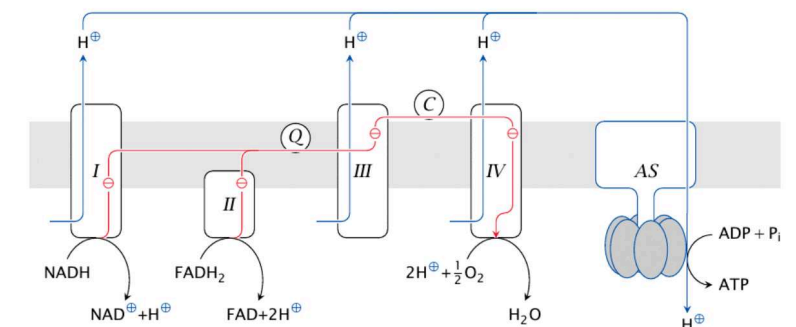
General features:

- (1) **Carriers** are arranged in order of **increasingly more positive  $E_0'$**  (reduction potential)
- (2) **Alternation of electron-only and electron-plus-proton carriers** in the chain
- (3) Net result is **reduction of terminal electron acceptor** (such as  $O_2$ ) + **generation proton motive force** (PMF, thanks to harnessing  $e^-$  flow)
- (4) ATP production by PMF (ATP synthesis is driven by an ion gradient through the activity of ATP synthase)



Environment

$H^+$  flow



$e^-$  flow

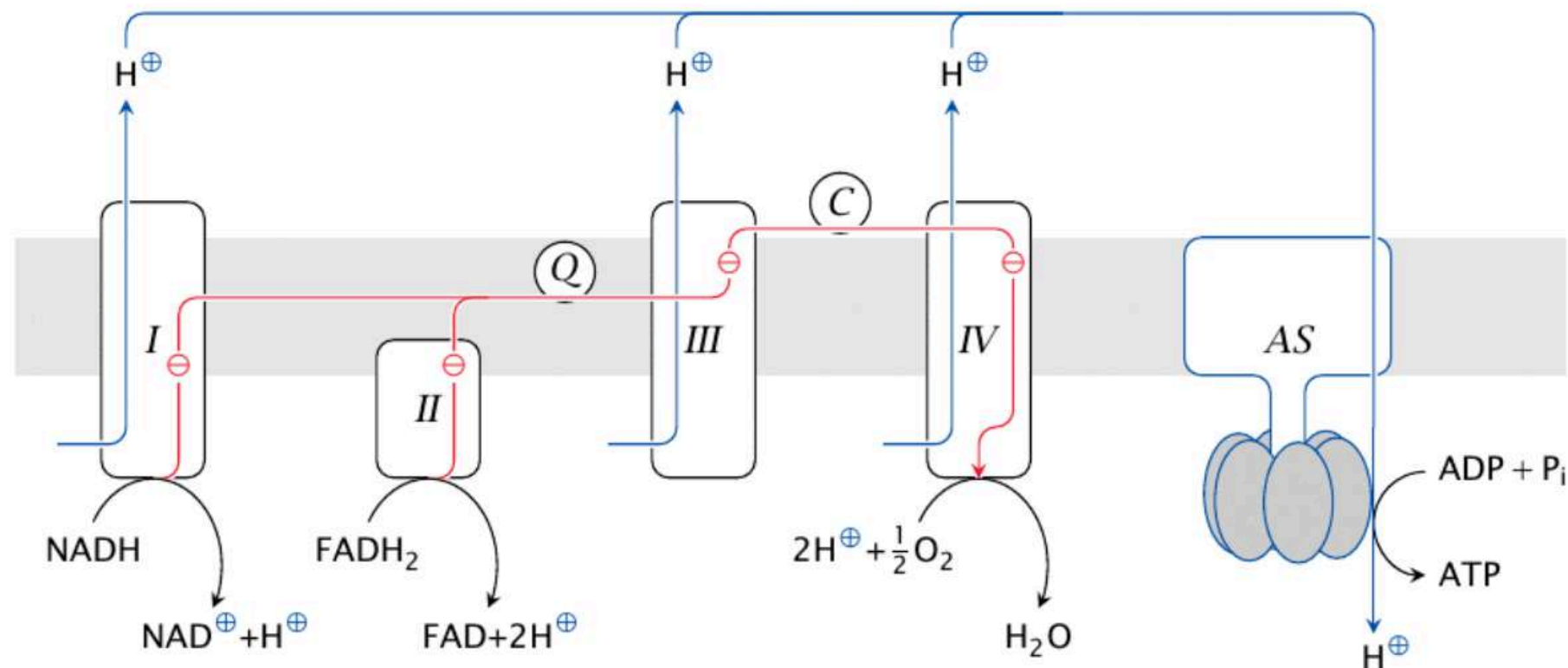
Cytoplasm



# Electron transport chain, III

General features:

- (1) **Carriers** are arranged in order of **increasingly more positive  $E_0'$**  (reduction potential)
- (2) **Alternation of electron-only and electron-plus-proton carriers** in the chain
- (3) Net result is **reduction of terminal electron acceptor** (such as  $O_2$ ) + **generation proton motive force** (PMF, thanks to harnessing  $e^-$  flow)
- (4) ATP production by PMF (ATP synthesis is driven by an ion gradient through the activity of ATP synthase)

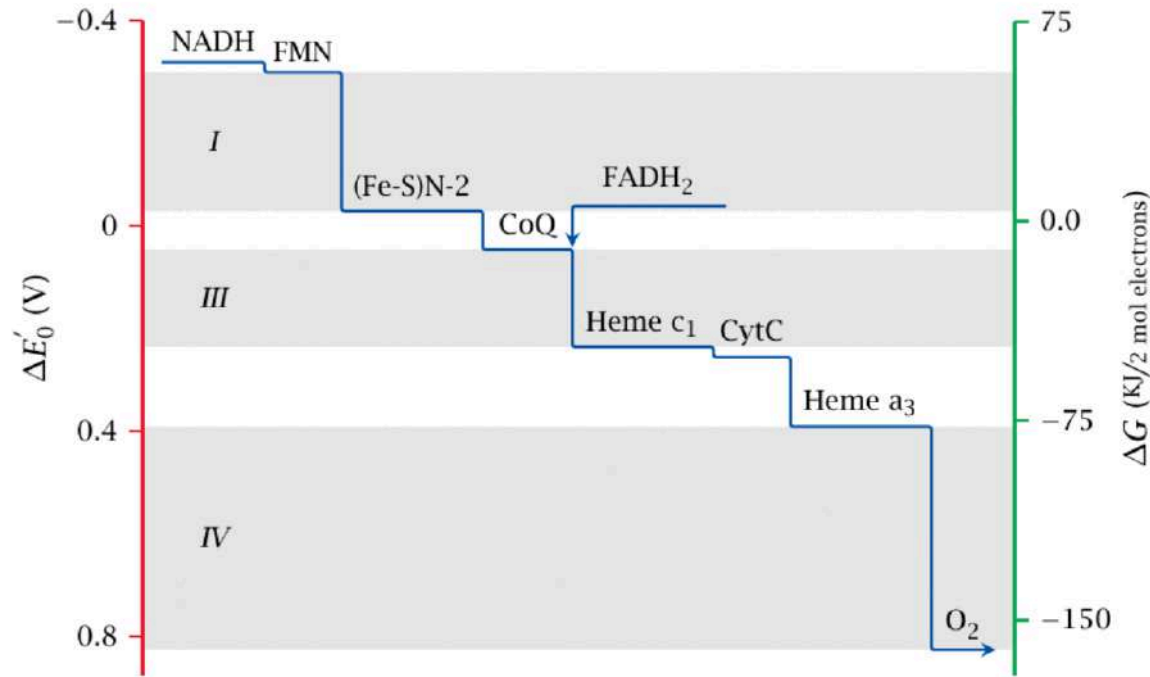


**$H^+$  flow**

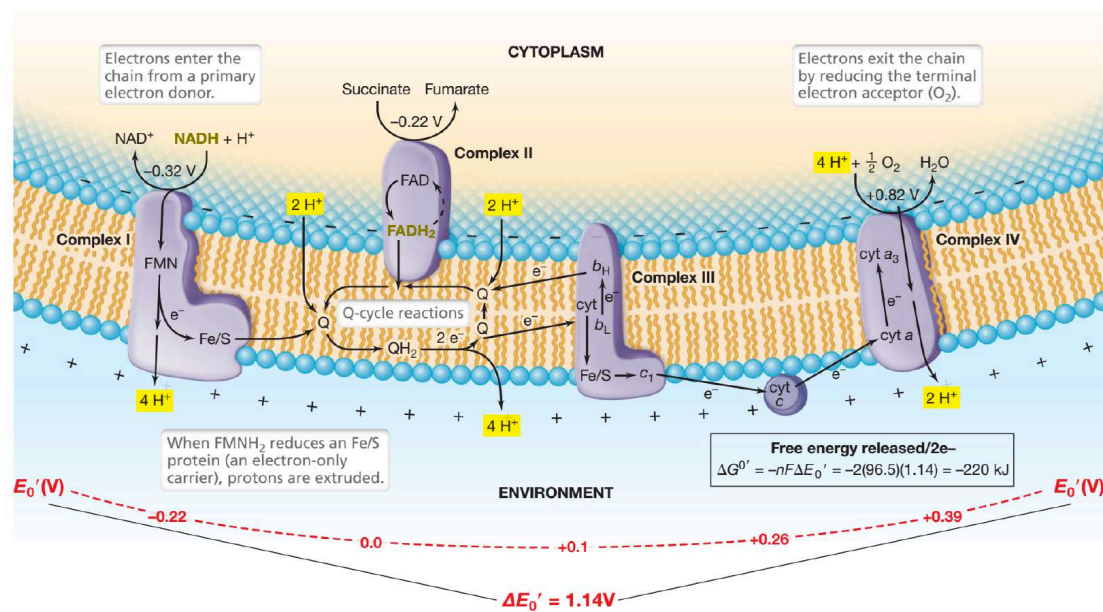
**$e^-$  flow**

# Structural orientation for ATP production

## Redox potentials and free energies in the respiratory chain



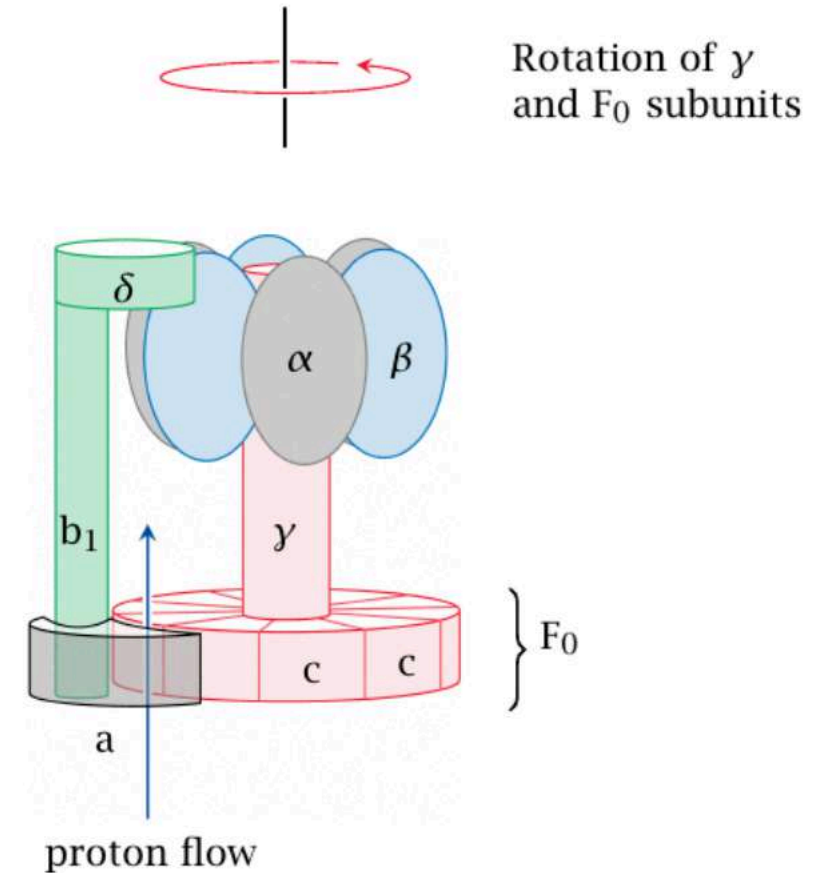
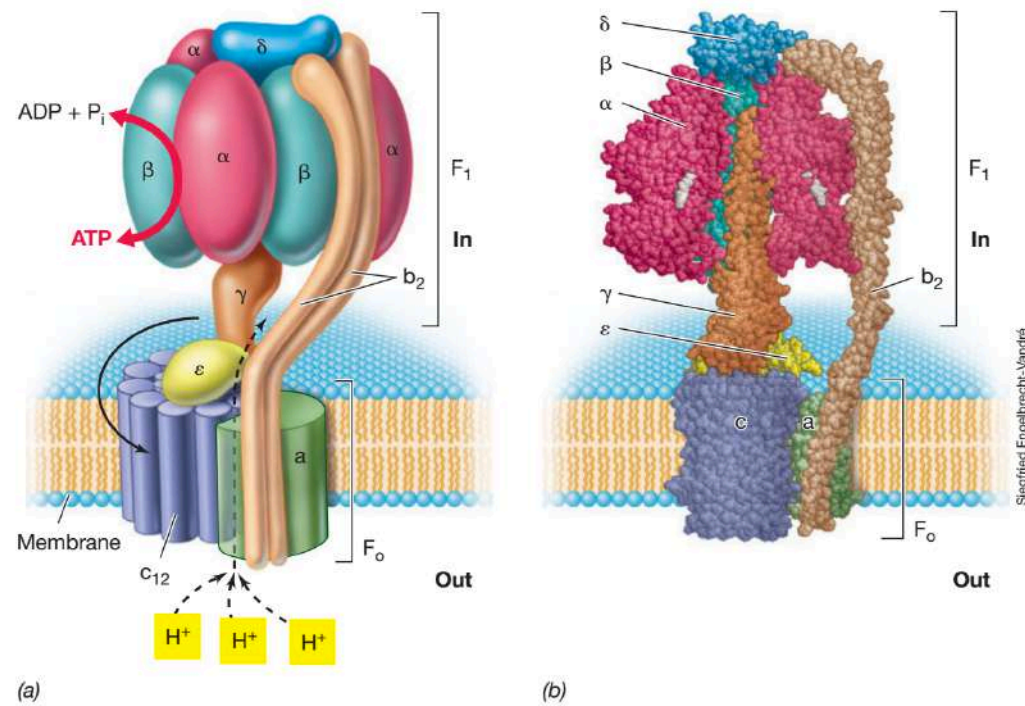
- Spontaneous flow of electrons ( $E_0'$ )
- $H^+$  are separated from  $e^-$  across membrane (spatial localization ETC)
- Inner and outer surfaces of the membrane differ in charge, pH, and electrochemical potential
- Electrochemical potential is proton motive force (PMF) and energizes the membrane, much like a battery
- Only three of the four mentioned electron carriers are capable of transporting protons from the matrix to the intermembrane space: I, III, and IV



http://wcut.uwaterloo.ca/webnotes/Metabolism/RespiratoryChain.html

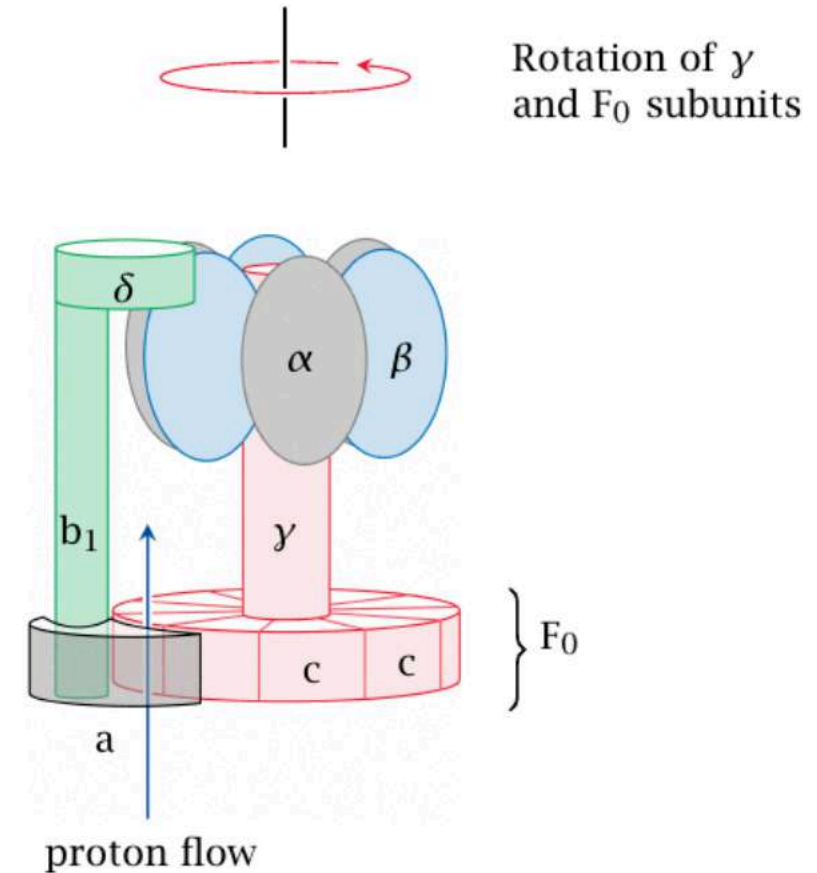
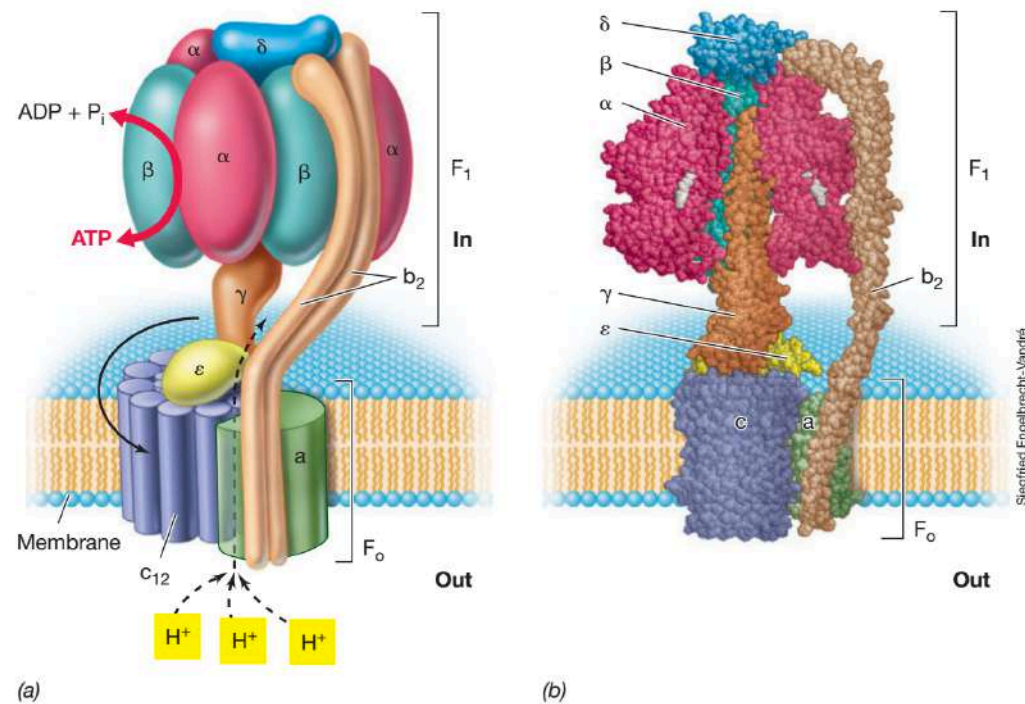


# ATP production



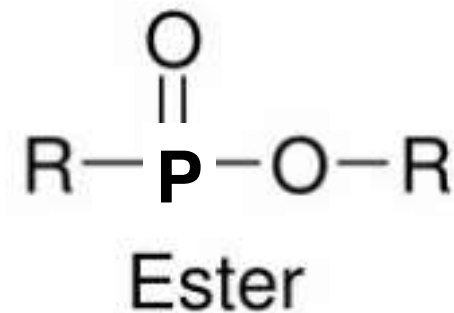
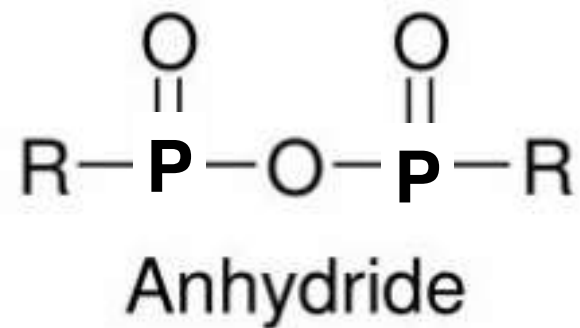
- $H^+$  gradient that drives phosphorylation of ADP to ATP as well as several other important transport systems (nutrient transport, flagellar rotation, and other energy-requiring reactions)
- $3 H^+ \rightarrow ATP$  (Noguchi et al., 2004): F1 is the catalytic complex responsible for the interconversion of ADP + Pi and ATP. F0, the rotor, is integrated in the membrane

# ATP production



- In analogy to how dissipation of the pmf applies torque that rotates the bacterial flagellum, the pmf also creates torque in the large membrane protein complex that synthesizes ATP
- This complex is called ATP synthase (ATPase)
- The activity of ATPase is driven by the pmf, and the formation of ATP from respiratory electron flow is called oxidative phosphorylation (contrast this with substrate-level phosphorylation in fermentation)

# C and P: Anhydrides and Esters in ATP



**ATP**

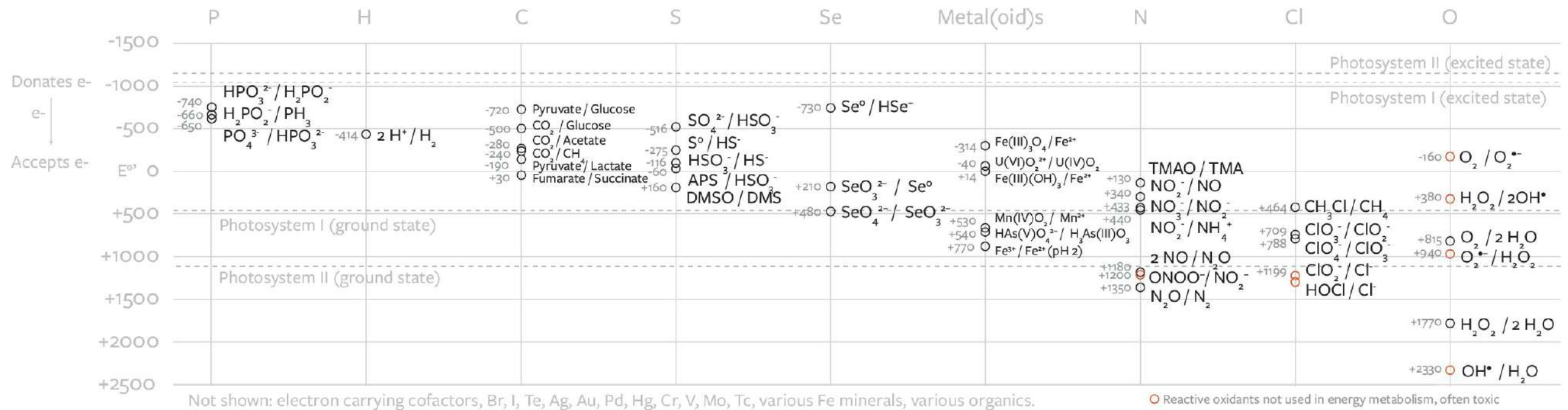


# Microbial Redox couples

## Redox couples and potentials (mV) for elements common in biology at pH 7 and temperature 25 C \*

Redox potential indicates the propensity for a compound to transfer electrons to another compound. A more-negative redox potential means a compound is more likely to donate electrons (e-).

All of life gets its energy by capturing the change in potential energy from the transfer of electrons from the reducing compound to the oxidizing compound.



- Across periodic table

- P, H, C, S, Se, Fe, U, Mn, As, N, Cl, O

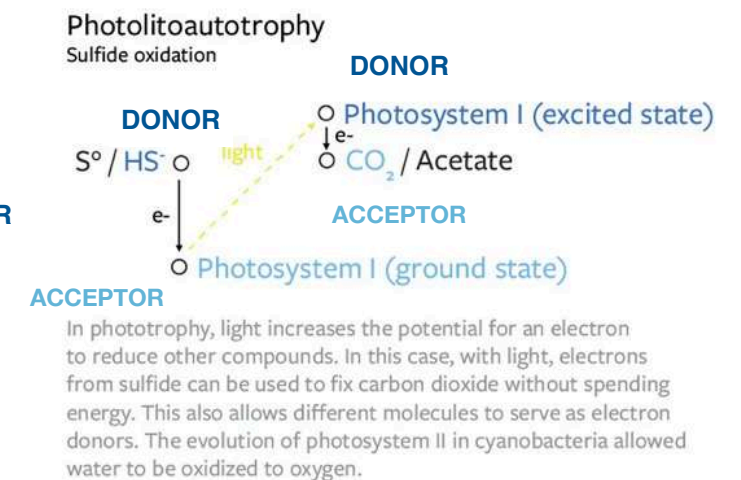
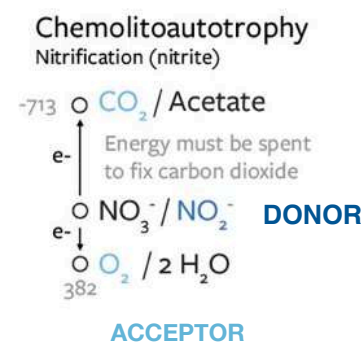
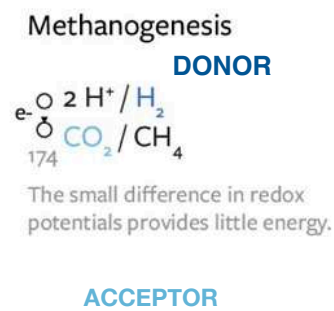
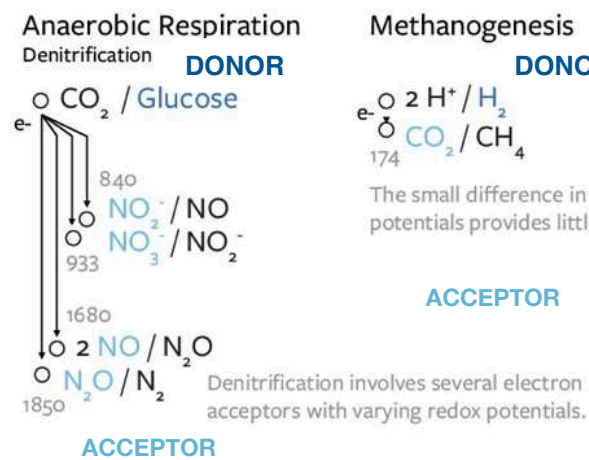
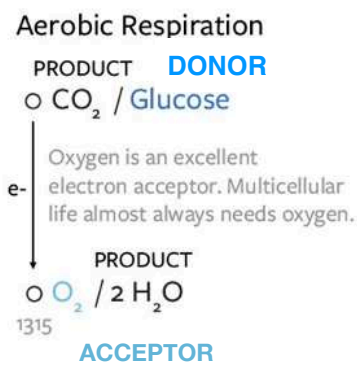
\* For teaching purposes only. Consult the scientific literature for exact values.

# Microbial Redox couples structure the metabolism

## Examples of energetically favorable redox metabolisms



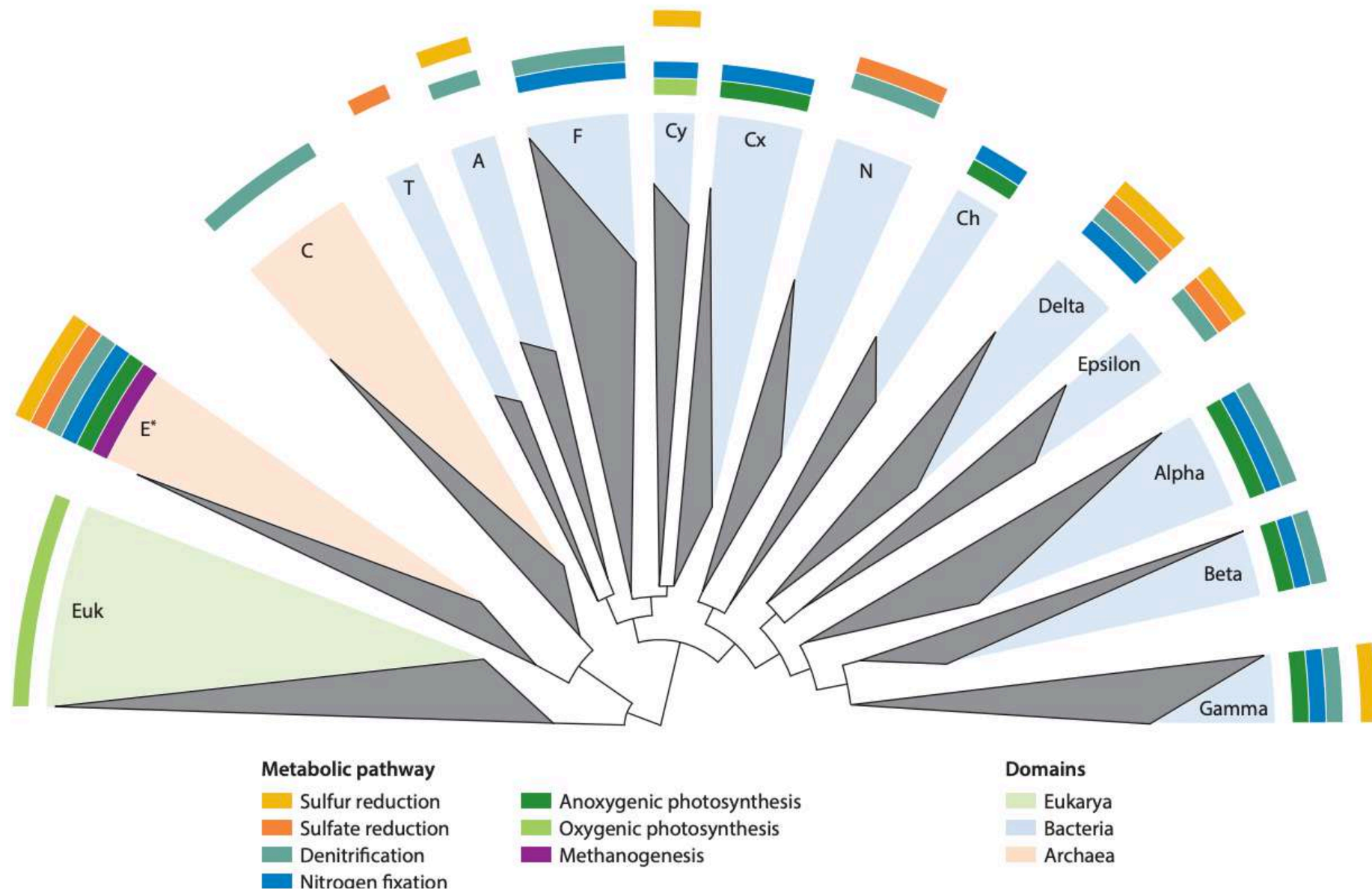
The electron acceptor in fermentation is the partially oxidized electron donor. While often a back-up metabolism, many microbes use only fermentation for energy.



\* For teaching purposes only. Consult the scientific literature for exact values.

Image produced by Tyler Barnum @tylerbarnumphd

# Microbial diversity and metabolic pathways to survive in the environment

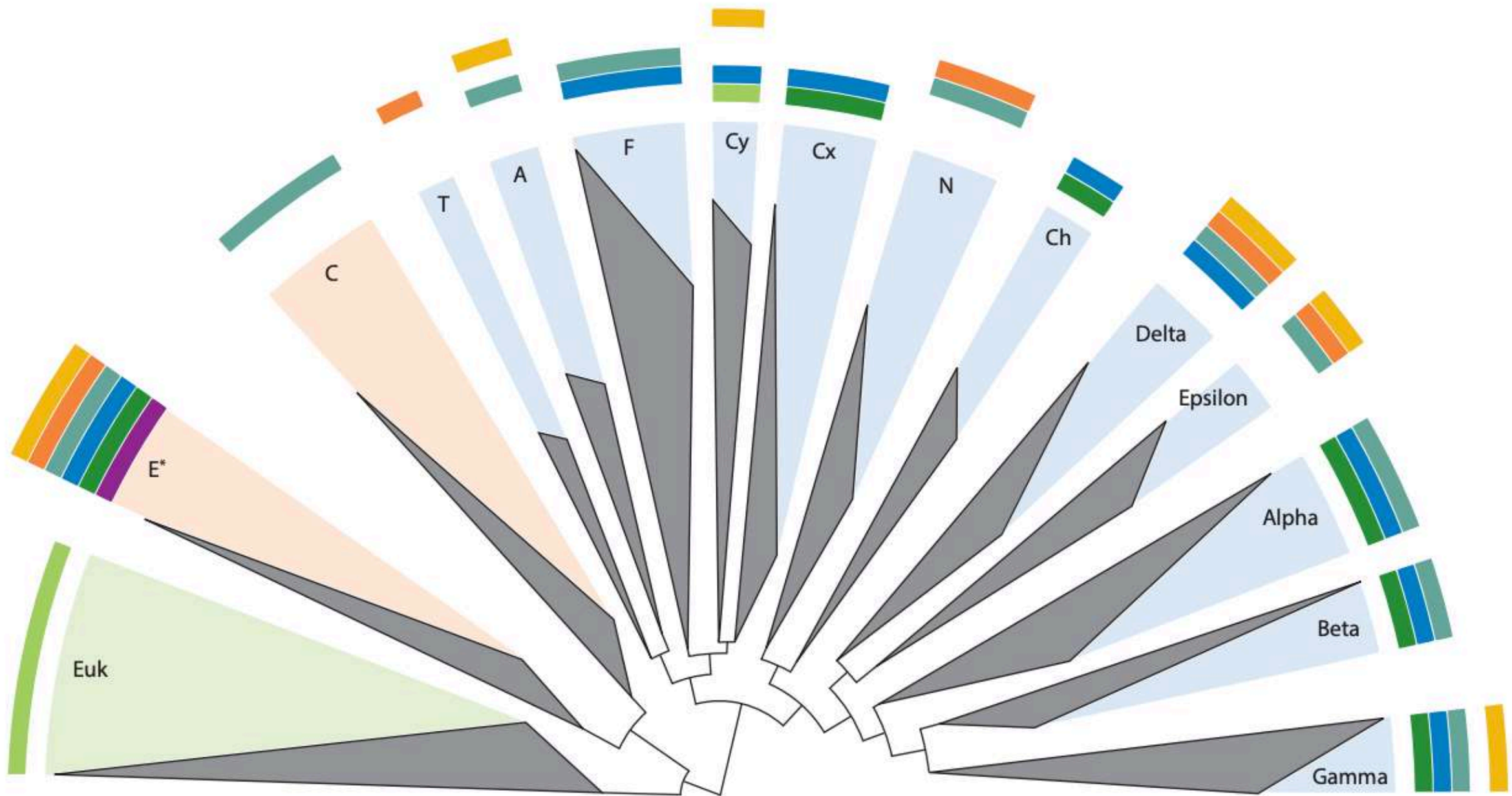


**Figure 1**

Distribution of selected metabolic pathways on the 16S rRNA tree of life. The tree (constructed with ARB; 104) was edited for clarity and shows selected bacterial and archaeal taxa. The area of each branch is proportional to the total number of 16S rRNA sequences present in the database. Metabolic pathways were assigned based on physiological data (**Supplemental Table 2**). Sulfate reduction includes sulfite and thiosulfate reduction pathways. \**Euryarcheota* are capable of bacteriorhodopsin-based photosynthesis only. Abbreviations: A, *Aquificae*; Alpha, *Alphaproteobacteria*; Beta, *Betaproteobacteria*; C, *Crenarchaeota*; Ch, *Chlorobi*; Cx, *Chloroflexi*; Cy, *Cyanobacteria*; Delta, *Deltaproteobacteria*; E, *Euryarchaeota*; Epsilon, *Epsilonproteobacteria*; Euk, *Eukarya*; F, *Firmicutes*; Gamma, *Gammaproteobacteria*; N, *Nitrospirae*; T, *Thermodesulfobacteria*.



# Microbial diversity and metabolic pathways to survive in the environment



## Metabolic pathway

- Sulfur reduction
- Sulfate reduction
- Denitrification
- Nitrogen fixation

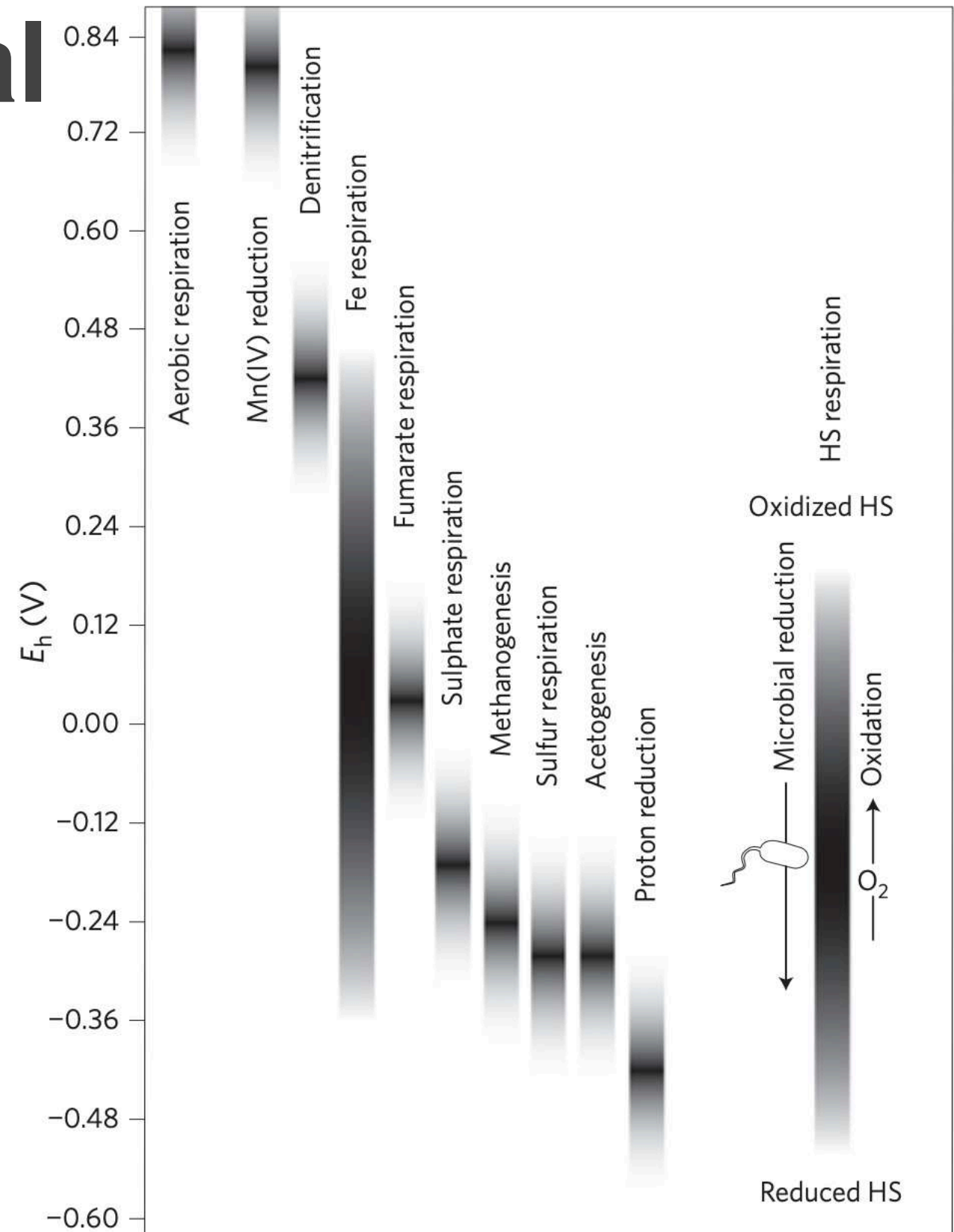
- Anoxygenic photosynthesis
- Oxygenic photosynthesis
- Methanogenesis

## Domains

- Eukarya
- Bacteria
- Archaea

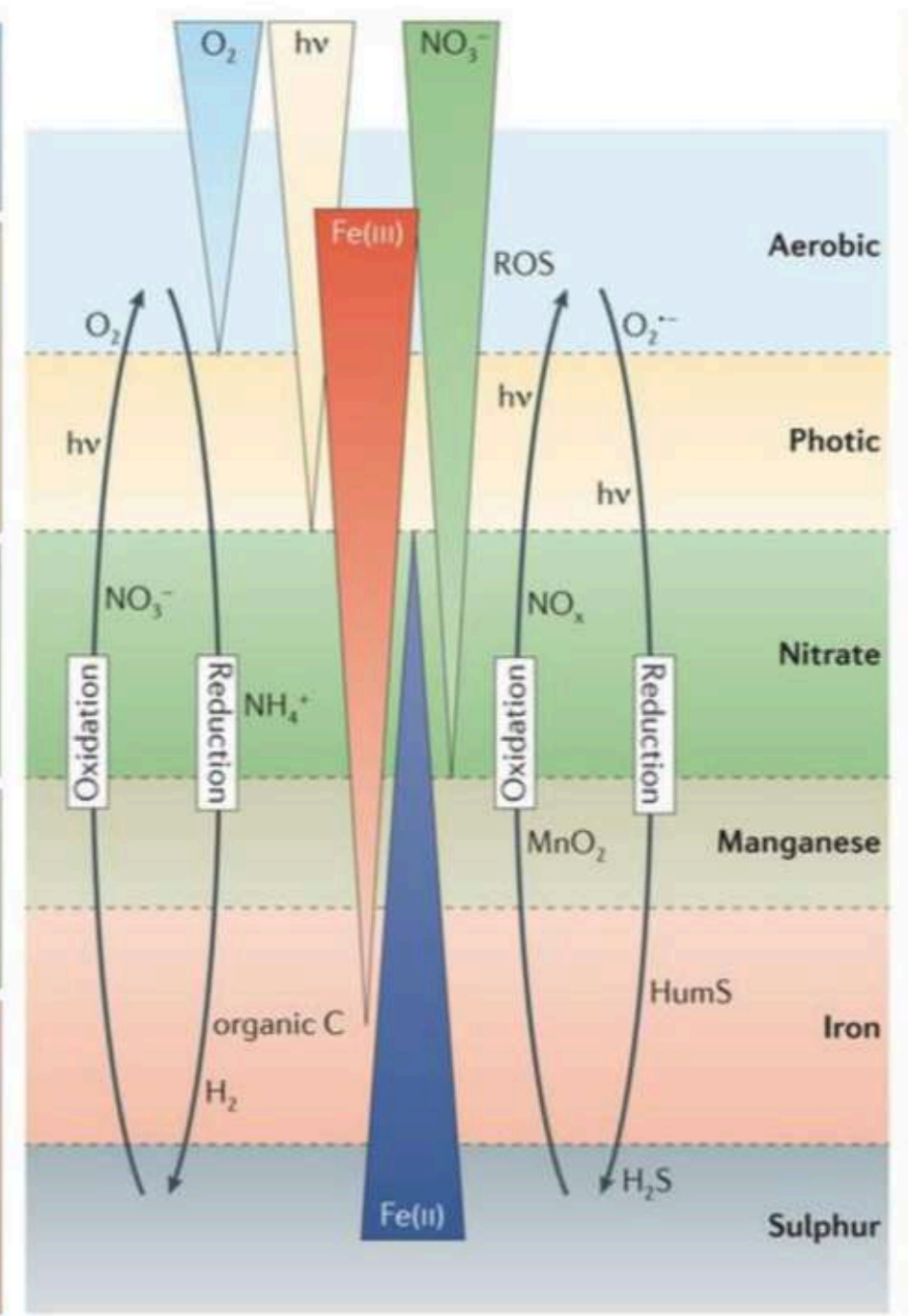
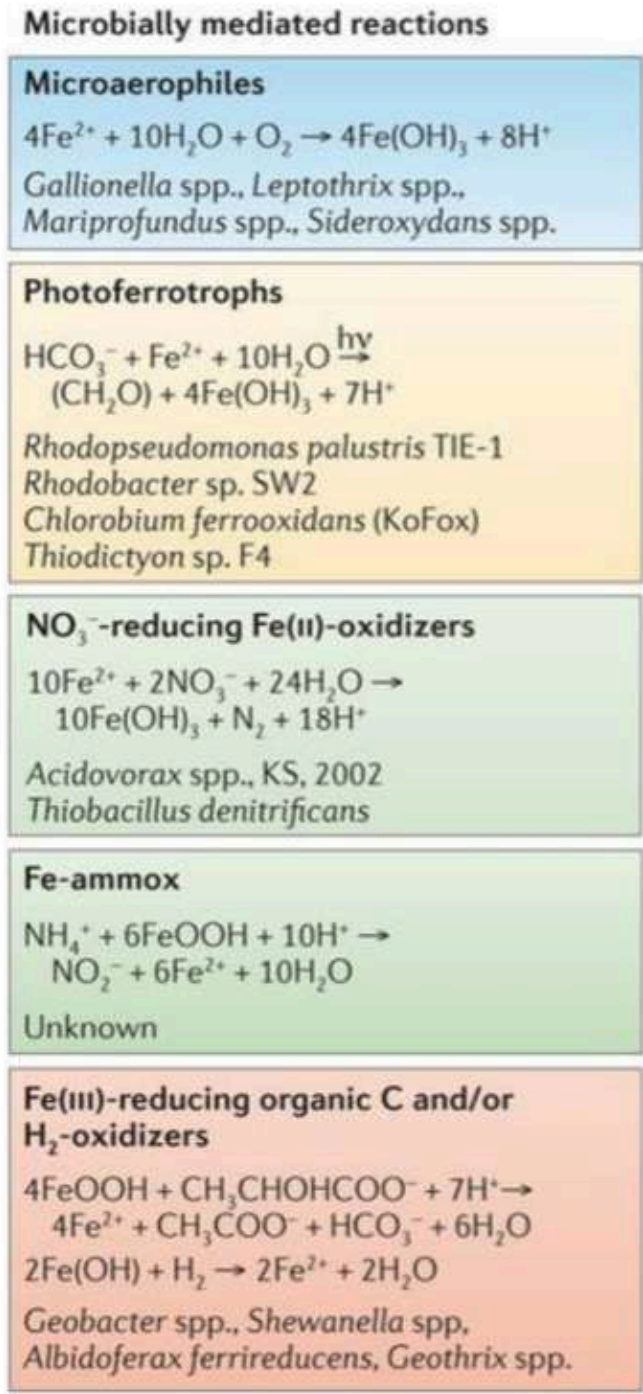
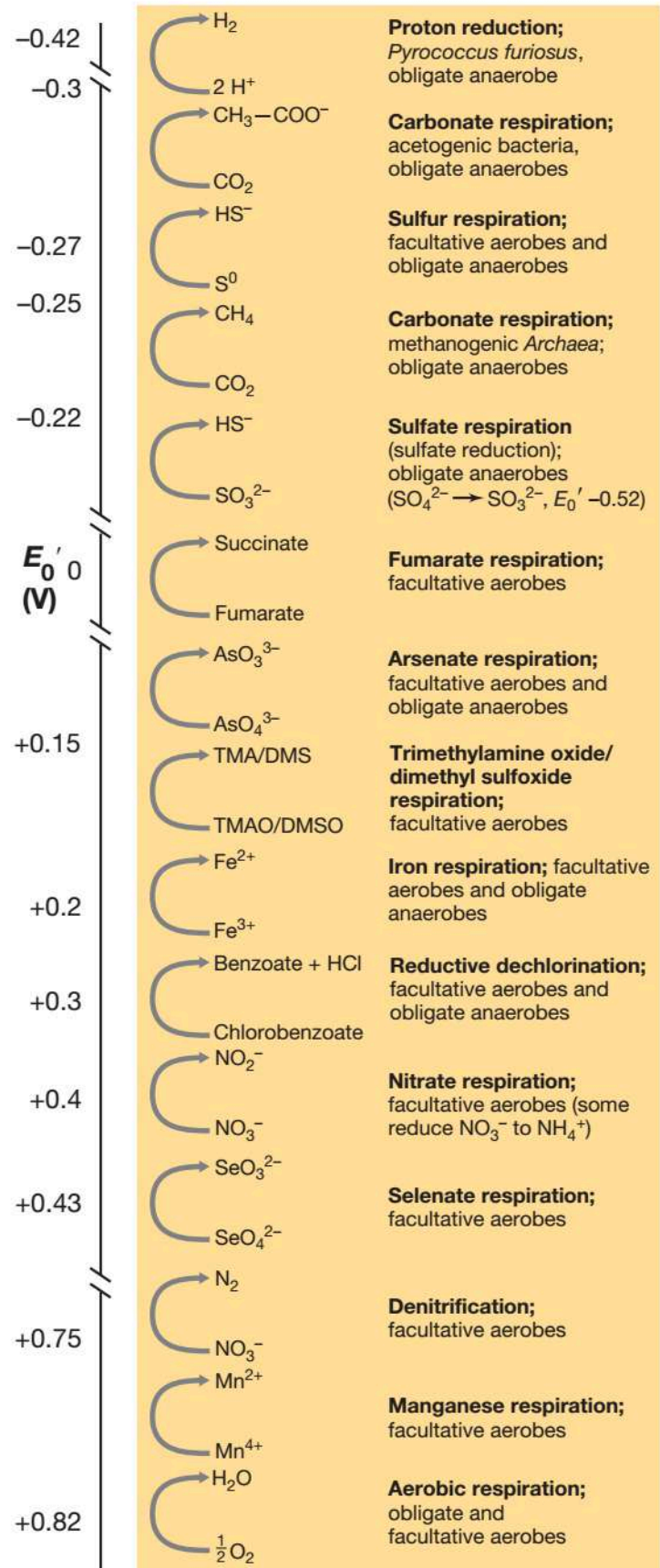
# Reduction potential ranges of microbial respiration

- The achievable energy yield of ETC depends on the difference in electrical potential between electron donor and acceptor
- Microbes able to respire in multiple ways will always choose available acceptors with the **biggest potential difference** to the donor (e.g., *E. coli*  $O_2 > NO_3^- > \text{fumarate}$ )





# Anaerobic respiration





# Fermentation/*Respiration*

- Fermentation is a form of anaerobic catabolism in which organic compounds both donate electrons and accept electrons, and redox balance is achieved without the need for external electron acceptors
- ATP is made from these energy-rich compounds by substrate-level phosphorylation, a process whereby the energy-rich phosphate bond on the organic compound is transferred directly to ADP to form ATP
- Glucose fermentation into alcoholic or lactic acid: 2 ATP
- *Respiration is a form of aerobic or anaerobic catabolism in which an organic or inorganic electron donor is oxidized with O<sub>2</sub> (in aerobic respiration) or some other compounds (in anaerobic respiration) functioning as electron acceptors*
- *ATP is made by PMF*
- *Glucose aerobic respiration into CO<sub>2</sub>: 38 ATP*

# Fermentation, II

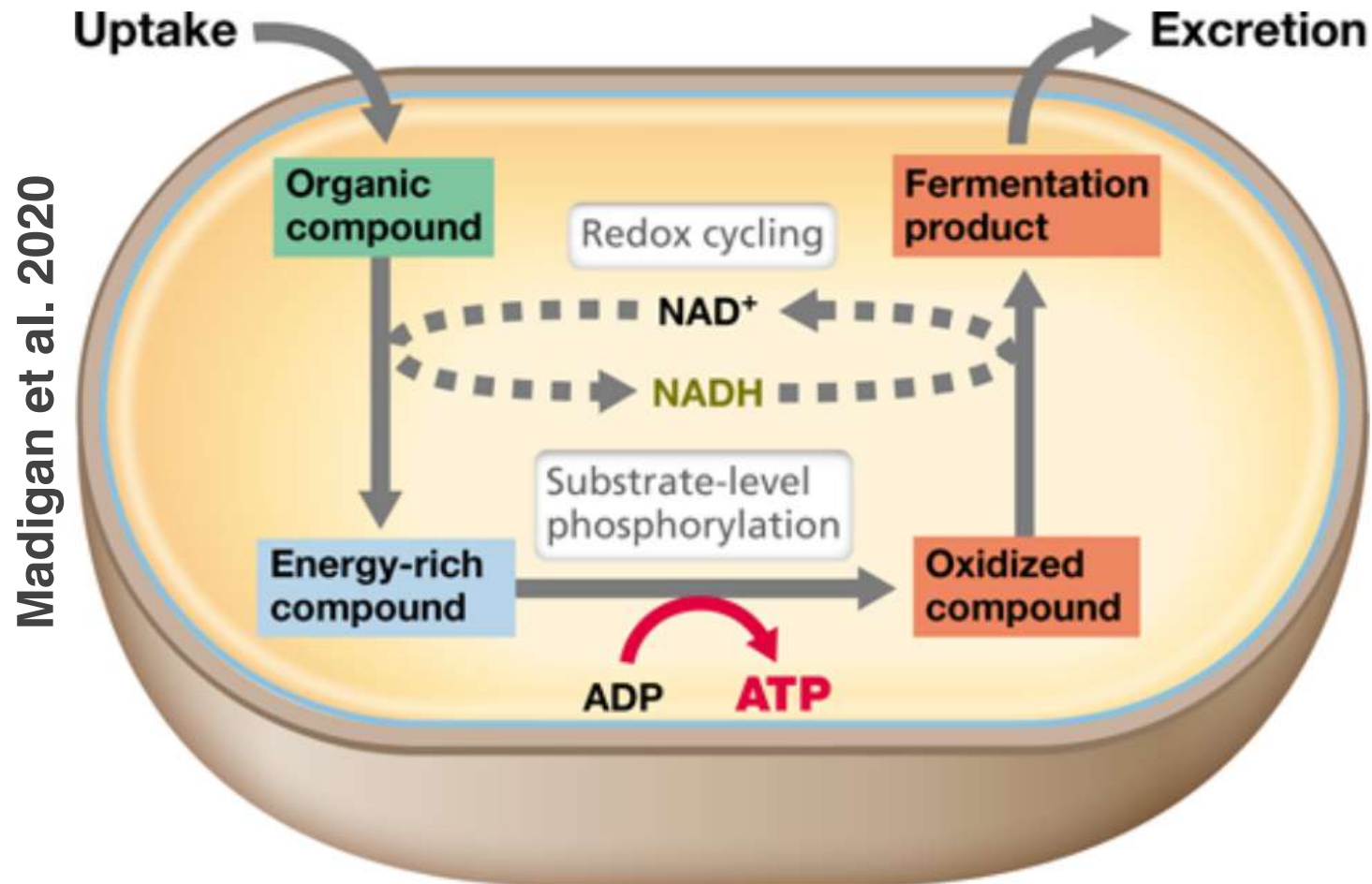
**TABLE 3.4** Common fermentations and some of the organisms carrying them out

Type	Reaction (substrate → products)	Organisms
Alcoholic	Hexose <sup>a</sup> → 2 ethanol + 2 CO <sub>2</sub>	Yeast, <i>Zymomonas</i>
Homolactic	Hexose → 2 lactate <sup>-</sup> + 2 H <sup>+</sup>	<i>Streptococcus</i> , some <i>Lactobacillus</i>
Heterolactic	Hexose → lactate <sup>-</sup> + ethanol + CO <sub>2</sub> + H <sup>+</sup>	<i>Leuconostoc</i> , some <i>Lactobacillus</i>
Propionic acid	3 Lactate <sup>-</sup> → 2 propionate <sup>-</sup> + acetate <sup>-</sup> + CO <sub>2</sub> + H <sub>2</sub> O	<i>Propionibacterium</i> , <i>Clostridium propionicum</i>
Mixed acid <sup>b,c</sup>	Hexose → ethanol + 2,3-butanediol + succinate <sup>2-</sup> + lactate <sup>-</sup> + acetate <sup>-</sup> + formate <sup>-</sup> + H <sub>2</sub> + CO <sub>2</sub>	Enteric bacteria including <i>Escherichia</i> , <i>Salmonella</i> , <i>Shigella</i> , <i>Klebsiella</i> , <i>Enterobacter</i>
Butyric acid <sup>c</sup>	Hexose → butyrate <sup>-</sup> + 2 H <sub>2</sub> + 2 CO <sub>2</sub> + H <sup>+</sup>	<i>Clostridium butyricum</i>
Butanol <sup>c</sup>	2 Hexose → butanol + acetone + 5 CO <sub>2</sub> + 4 H <sub>2</sub>	<i>Clostridium acetobutylicum</i>
Caproate/Butyrate	6 Ethanol + 3 acetate <sup>-</sup> → 3 butyrate <sup>-</sup> + caproate <sup>-</sup> + 2 H <sub>2</sub> + 4 H <sub>2</sub> O + H <sup>+</sup>	<i>Clostridium kluyveri</i>
Acetogenic	Fructose → 3 acetate <sup>-</sup> + 3 H <sup>+</sup>	<i>Clostridium aceticum</i>

- Not all compounds are inherently fermentable, but sugars (e.g. glucose, other hexoses, most disaccharides, other relatively small sugars) —are fermentable
- Polysaccharides (e.g. cellulose, starch, chitin) are also fermentable by bacteria that produce enzymes that attack these large molecules and produce sugars from them if the latter are not glucose, they must first be converted to glucose before they enter glycolysis
- 2 net ATP molecules in glycolysis
- More ATP synthesis by substrate-level phosphorylation if fatty acid because the fatty acid is formed from its coenzyme-A precursor (energy-rich molecules)

# Fermentation

Figure 3.14 The essentials of fermentation.



- Both organic compounds accept and donate e-
- No need to external e-acceptor to achieve balance

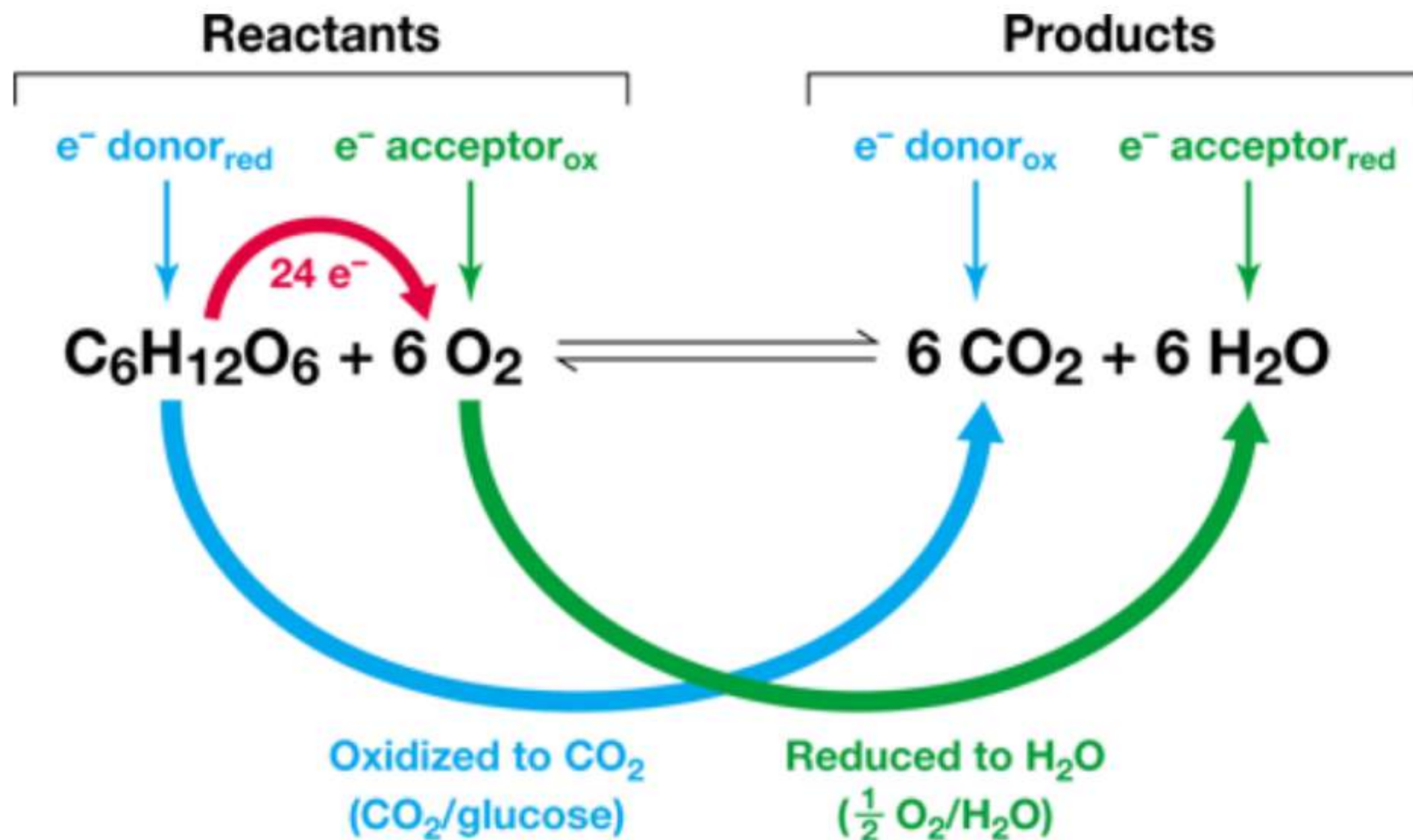
- An organic compound is oxidized
- e- are recycled back to one of the oxidized organic products because an external e-acceptor is lacking
- Product is excreted from the cell and ATP is produced by substrate-level phosphorylation



# Fundamentals in Metabolisms

- Transfer e<sup>-</sup> and conserve energy
- Reactions are not performed in single-step → consecutive reactions in different part of the cells
- Need of soluble e<sup>-</sup> carriers: NAD<sup>+</sup>/NADH, FAD<sup>+</sup>/FADH<sub>2</sub>

Madigan et al. 2020

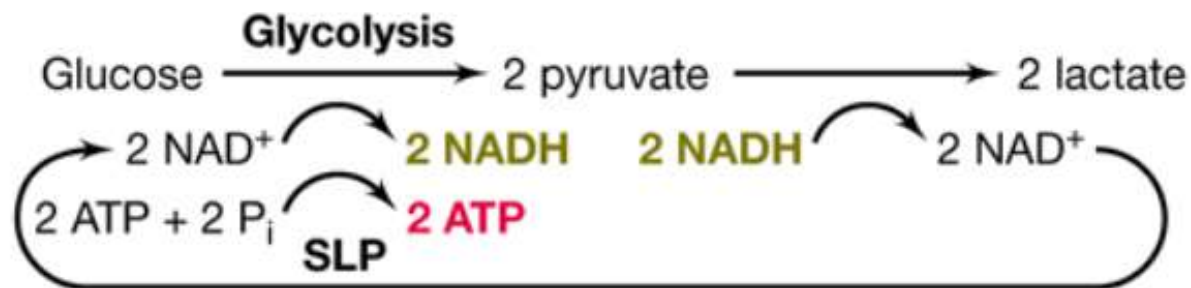


# Substrate-Level-Phosphorylation

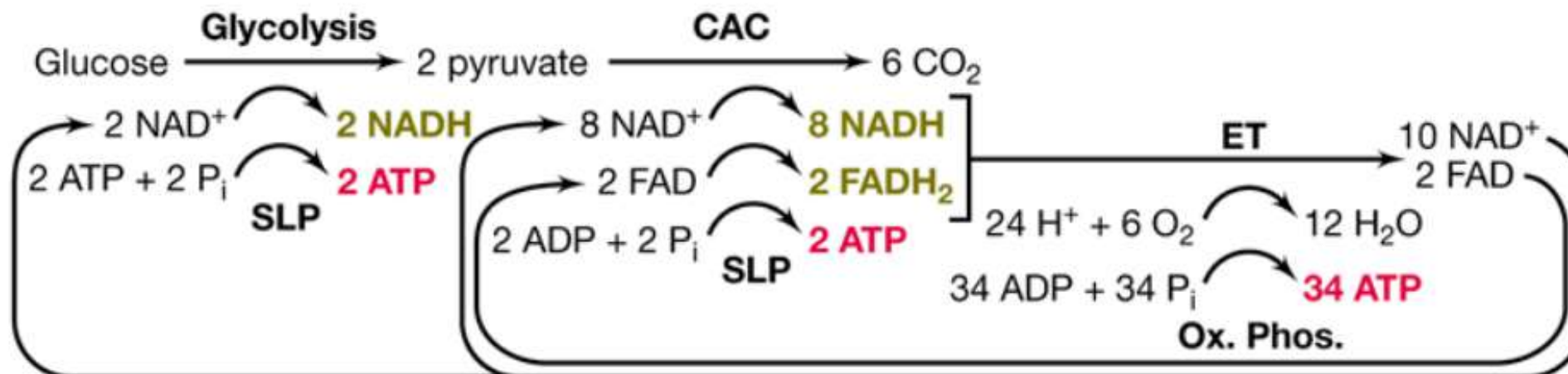
- Glycolysis can generate ATP in the absence of oxygen: anaerobic metabolism
- Glycolysis and citric acid cycle (CAC) result from substrate-level phosphorylation (SLP)
- SLP is distinct from oxidative phosphorylation that occurs in ETC
- Substrate-level phosphorylation refers to the formation of ATP from ADP and a phosphorylated intermediate, rather than from ADP and inorganic phosphate,  $P_i$ , as is done in oxidative phosphorylation (ET)

# Figure 3.21 Energetics in fermentation and aerobic respiration.

## Lactic acid fermentation

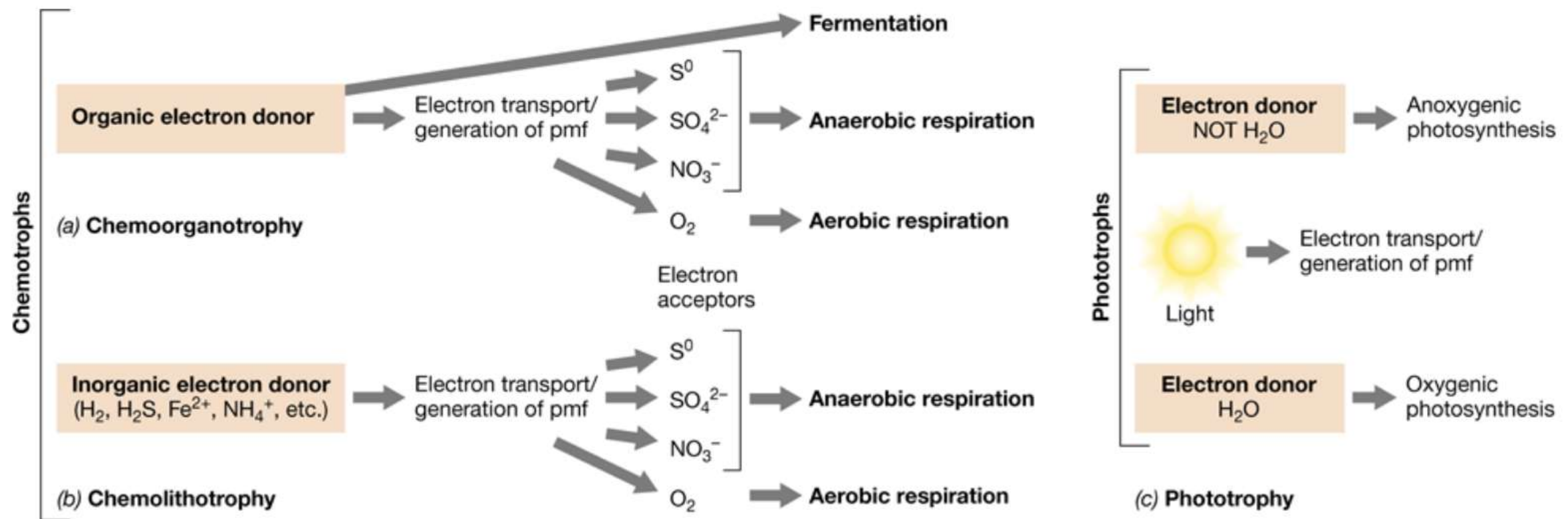


## Aerobic respiration

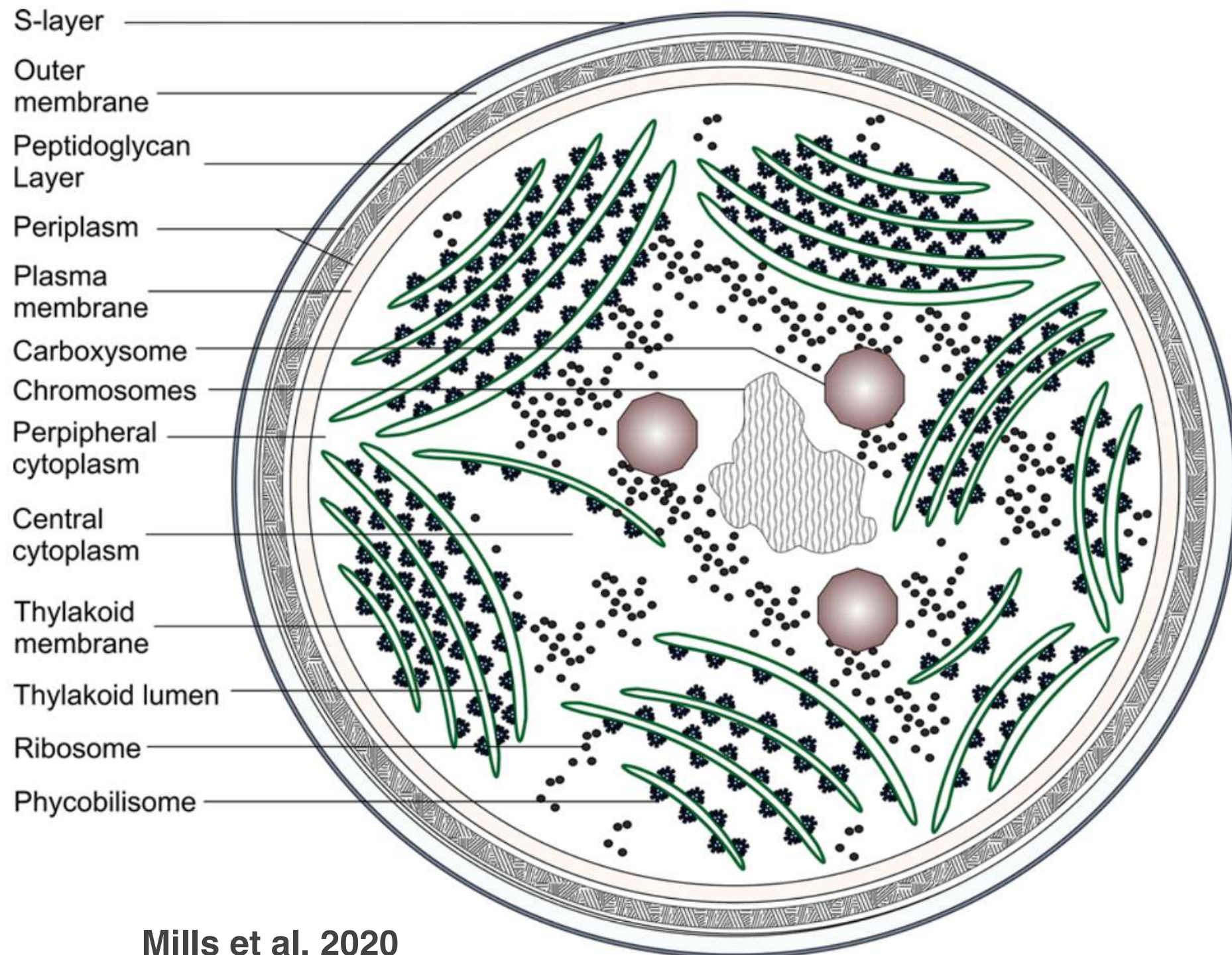




# Figure 3.22 Metabolic diversity and its relationship to oxygen.



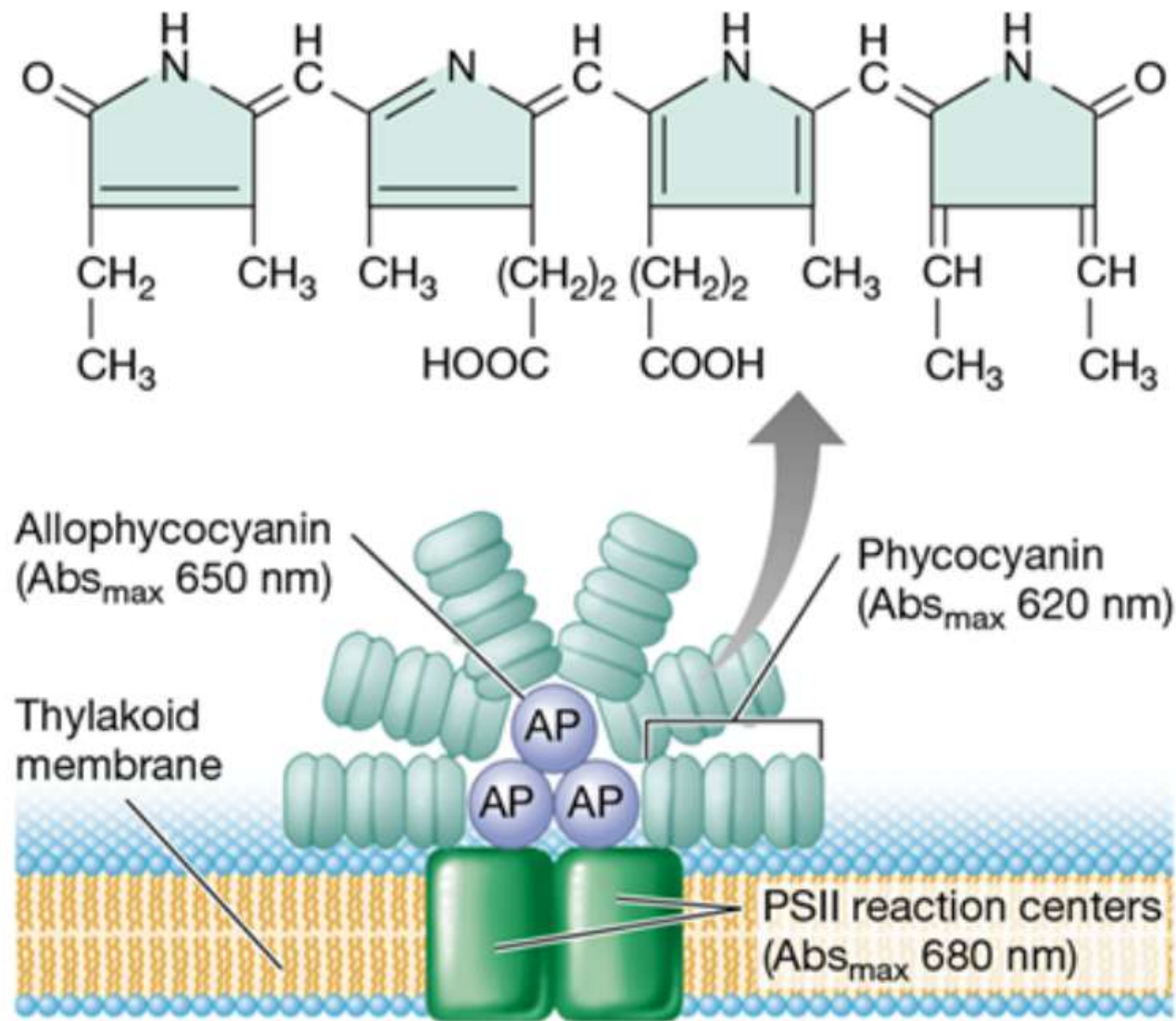
# Photo Synthesis: Calvin–Benson–Bassham



- Carboxysomes are made of polyhedral protein shells about 80 - 140 nm in diameter
- Concentrate carbon dioxide to overcome the inefficiency of RuBisCO (ribulose biphosphate carboxylase/oxygenase)
- RuBisCO predominant enzyme in carbon fixation and the rate limiting enzyme in the Calvin-Benson-Bassham cycle



# Oxygenic photosynthesis

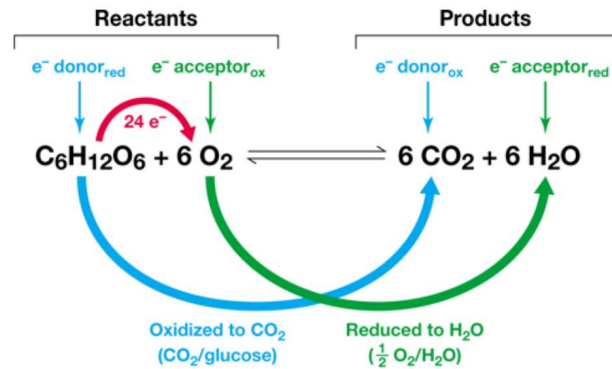


Madigan et al. 2020

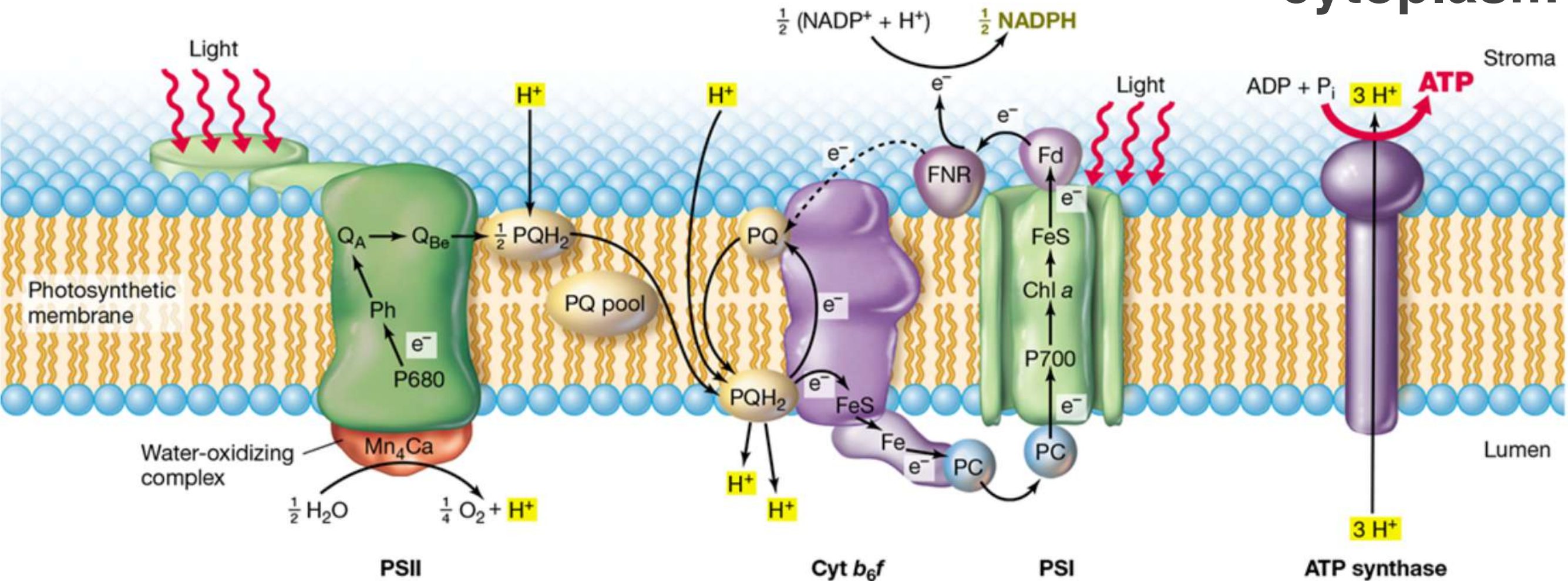
- Physical location within the cell (Cyanobacteria)
- Bilayer w. proteins and complex that capture light, phycobilisome



# Photo



# cytoplasm

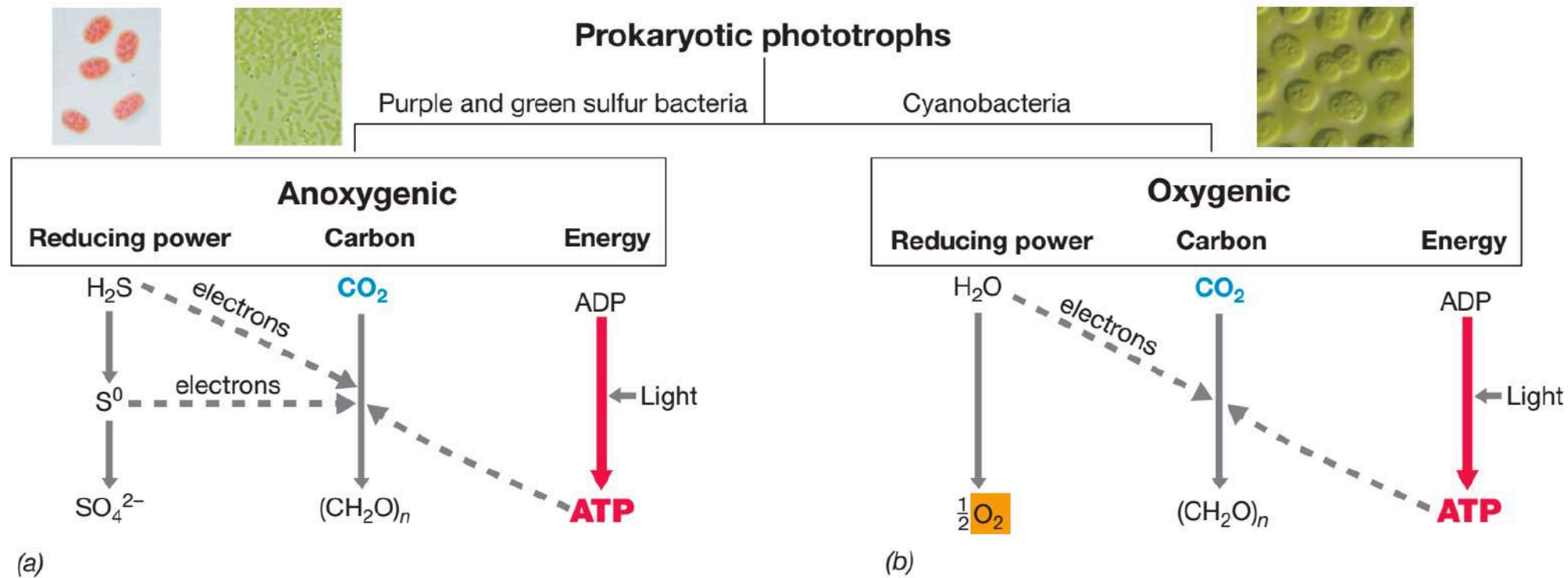


- Splitting of  $\text{H}_2\text{O}$
- Generation  $\text{H}^+$  motive force
- Generation of NADPH → **C fixation (from  $\text{CO}_2$ )** via Calvin–Benson–Bassham cycle
- ATP production

Madigan et al. 2020



# Light driven processes



Madigan et al. 2018

## Winogradsky columns

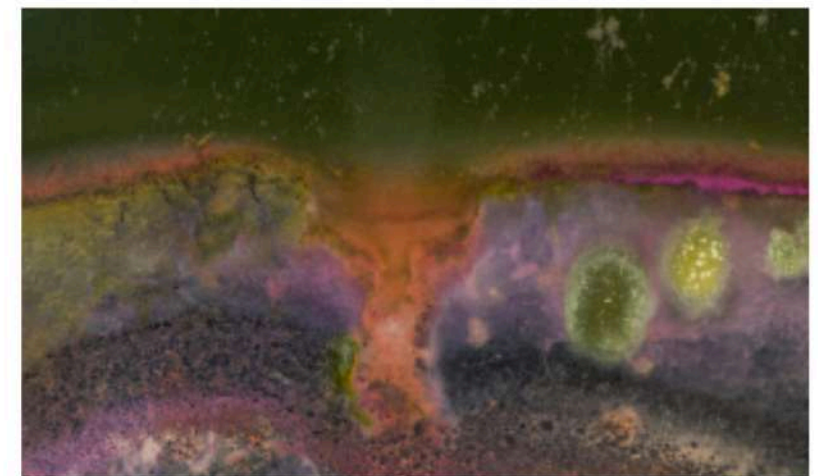
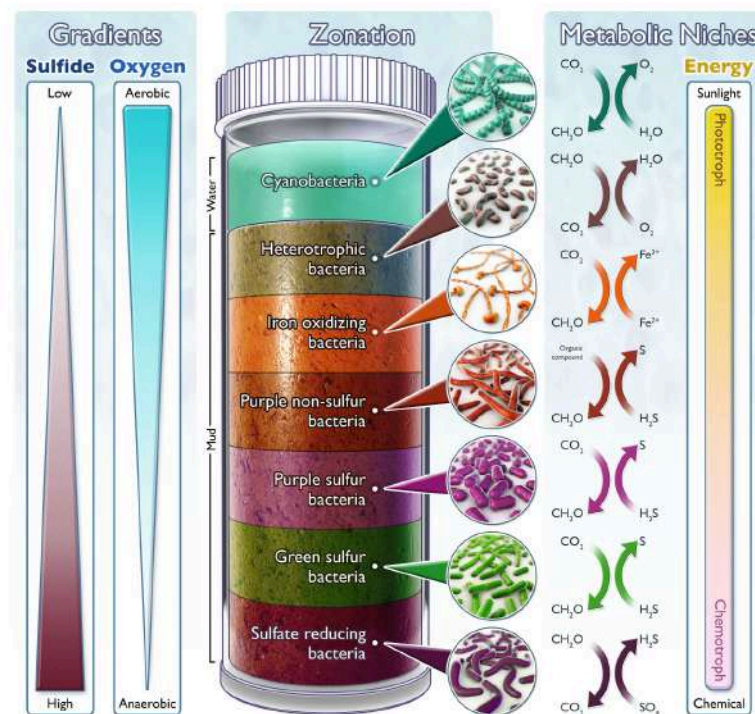


Figure 3. The upper sediment interface on day 15. Aerobic cyanobacteria and algae (upper aqueous phase), yellow-orange microaerophilic iron-oxidizing bacteria, and anaerobic green and purple photosynthetic bacteria develop into layered communities.

# Energy generating metabolic pathways

## •Oxygenic Photosynthesis

ATP and NADPH are made in large amounts

Produces oxygen as a bi-product during splitting of water for reducing power

## •Anoxygenic Photosynthesis

ATP made in large amounts

Reduction of NADP does not involve water; hence no oxygen produced

## •Aerobic Respiration

ATP and NADH are made in abundance

Requires oxygen

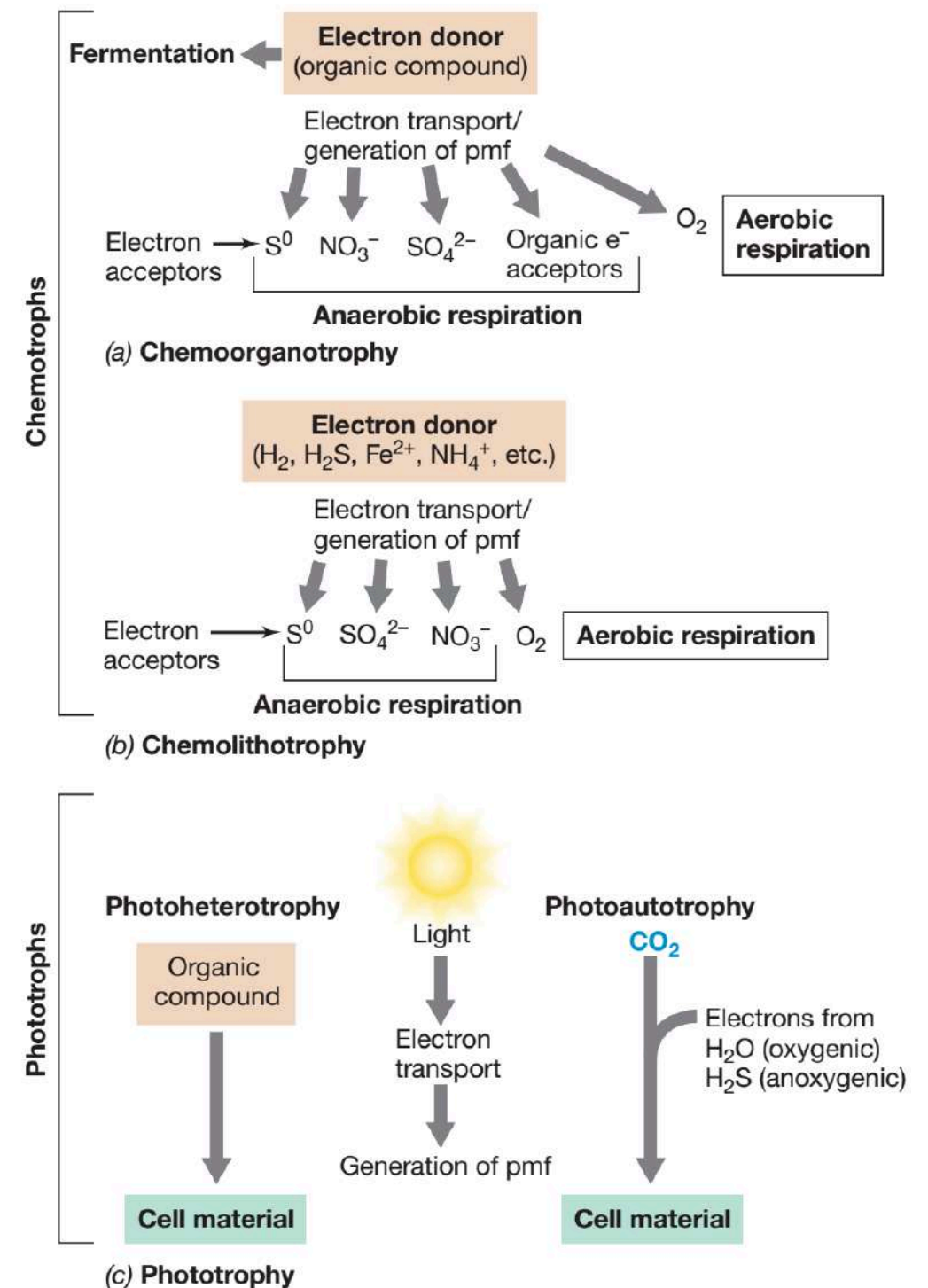
## •Anaerobic Respiration

Lower ATP yield than aerobic respiration; NAD easily reduced

Requires electron acceptor other than oxygen

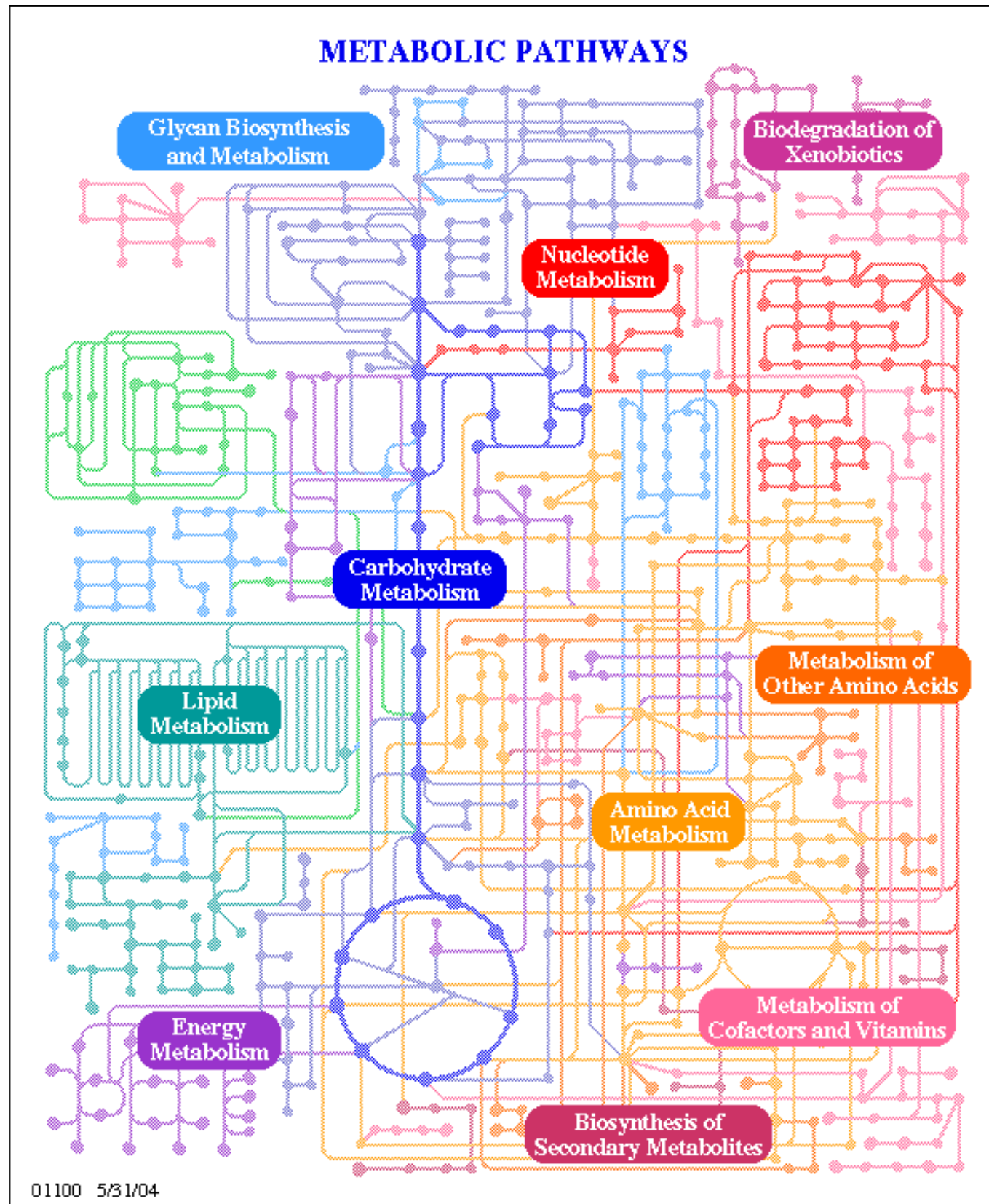
## Fermentation

Little ATP, no net NAD reduction, MOST SIMPLE SYSTEM





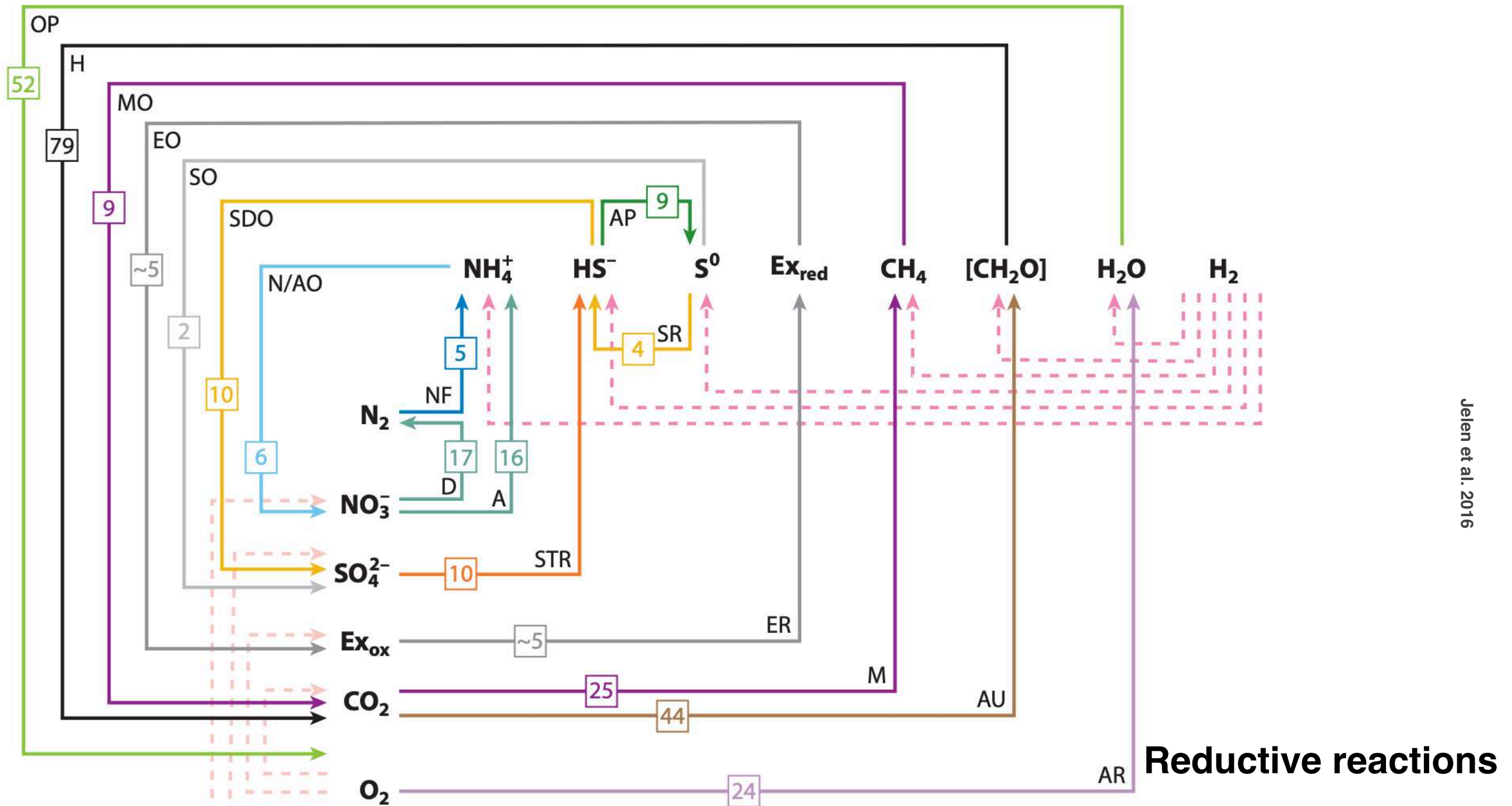
# Integrative approach, I



Metabolic pathways evolved to utilize available substrates produced as end products of other types of microbial metabolism, either by modification of existing metabolic pathways or by using established ones in reverse

# Integrative approach, II

## Oxidative reactions



Jelen et al. 2016

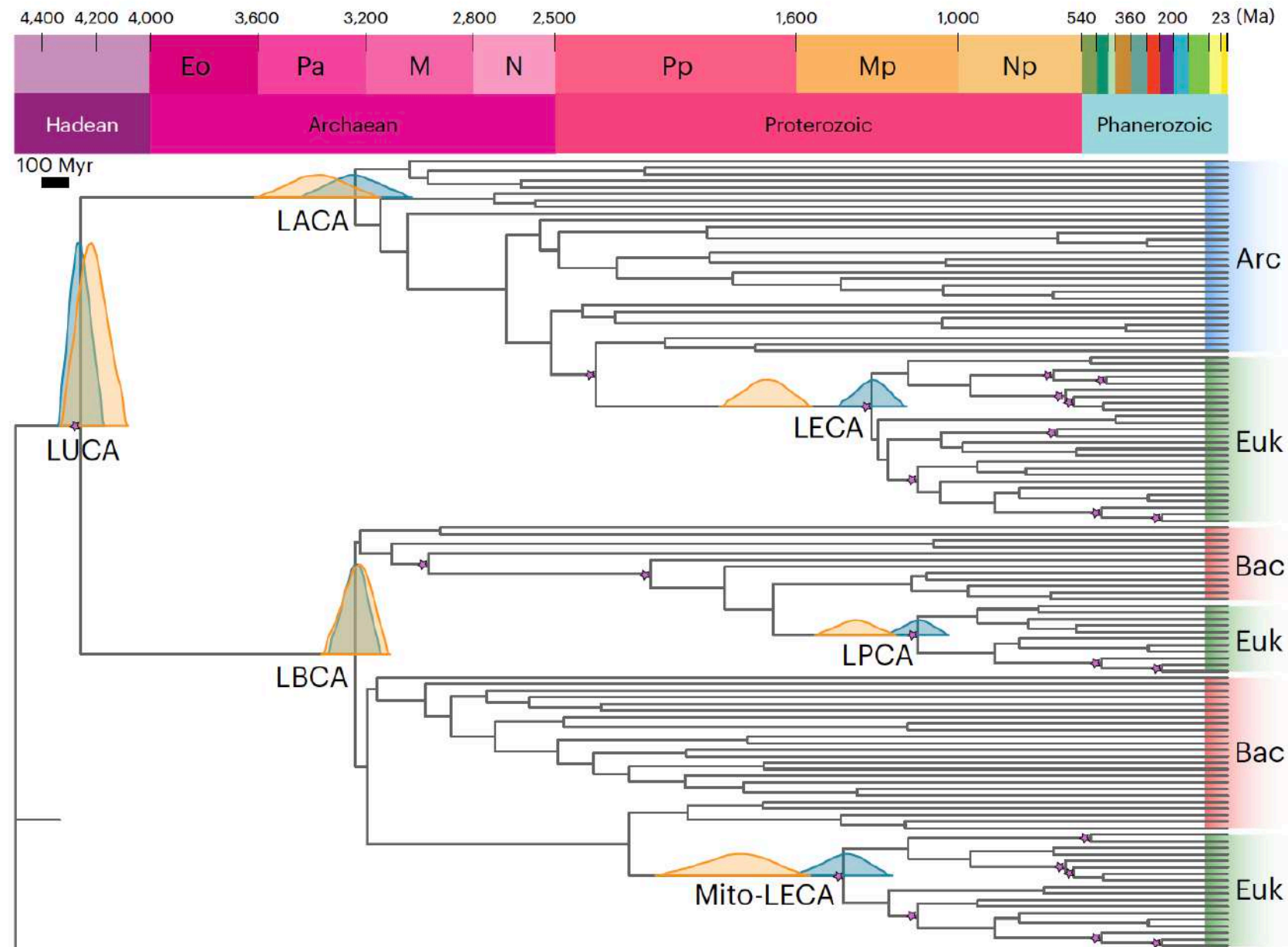
A, ammonification; AP, anoxygenic photosynthesis; AR, aerobic respiration; AU, autotrophy; D, denitrification; Ex<sub>ox</sub>, other elements oxidation; Ex<sub>red</sub>, other elements reduction; H, heterotrophy; M, methanogenesis; MO, methane oxidation/methanotrophy; N/AO, nitrification/ammonia oxidation; NF, nitrogen fixation; OP, oxygenic photosynthesis; SDO, sulfide oxidation; SO, sulfur oxidation; SR, sulfur reduction; STR, sulfate reduction

# Energy conservation

- The achievable energy gain (Gibbs free energy,  $\Delta G$ ) of ETC depends on the redox potential difference ( $\Delta E$ ) of all reactions between electron donor and acceptor
- Microbes able to respire in multiple ways will always choose available acceptors with the biggest potential difference to the donor (e.g., *E. coli*  $O_2 > NO_3^- \rightarrow$  fumarate)
- **Cellular metabolism coordinate the production, management and re-distribution of carbon building blocks and energy (ATP and NADPH) between various electron and carbon sinks**
- ATP and NAD(P)H are **essential energy carriers** for numerous biochemical reactions occurring
- With the exception of fermentation, in which substrate-level phosphorylation occurs all other mechanisms of microbial energy conservation are linked to the proton motive force (or gradient of sodium ions,  $Na^+$ , instead of protons)
- Whether electrons come from the oxidation of organic or inorganic chemicals or are mediated by light-driven processes, in both respiration and photosynthesis, **energy conservation is the result of electron transport reactions and the formation of a PMF  $\rightarrow$  ATP**
- **The oxidation of NADH and FADH, to  $NAD^+$  and FAD, respectively, is linked to energy conservation via ETC**

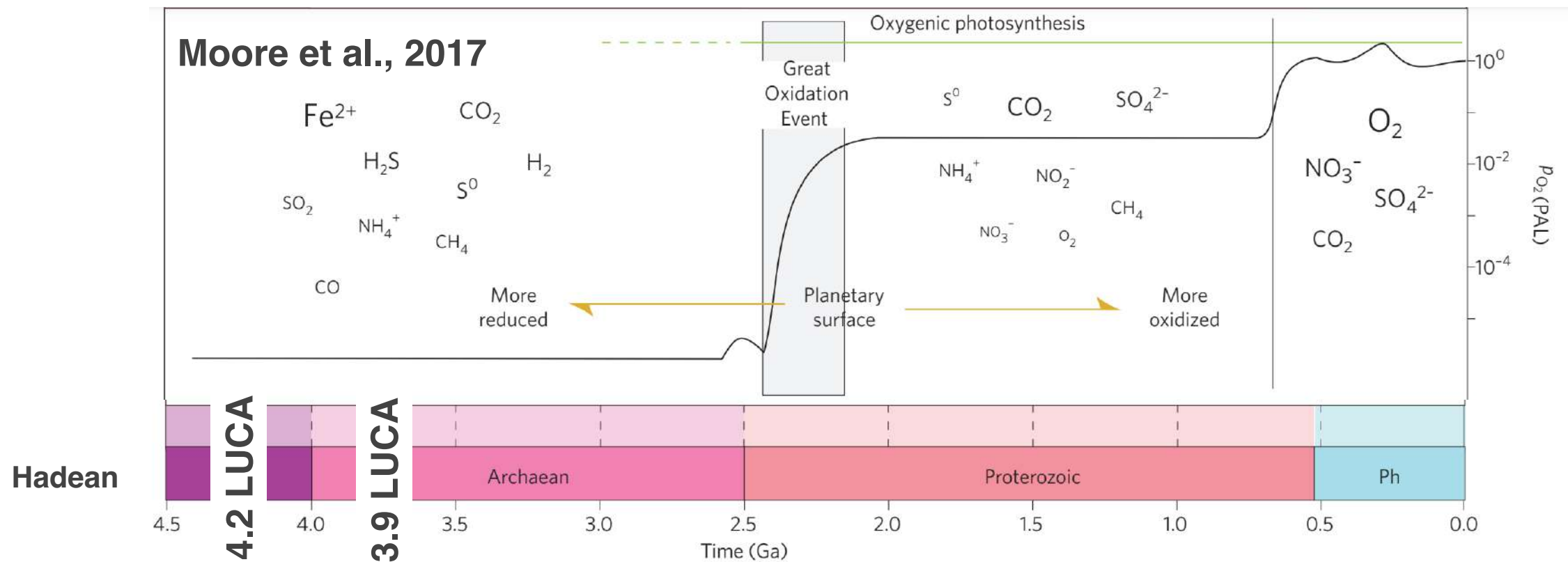


Inference: LUCA lived ~4.2 Ga (4.09–4.33 Ga) through divergence time analysis of pre-LUCA gene duplicates, calibrated using microbial fossils and isotope records under a new cross-bracing implementation



Moody et al., 2024

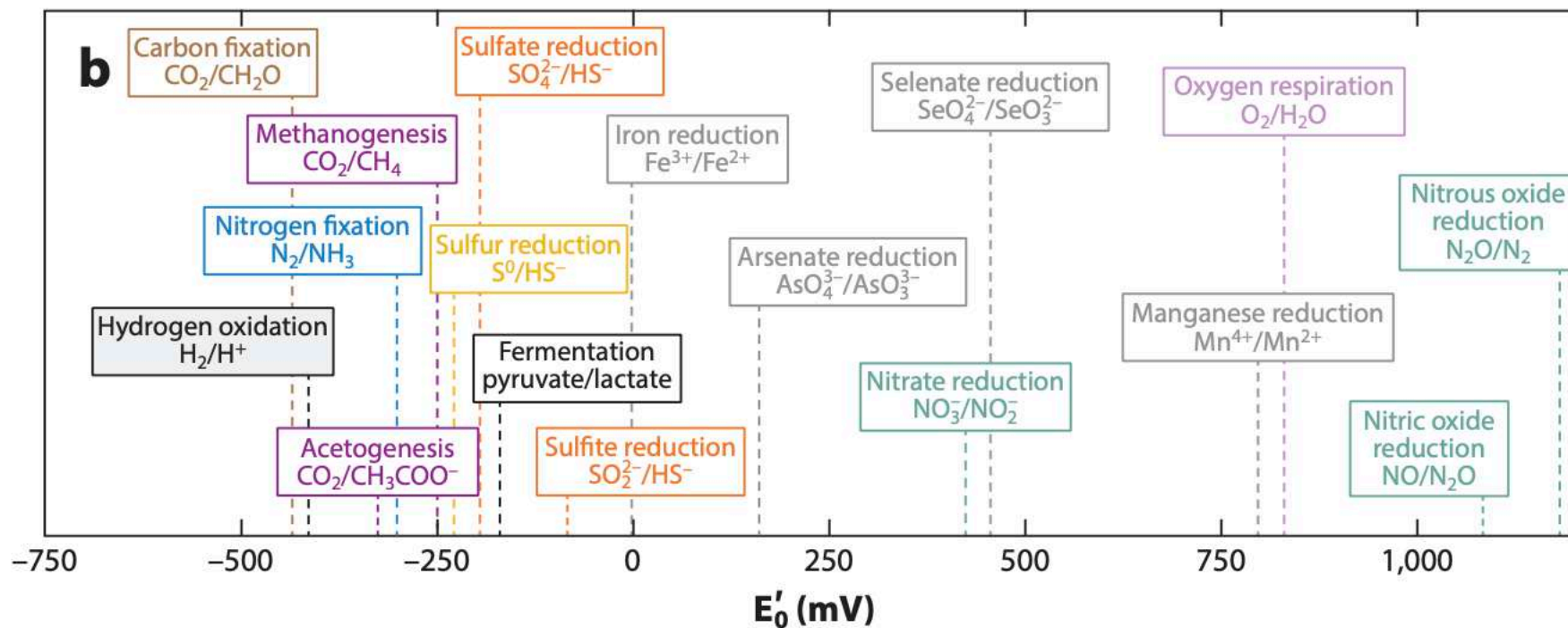
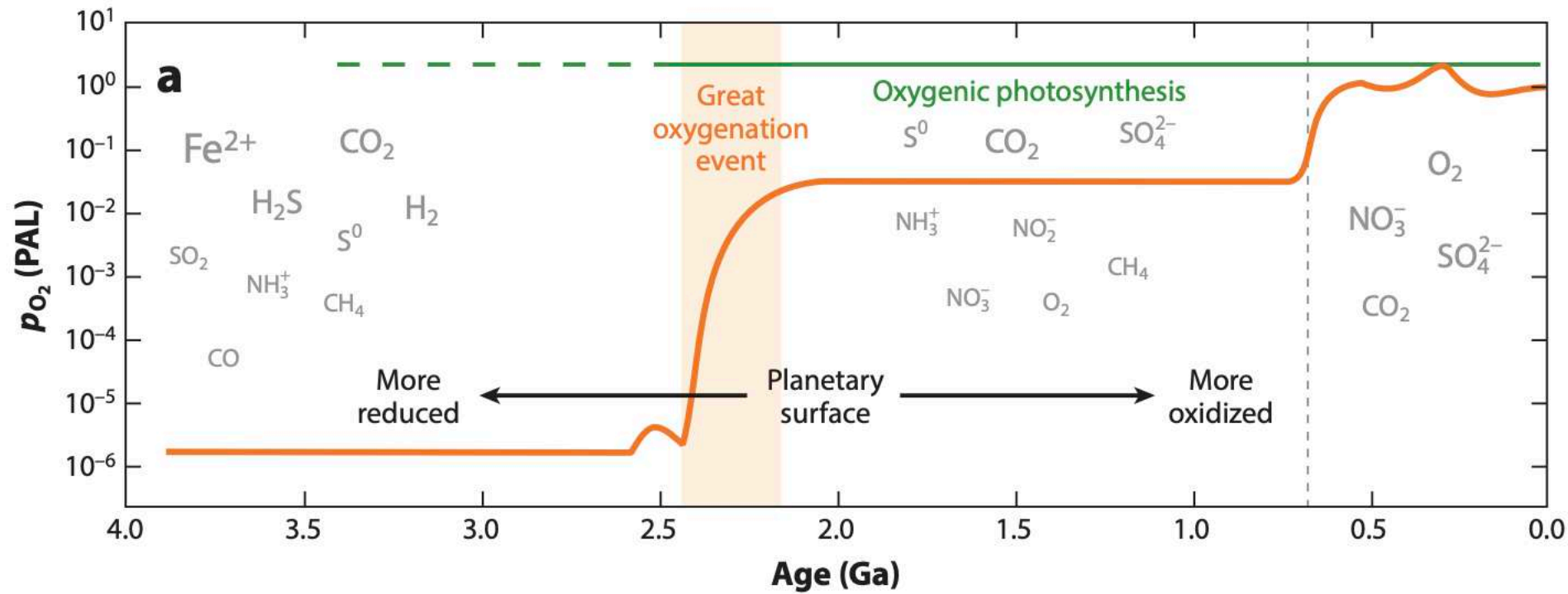
# Earth redox state changes



The availability of different metals and substrates has changed over the course of Earth's history as a result of secular changes in redox conditions of the mantle

Solar energy used by early microbes

# Coevolution of geosphere and biosphere through time as depicted by change in planetary redox state, availability of redox couples



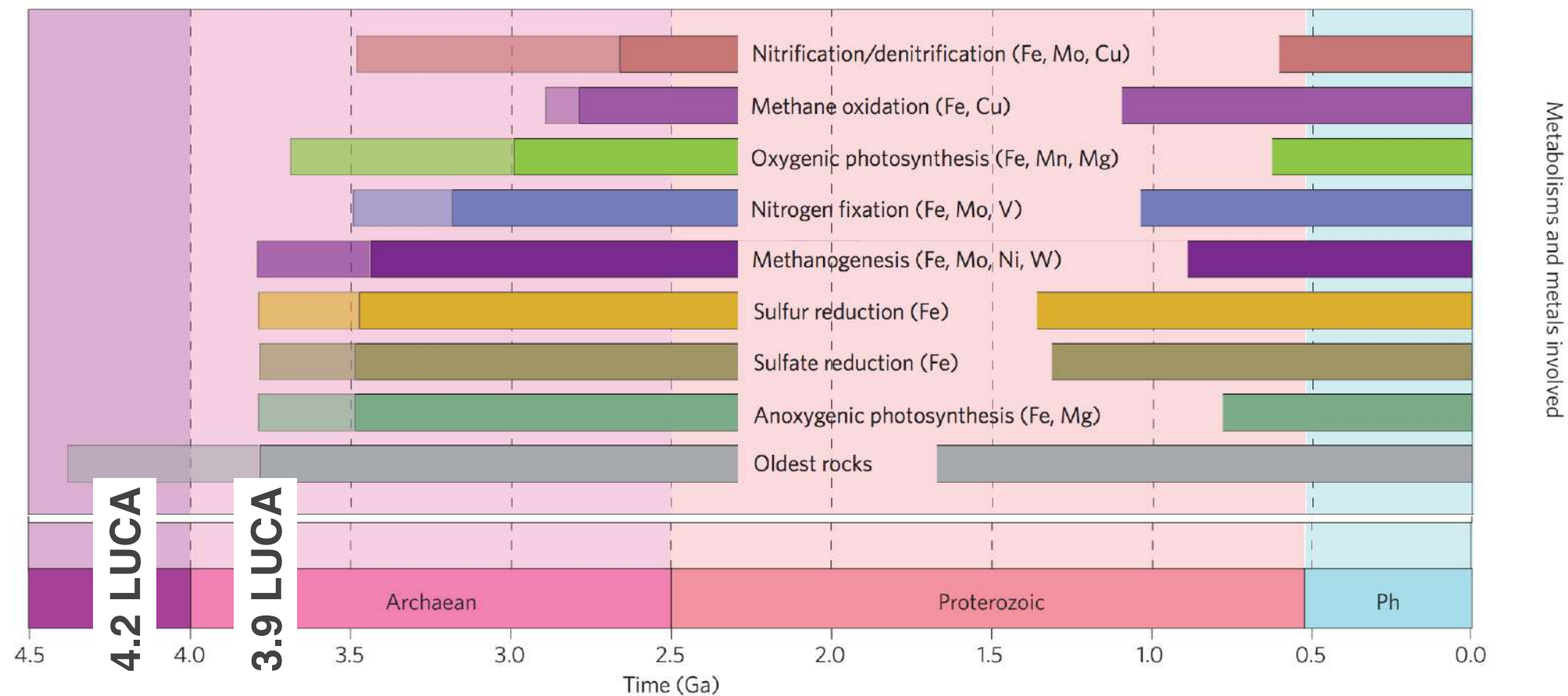
Standard reduction potential at pH 7 ( $E'_0$ ) of biologically relevant redox pairs. Redox halfreactions represent the reductive side (i.e., terminal electron acceptor) of given pathways

Jelen et al., 2016



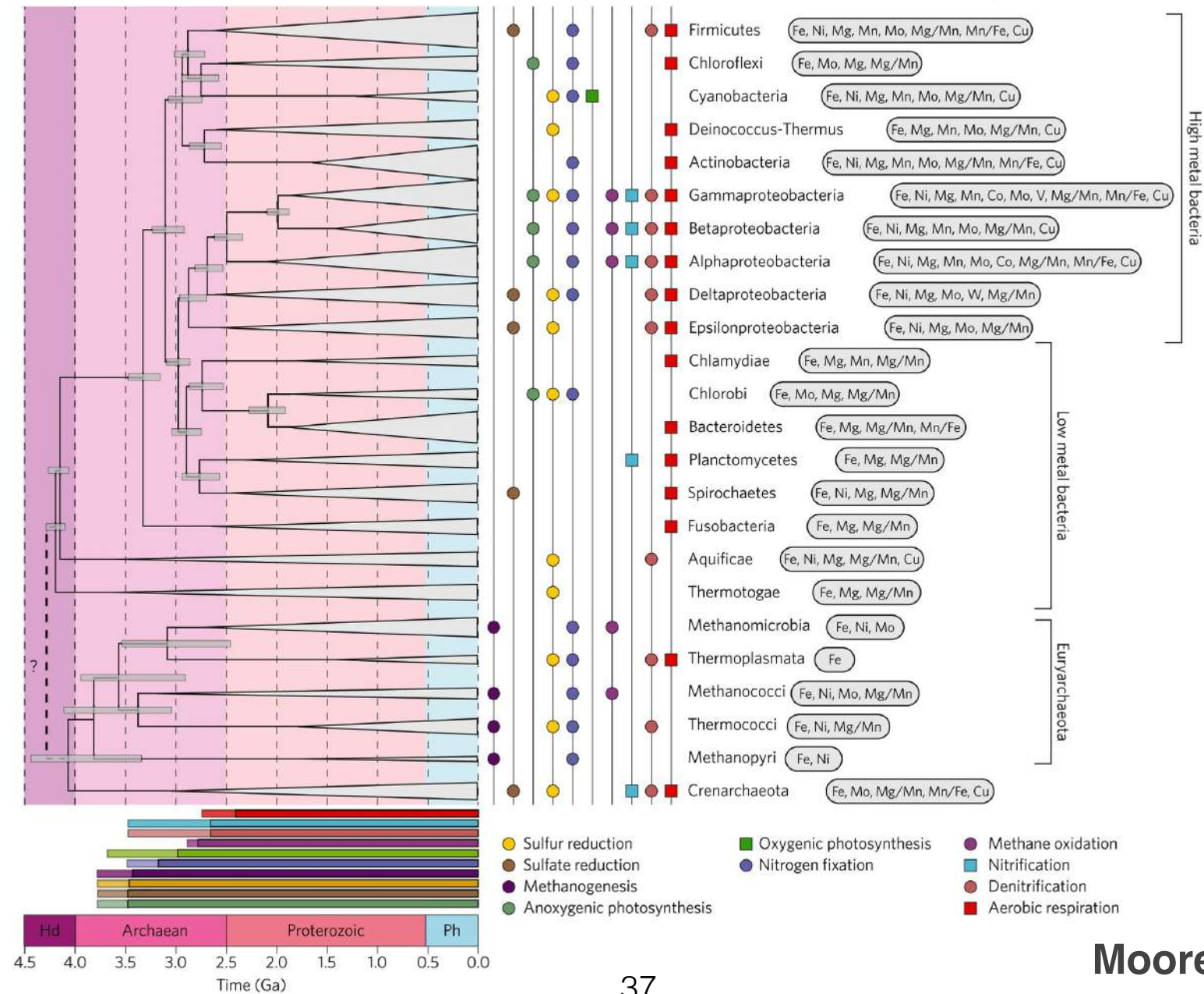
# Emerging microbial metabolisms

Moore et al., 2017



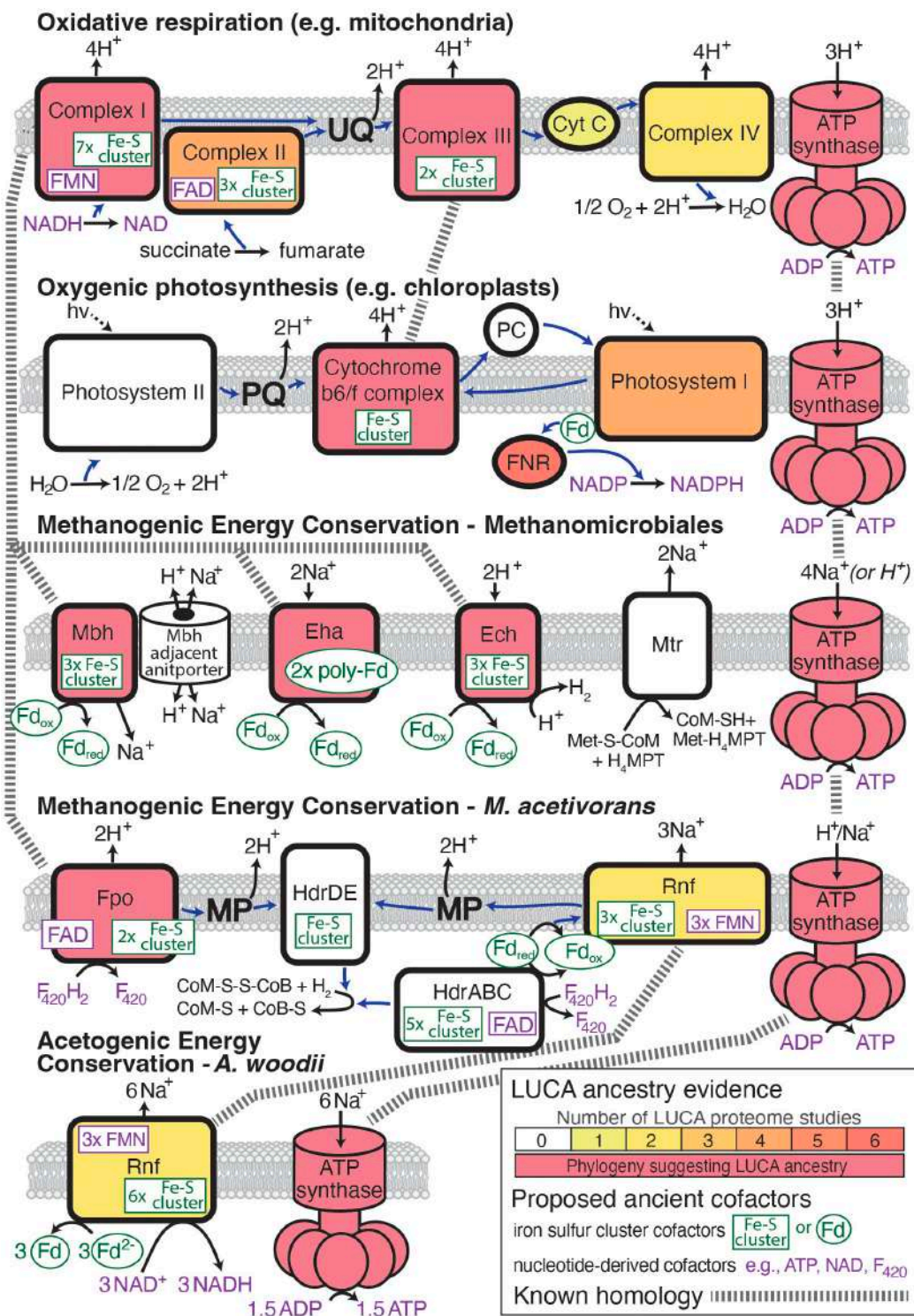
**The oxidoreductases responsible for these metabolisms incorporated metals that were readily available in Archean oceans: iron and iron–sulfur clusters**

# Phylogenetic tree of the main lineages of Bacteria and Archaea and their putative divergence times



Moore et al., 2017

# Rewiring of exhibiting membrane-associated micromachies



## Electron transport chains as a window into the earliest stages of evolution

Signatures of early evolution across different types of chemiosmotic energy conservation.

Electron flow is shown as blue arrows.

Likely ancestry from the LUCA is reflected by either direct phylogenetic evidence or the number of different LUCA proteome studies (out of eight total) that predict a component of the complex to be descended from the LUCA.

Protein cofactors that are potential relics of prebiotic mineral catalysis or ribozyme catalysts are highlighted in green and purple, respectively.

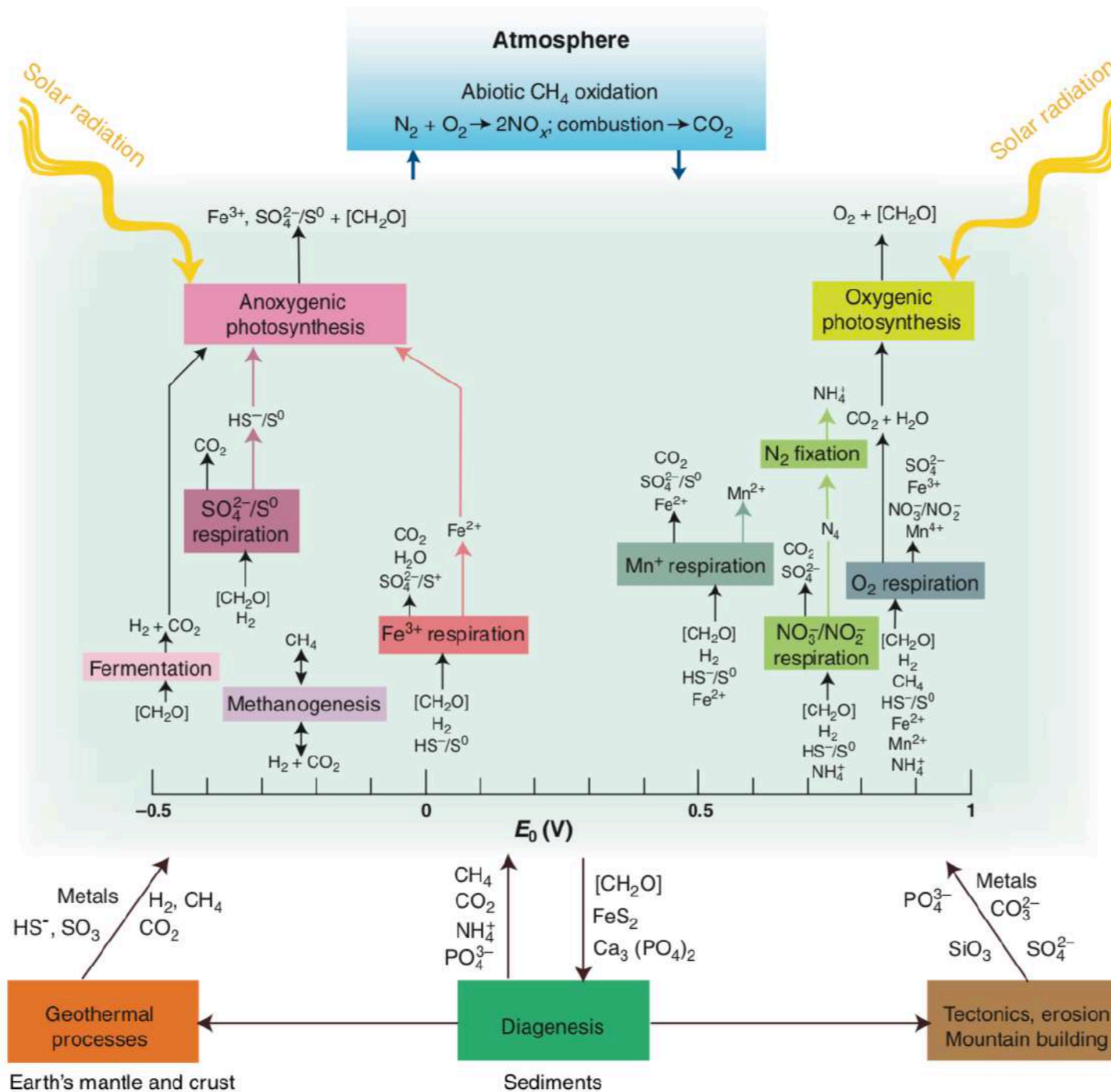
Homology across different ETC components is indicated by a dashed line.

Electron carrier proteins that are components of ETC complexes such as cytochrome B are not shown.



# Biosphere model of energy fluxes and elemental cycles

Falkowski, Fenchel and Delong, 2008



**Microbial microscale actions structure planet-scale functioning**