



Extracellular vesicles: grounds for hope in disease diagnostics and therapy





Extracellular Vesicles (EVs)





Small EVs in cancer





EVs: challenges





Margolis & Sadovsky, PLOS biology 2019





Exploring the role of small EVs in cancer spreading

- Role of pathological s-EVs in modulating physical properties of cells
- Uptake routes and dynamics: model membrane systems





Part I: s-EVs in metastatic cancer



Albritton et al. 2018, Dis Model Mech.

From literature:

- metastatic potential of cancer cells requires specific biomechanical properties
- metastatic cancer cells transfer oncogenic proteins, mRNAs, and miRNAs to target cells through EVs

Questions:

- are s-EVs regulators of cell biomechanical changes?
- which type of phenotypic changes do they induce?

During metastasis cancer cells undergo different phenotypic changes

Cell rearrangements: cytoskeleton, nuclear morphology, focal adhesion





s-EVs from breast cancer cells



Triple negative breast cancer (TNBC): aggressive, fast spread, highly metastatic sEV extracted from TNBC cells (MB-MDA-231) with respect to the ones from non-metastatic breast cancer cells (MCF7)





IV. Extracellular Vesicles (EVs)



Cell expansion from a GMP compliant master cell bank



Fill & finish as ready to use solutions





Small Angle X-Ray Scattering

> Ultraviolet Resonant Raman spectroscopy

spectroscopy



T51.8

1118 nm

NTA

Name	EV isolation method	NTA Concentration (particles/ml)	NTA Average Size (nm)
sEV1	TFF 750 kD	1-4 x 10 ¹¹	100.8
sEV2	TFF 750 kD + UC	1-4 x 10 ¹¹	114
sEV3	TFF 750 kD +UC + SEC	0.8-2x10 ¹⁰	116

 Table 1: Samples analysed with description of isolation method and concentration and average size measured my means of Nanoparticle Tracking Analysis.



IV. Extracellular Vesicles (EVs)

SAXS

scattering intensity versus the momentum transfer q is function of the defined size d, as $q=2\pi/d$.





s-EVs uptake and cell biomechanics

Are sEVs modulators of biomechanics properties?



Cell proliferation assay

Effect of sEVs from Triple Negative Breast Cancer cell lines (MDA-MB-231) on the biomechanics of non metastatic breast cancer cells (MCF7)



s-EVs uptake and cell biomechanics

Are sEVs modulators of biomechanics properties?



breast cell line (MCF10A)





 $E = A_L/A_S - I$

E<0.5 spherical cells E>0.5 elliptical cells













* = p < 0.05; ** = p < 0.01; *** = p < 0.001; **** = p < 0.001; **** = p < 0.0001





Pyramidal tips k = 0.200 N/m, tip curvature < 20 nm; two-slope modified Hertz-Sneddon model to fit the force curves

231_sEVs-treated MCF7 cells:

Y2 significantly lower than MCF7 cells Y1 comparable to MCF7 cells.





* = p < 0.05; ** = p < 0.01; *** = p < 0.001; **** = p < 0.0001; **** = p < 0.0001









PROTEIN CONTENT INCREASES IN TREATED CELLS MDA-MB-231 have more lipids than both MCF7 cells



Part I: conclusions

- s-EVs soften non-metastatic breast cells
- s-EVs induce modification of nuclear shape and size, actin fiber formation
- The Young modulus changes at the level of nucleous, not membrane-associated cortical fibers
- Chromatin decondensation might play a relevant role in biomechanical changes

Inducede biomechanical changes on model cell cultures by patientderived s-EVs as a new functional assay in metastatic cancer diagnosis and therapy.







Part II: s-EVs uptake mechanism



Model system:

Cell expansion

Fill & finish

as ready to use solutions

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EVs from Umbilical Cord multipotent Mesenchymal **StemCells**

OEVs . Protein

Small Angle X-Rav Scatterin

FTIR

Model Plasma Cell Membrane with Lipid Rafts

Purification

by filtration and centrifugation steps

Characterization

Ultraviolet Resonant Raman spectroscopy

Atomic Force



s-EVs uptake mechanism



Atomic Force Microscopy

- Morphological/mechanical characterization
- ✓ (Sub)Nanometer resolution
- Liquid environment

Neutron Reflectometry

AFM





The Reflectivity spectrum obtained is given by the interference of the waves reflected from the top and bottom of each layer



- ✓ Sample cross sectional profile extracted from data analysis
- ✓ Sensitive to light elements (H,O,N,C..)

 Sample cross-sectional profile: thickness, composition, compactness, roughness

Playing with selective deuteration, protiated molecules can be evidenced in a ghost phospholipid matrix



Model lipid bilayers

Multicomponent artificial lipid bilayers for mimicking cellular membranes





Nanoparticles + artificial membranes/EVs



Montis et al., Journal of Colloid and Interface Science (2020)





Model lipid bilayers

>DOPC + SM (66:33) + 5% Chol



68/22/10

DOPC



31.2







s-EVs interaction with membrane



Patches protruding 3-4 nm above SLB tend to expand in a more favourable fashion in L_d phase

sEVs preferentially dock and break at phases borders!

F. Perissinotto et al., Nanoscale, 2021, 13, 5224



s-EVs interaction with membrane

L_o phase re-shaping, borders granularity



From literature:

- -> cholesterol depletion
- -> components redistribution

Distinct processes occur:

I. Fast diffusion of lighter elementslaterally migrating along phaseboundaries

2. Diffusion of bulkier sEVs components mixing with target membrane



The uptake process is different from 'standard' membrane fusion



Playing with selective deuteration: protiated molecules in a ghost phospholipid matrix

- 20% volume penetration
- Change in contrast spans whole membrane thickness
- Asymmetry

	AFM ∆ Z (nm)	NR h (nm)
PC	5.1 ± 0.6	4.2 ± 0.3
PC+EVs	6 ± 2	5.4 ± 0.3

F. Perissinotto et al., Nanoscale, 2021, 13, 5224



s-EVs interaction with membrane





F. Perissinotto et al., Nanoscale, 2021, 13, 5224



s-EVs interaction with membrane

DOPC







Part II: conclusions

- s-EVs on artificial lipid bilayers break and form EVs-membrane domains
- Phase borders are docking sites
- Different s-EVs components spread with different kinetics
- The area of s-EVs-membrane domains increases over time: initial nucleation seeds act as docking sites for other s-EVs from solution
- The final membrane is asymmetric
- Cargo release in this case seems favoured





WHAT about cholesterol?





EV FROM TRBC*: ADSORPTION



LbL composition: DOPC, SM (2:1) with 17 mol% CHOL







... AND IF WE CHANGE ev CELL ORIGIN*?









³⁴ Paba, et al. Accepted at JCIS – 19.08-23

... WHAT if we remove chol?



35 Paba, et al. Accepted at JCIS – 19.08-23

*







FUTURE PLANS: CAVEOLIN-1



Campos, A., et al., Biomolecules, 2019.





PORE SPANNING MEMBRANE



LUO, Yitian, et al., Structure, 2021. Zhang, Y., et al., Molecules, 2021. Gunduz, M., et al., 2011.







Aknowledgments

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Thank you for your attention!

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https://www.elettra.trieste.it/userarea/ access-request.html BEAMTIME expenses covered for EU scientists

OWSD PhD Fellowship offered to women scientists from science- and technology-lagging countries (STLCs) to undertake PhD research in the natural, engineering and information technology sciences

U. Of the French West Indies might be interested to participate to the programme.













Mechanisms of EVs uptake





Koponen et al., Biosens. Bioelectronics, 2020, 168, 112510 Suutari et al., Small, 2016, 12, 6289-6300



Bonsergent P6.91a Semigaglies 14 Beatrice (Wednesday)

Lara et al.., J Nanobiotechnol 18, 2020, 20

Role of Extracellular Vesicles in modulation of biomechanical properties



Verdera et al., Journal of Controlled Release, 2017, 266, 100-108



Joshi et al., ACS Nano 2020, 14, 4444 4455



Multicomponent artificial lipid bilayers for mimicking cellular membranes



Proteoliposomes + artificial membranes



> Nanoparticles + artificial membranes/EVs



Montis et al., Journal of Colloid and Interface Science (2020)

How do EVs interact with cell membranes? An integrated biophysical approach







Good contrast for molecules with high electron densities (eg. SUGARS)

Neutron

Playing with selective deuteration, protiated molecules can be evidenced in a ghost phospholipid matrix

Membrane components distribution





- ✓ Non-destructive
- \checkmark Possibility of selective deuteration to play with contrast
- ✓ Deep material penetration (buried systems)



External interacting molecules distribution



AFM imaging of single EVs



50

100

150 Offset (nm)

EVs derived from Umbilical cord Stem Cells

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I.Prada, J. Meldolesi, Int. J. Mol. Sci. 2016