


Practical and technical applications of thermostable enzymes

- higher reaction rate at higher T
- lower risk of microbial contaminations

Enzyme	Temperature range (°C)	Bioconversions	Applications
α -Amylase (bacterial)	90-100	Starch \rightarrow dextrose syrups	Starch hydrolysis, brewing, baking, detergents
α -Amylase (fungal)	50-60	Starch \rightarrow dextrose syrups	Production of maltose
Pullulanase	50-60	Starch \rightarrow dextrose syrups	Production of glucose syrups
Xylanase	45-65, 105 ^a	Craft pulp \rightarrow xylan + lignin	Pulp and paper industry
Chitinase	65-75 ^b	Chitin \rightarrow chitobiose	Food, cosmetics, pharmaceuticals, agrochemicals
		Chitin \rightarrow N-acetyl glucosamine (chitinase)	
		N-acetyl glucosamine \rightarrow glucosamine (deacetylation)	
		Chitin \rightarrow chitosan (deacetylase)	
Cellulase	45-55, 95 ^c	Cellulose \rightarrow glucose	Cellulose hydrolysis, polymer degradation in detergents
Protease	65-85	Protein \rightarrow amino acids and peptides	Baking, brewing, detergents, leather industry
Lipase	30-70	Fat removal, hydrolysis, interesterification, alcoholysis, aminolysis	Dairy, oleo chemical, detergent, pulp, pharmaceuticals, cosmetics and leather industry
DNA polymerase	90-95	DNA amplification	Genetic engineering/PCR 

DNA polymerase from *Thermus aquaticus*: Taq polymerase

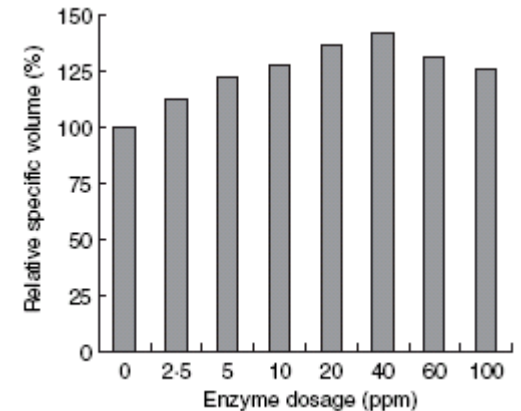
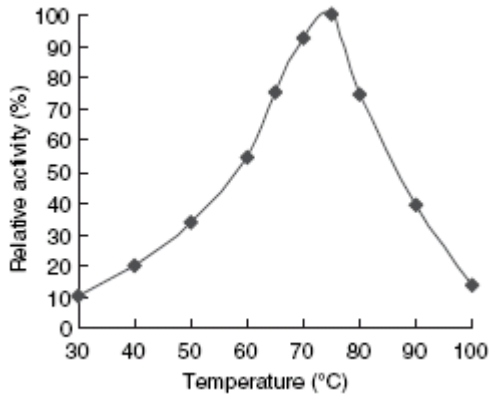
XYLANASE

Pulp and paper industry

Bakery

Higher quality of bread

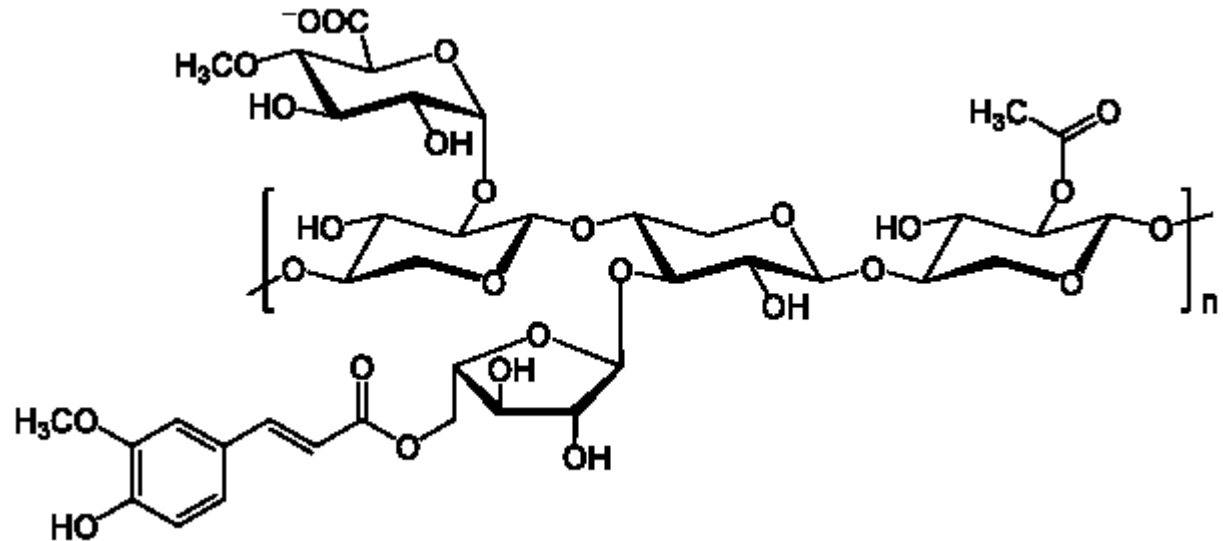
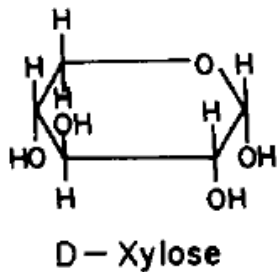
Enzyme active at 60-75°C, also during cooking



Xylan: a group of hemicelluloses present in plant cell walls

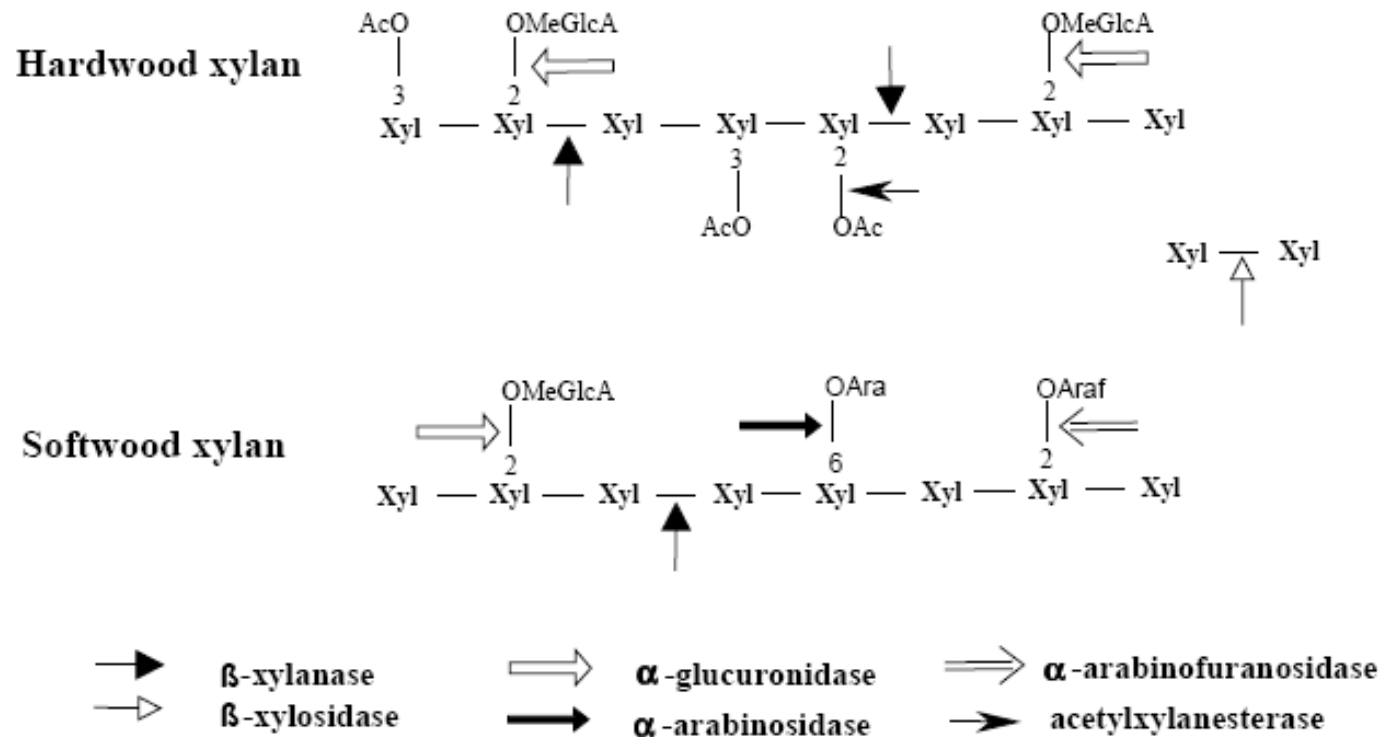
Ubiquitous as cellulose in plant cell walls and contain predominantly β -D-xylose units.

Typically the content of xylans in hardwoods are 10 - 35 % of the hemicelluloses and in softwood are 10 - 15 % of the hemicelluloses.



Example of xylane structure (ester of ferulate)

Enzymes degrading hemicellulose



Endo- β -1,4-xylanases catalyse the random hydrolysis of β -1,4-glycosidic bonds in xylans.

Proteases in detergents

Source microorganisms and properties of thermostable proteolytic enzymes

Organism	Enzyme properties	
	Optimal temperature (°C)	Optimal pH
<i>Bacillus brevis</i>	60	10.5
<i>Bacillus licheniformis</i>	70	9.0
<i>Bacillus stearothermophilus</i>	60	–
<i>Bacillus stearothermophilus</i>	85	–
<i>Bacillus</i> sp. JB-99	80	6–12
<i>Bacillus stearothermophilus</i> TP26	75	–
<i>Bacillus</i> sp. no. AH-101	80	12.0–13.0
<i>Bacillus thermoruber</i>	45	9
→ <i>Pyrococcus</i> sp. KOD1	100	7
<i>Staphylothermus marinus</i>	–	9
<i>Thermoacidophiles</i> (archeal and bacterial origin)	60–70	7.0–8.5
<i>Thermococcus aggregans</i>	90	7.0
<i>Thermococcus celer</i>	95	7.5
<i>Thermococcus litoralis</i>	85	8.5
<i>Thermotoga maritima</i>	95	9.5

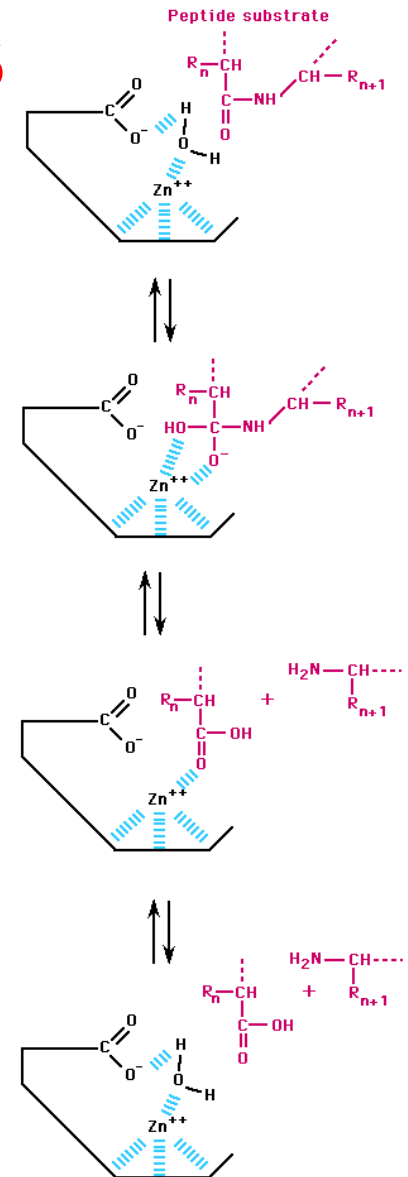
Proteases in detergents

Bacillus thermoproteolyticus neutral proteinase
Thermolysine

Stable at 50-60°C

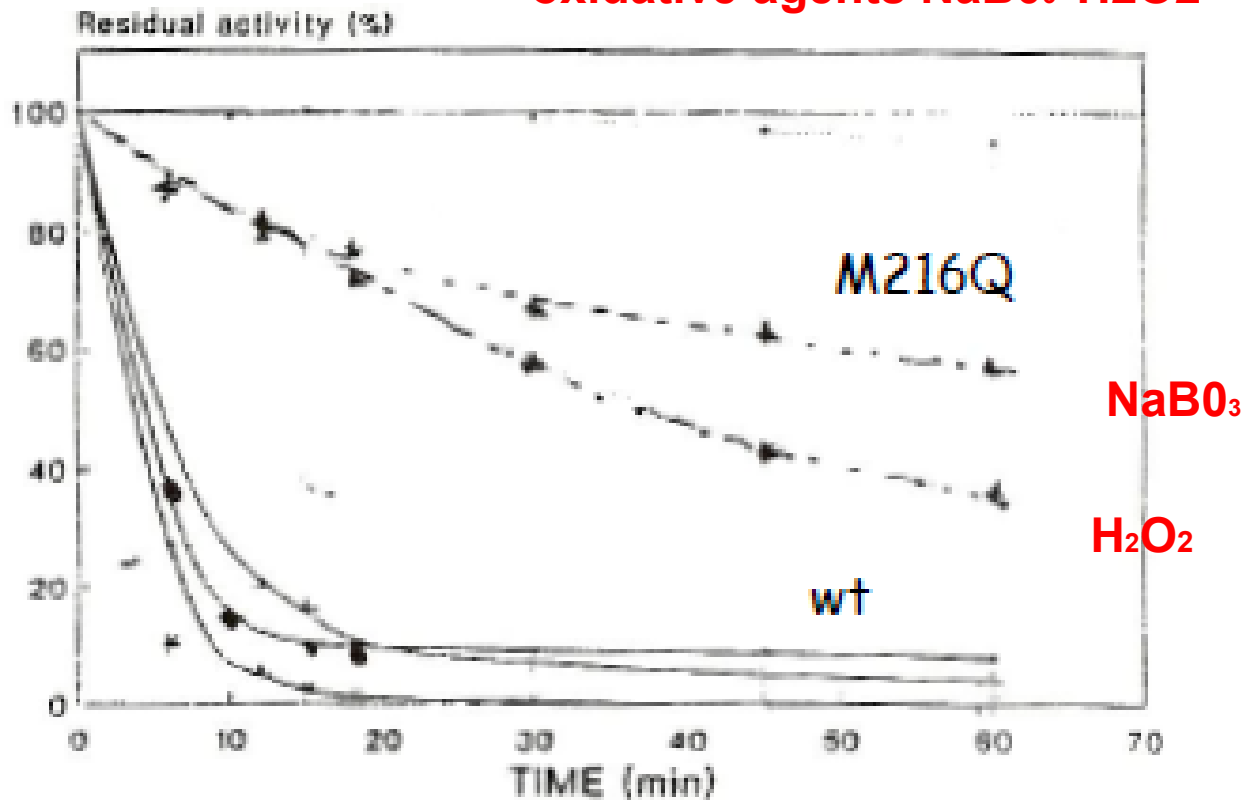


Zn protease



Stabilization of a protease towards oxidative agents (peracids and H₂O₂): Genetic engineering

Mutation of Metionine into Glutamine:
mutant displays higher stability towards
oxidative agents NaB₃O₆ H₂O₂



«natural» methods for
increasing enzyme stability

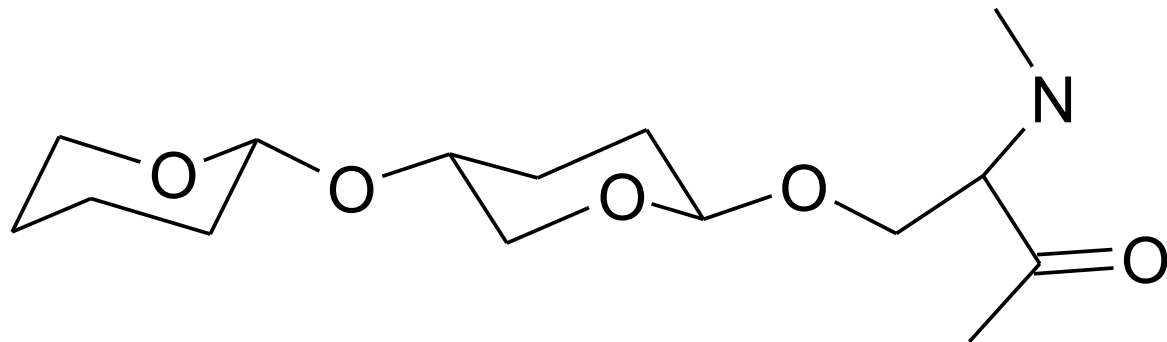
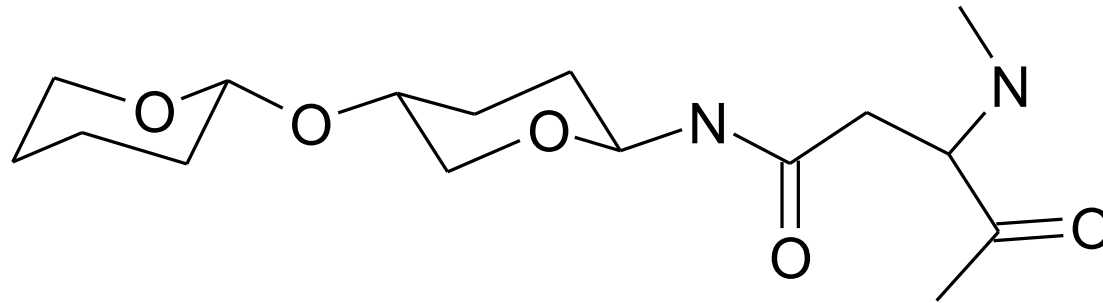
Glycosylation of proteins is a natural tool for increasing protein stability

Several roles have been suggested for the carbohydrate moieties of glycoproteins among which

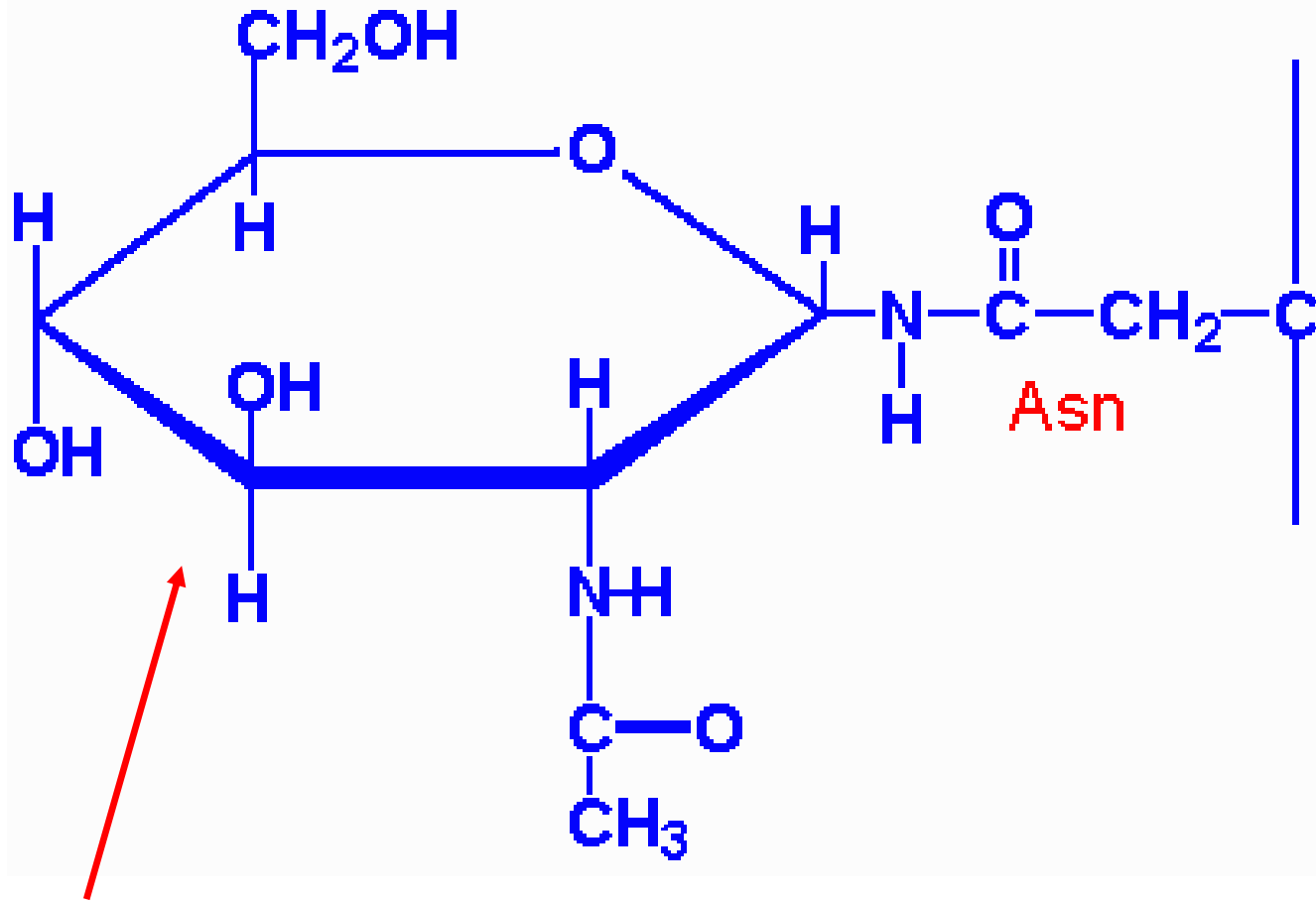
- **stabilization of protein conformation,**
- **protection from proteolysis**
- **Prevention of aggregation**

There are two main type of protein glycosylation:

- **N-glycosylation**, in which the glycan (10-35 mannose residues) is attached to an **Asn** residue present in the tripeptide consensus sequon **Asn-XSer/Thr** (where X can be any amino acid except Pro),
- **O-glycosylation**, in which the glycan is attached to a **Ser** or **Thr** residue.

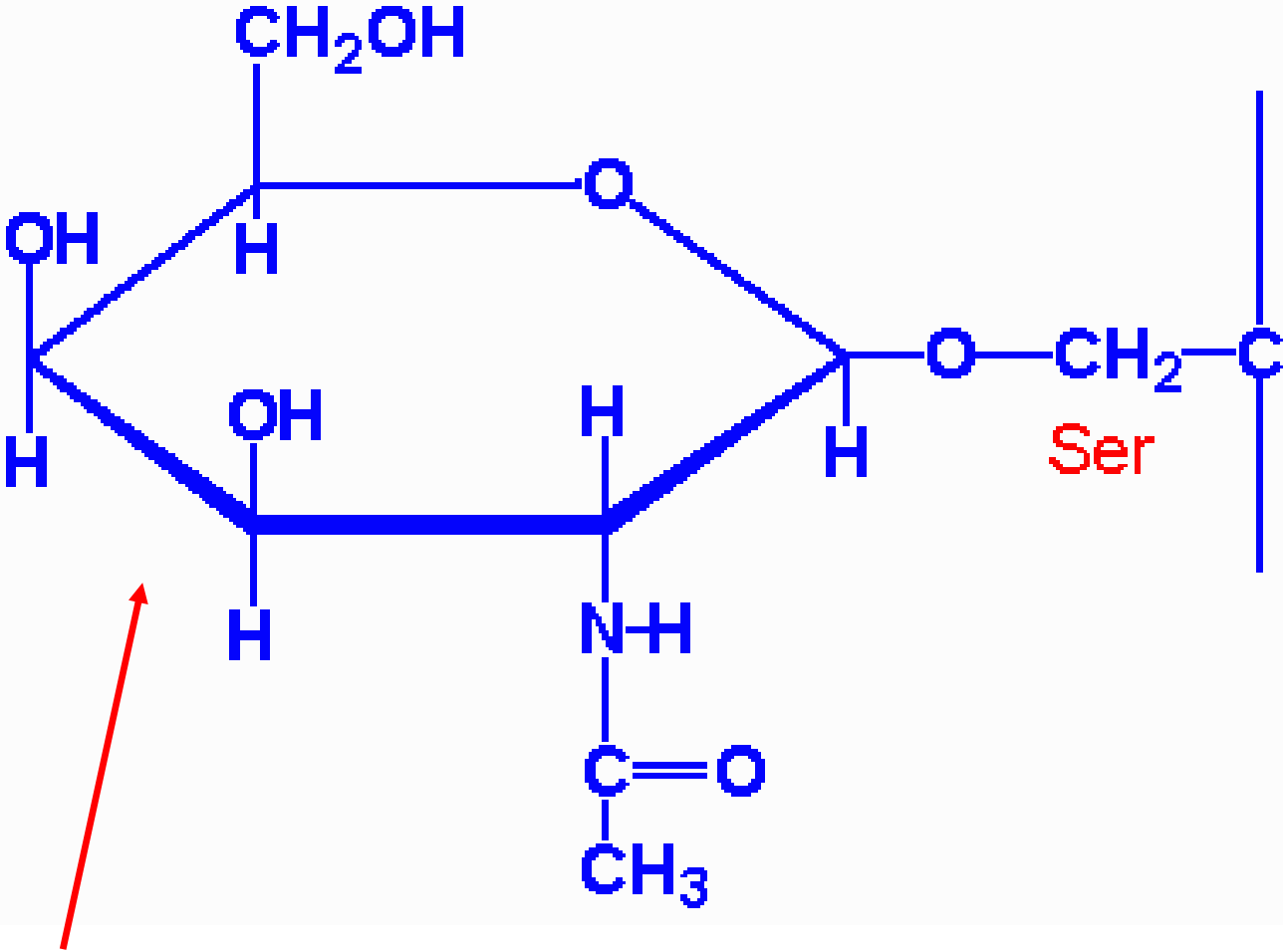


N-glycosidic bond



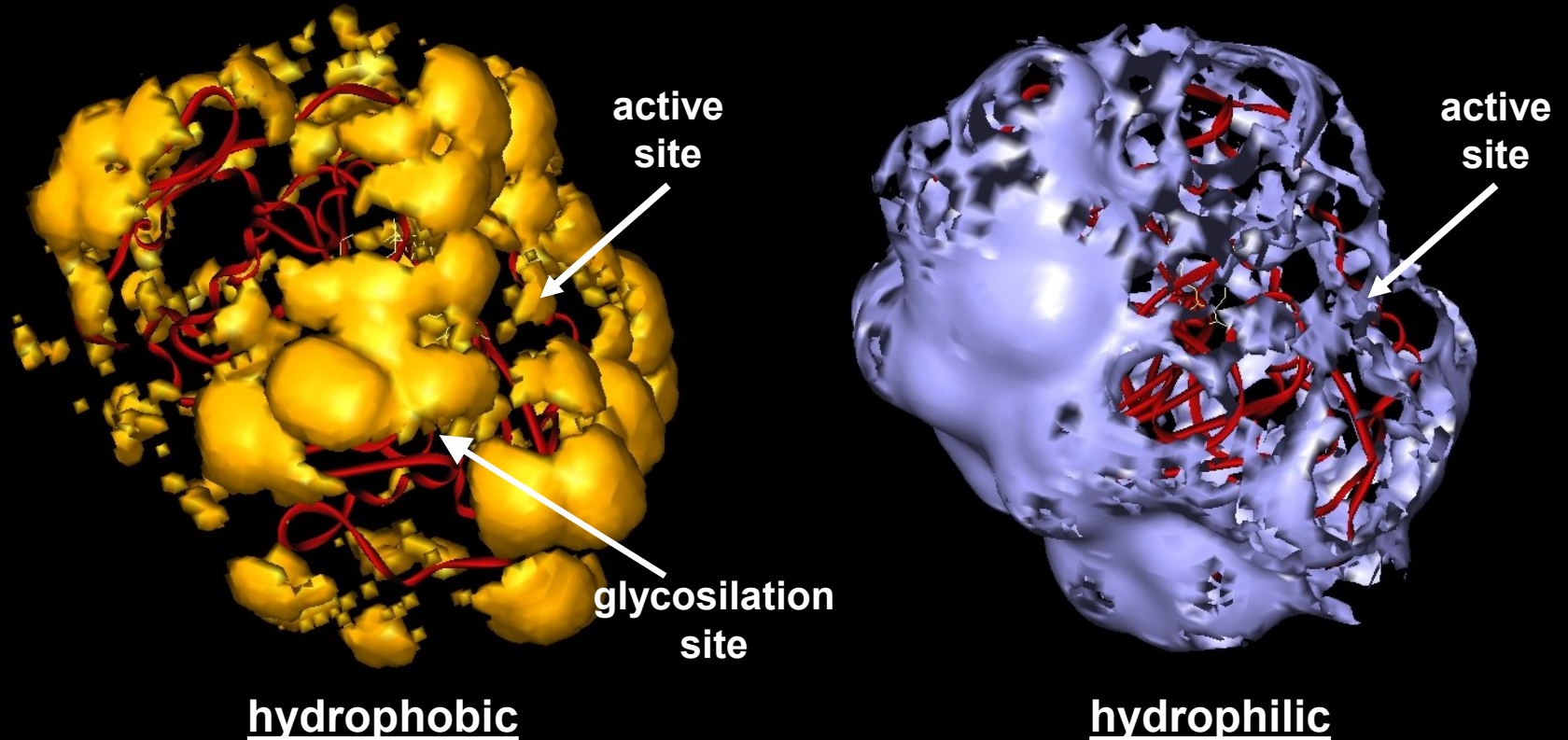
2-N-Acetylglucosamine

O-glycosidic bond



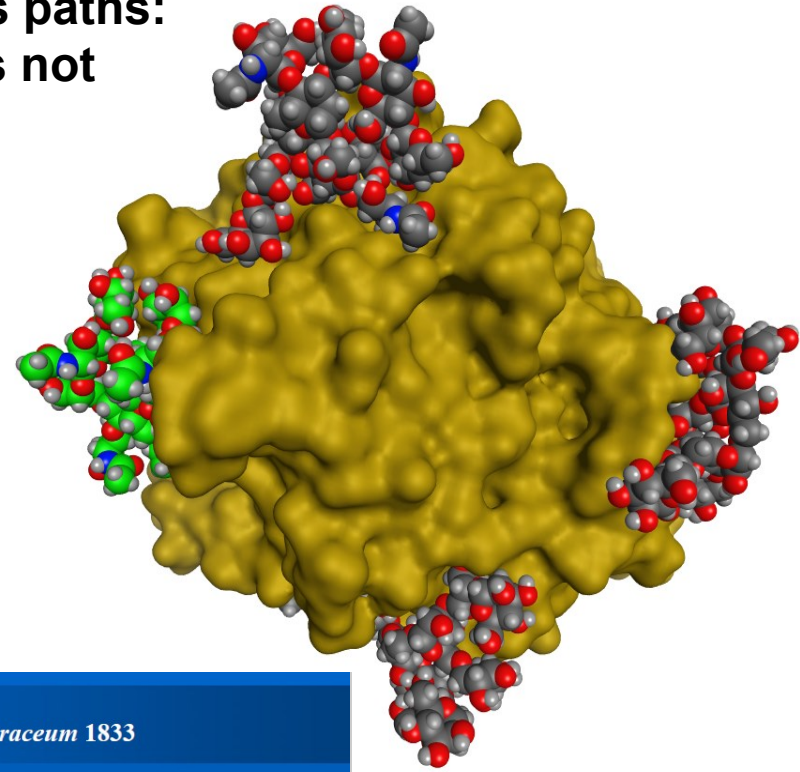
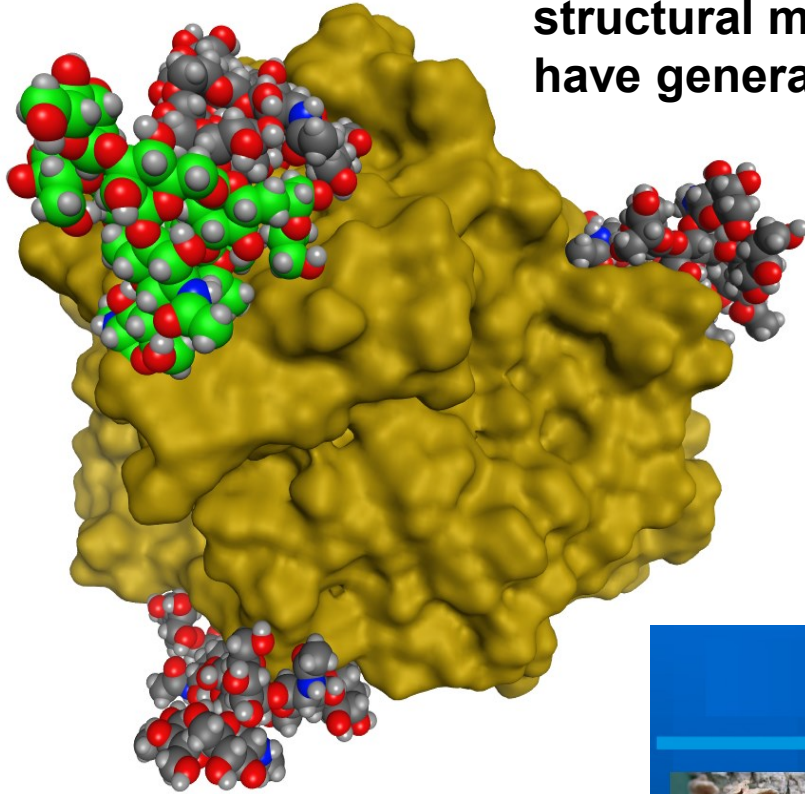
2-N-Acetylglucosamine

Lipasi CAL-B





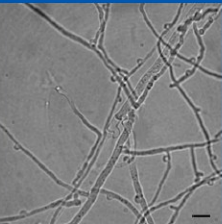
Glycans are generally inserted in the proximity of hydrophobic regions on the surface for stabilizing against hydration

Glycosylation occurs with broadly heterogeneous paths: structural models does not have general validity



Laccase from fungi:
degrades lignin

Steccherinum ochraceum 1833

		
<i>S. ochraceum</i> , natural fruit-bodies	<i>S. ochraceum</i> 1833, culture on MEA	<i>S. ochraceum</i> 1833, hyphae with clamps

Myasoedova, N.M., Chernykh, A.M., Psurtseva, N.V., Belova, N.V. and Golovleva, L.A. (2008)
New efficient producers of fungal laccases. *Appl Biochem Microbiol (Russia)* 44, 84-89.

Glycosylation improves thermal stability

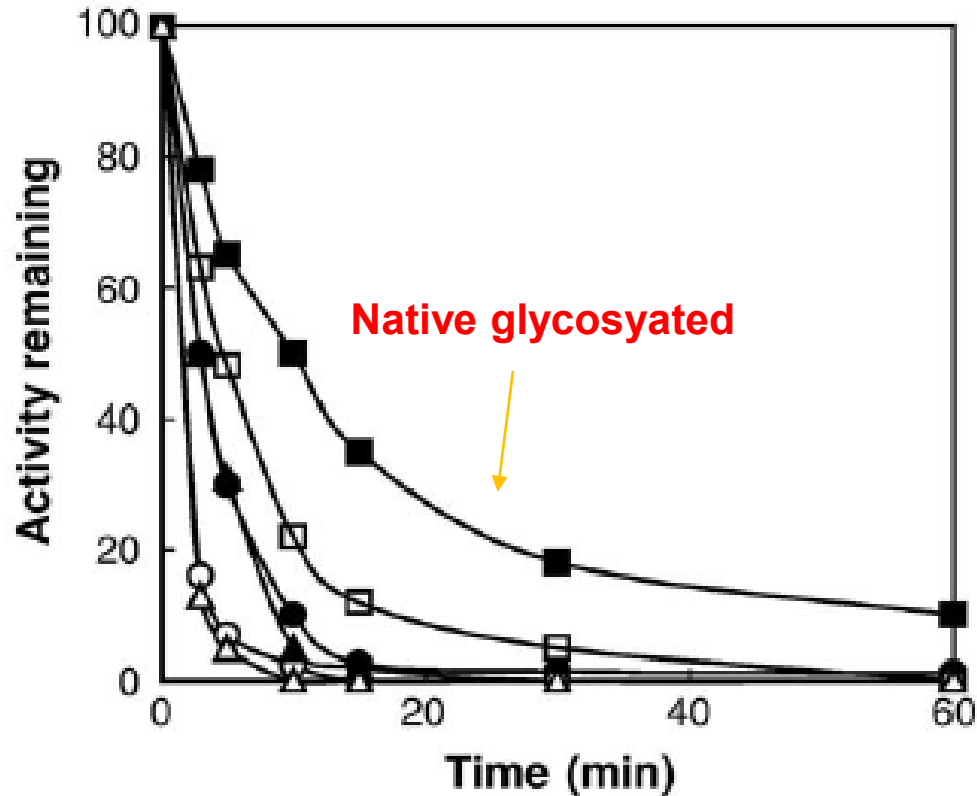


Fig. 4. Irreversible thermoinactivation of native and deglycosylated forms of glucoamylase at 70°C: Native (●), deglycosylated (○) at pH 3; native (■), deglycosylated (□) at pH 4.8; native (▲), deglycosylated (△) at pH 8. Further details are described under Materials and methods.

Glycosylation prevents aggregation

- improves the solubility of unfolded or partially folded proteins

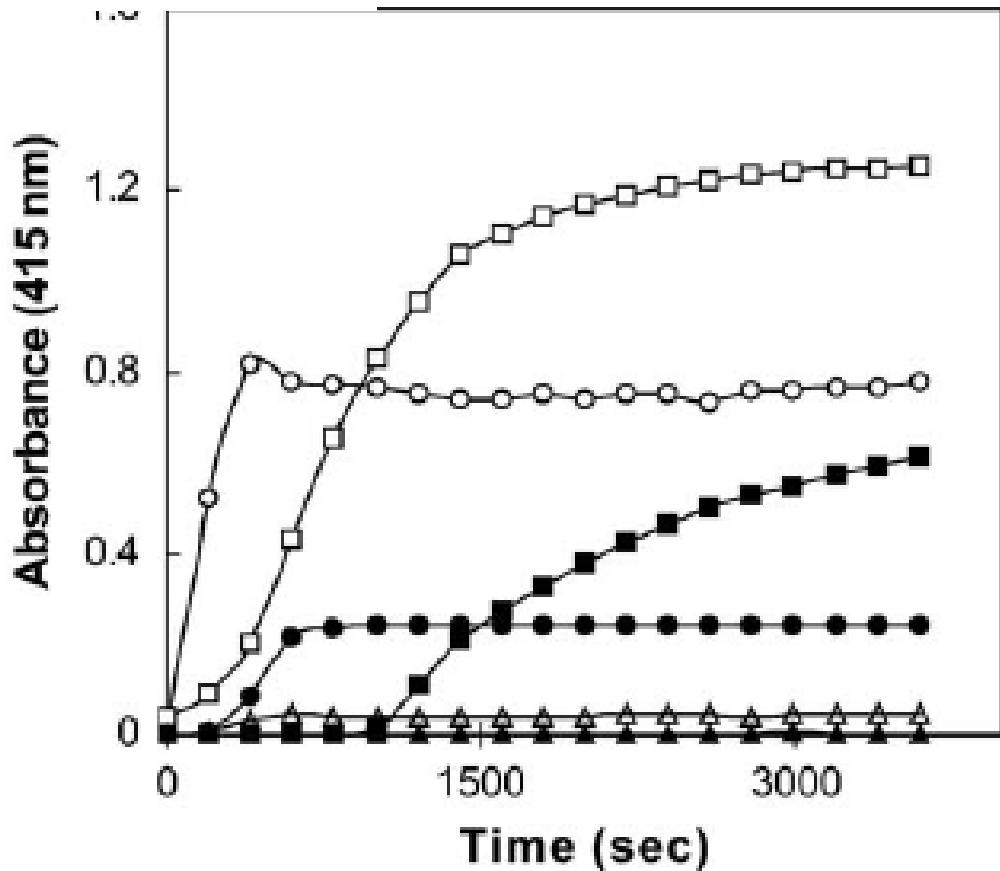
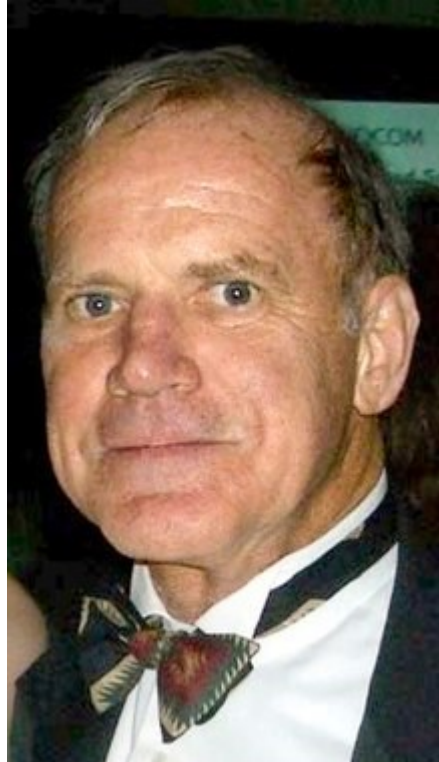


Fig. 5. Aggregation of native and deglycosylated forms of glucoamylase at 70°C: Native (●), deglycosylated (○) at pH 3; native (■), deglycosylated (□) at pH 4.8; native (▲), deglycosylated (△) at pH 8. For further details, please see Materials and methods.

Thermostable DNA polymerase in molecular biology



**Kary Banks Mullis (Lenoir, 28 dicembre 1944)
biochimico statunitense,
vincitore del Premio Nobel per la Chimica nel 1993 assieme a Michael Smith.**

Ha ottenuto il dottorato di ricerca all'[Università della California a Berkeley](#) nel [1973](#).

Mullis ha ricevuto il premio Nobel per lo sviluppo della tecnica della [reazione a catena della polimerasi](#) (*Polymerase Chain Reaction* o PCR), un processo già descritto da Kjell Kleppe e da [Har Gobind Khorana](#), Nobel nel 1968. La tecnica consente l'amplificazione [in vitro](#) di frammenti di [DNA](#). I miglioramenti apportati da Mullis hanno reso la PCR una tecnica fondamentale in [biochimica](#) e nella [biologia molecolare](#), con innumerevoli applicazioni in campo medico, agricolo, e investigativo.

Mullis è stato insignito nel 1993 anche del [Japan Prize](#).

[Surfista](#) e [contestatore](#) nella [Berkeley](#) degli [anni sessanta](#), Mullis è considerato un personaggio alquanto originale e discusso, che spesso si è scontrato con le posizioni "ortodosse" della [comunità scientifica](#). È noto per essere fortemente critico nei confronti della teoria diffusamente accettata del legame [HIV - AIDS](#), per lo scetticismo rispetto alle cause del [riscaldamento globale](#) e del [buco nell'ozono](#). In una famosa intervista si domandò provocatoriamente se avrebbe mai scoperto la PCR se non avesse assunto [LSD](#), concludendo che ne dubitava seriamente, in quanto poteva letteralmente veder lavorare i singoli polimeri e ammettendo di avere imparato parecchio grazie a tale esperienza.

Nel suo libro del [1998](#) ("*Ballando nudi nel campo della mente. Le idee (e le avventure) del più eccentrico tra gli scienziati moderni*") Mullis parla della sua visione del mondo e narra di episodi curiosi ed esperienze alquanto insolite da lui esperite, parla dell'[astrologia](#), della sua partecipazione al processo di [O. J. Simpson](#), del suo uso di droghe, per finire addirittura con l'ipotesi di essere stato rapito dagli [alieni](#); gli accadde una notte del 1985, in un bosco nei pressi di [Mendocino County](#) in [California](#).

Mullis racconta come ai tempi dei suoi studi a Berkeley mandò all'autorevole rivista inglese [Nature](#) un articolo fortemente speculativo nel quale sosteneva che metà della materia dell'universo andrebbe all'indietro nel tempo: l'articolo fu pubblicato. Un ventennio dopo propose alla stessa rivista un articolo in cui documentava la tecnica della PCR, che gli avrebbe poi valso il Nobel, e non fu pubblicato.

Recombinant DNA

- DNA molecules formed by laboratory methods of genetic recombination to bring together genetic material from multiple sources
- Proteins resulting from the expression of recombinant DNA within living cells are termed *recombinant proteins*
- Expression of foreign proteins requires the use of specialized *expression vectors*

**The first prerequisite is to obtain a sufficient DNA quantity
(amplification)**



DNA amplification is obtained by *molecular cloning* or by PCR (Polymer Chain Reaction)

PCR - Polymerase Chain Reaction

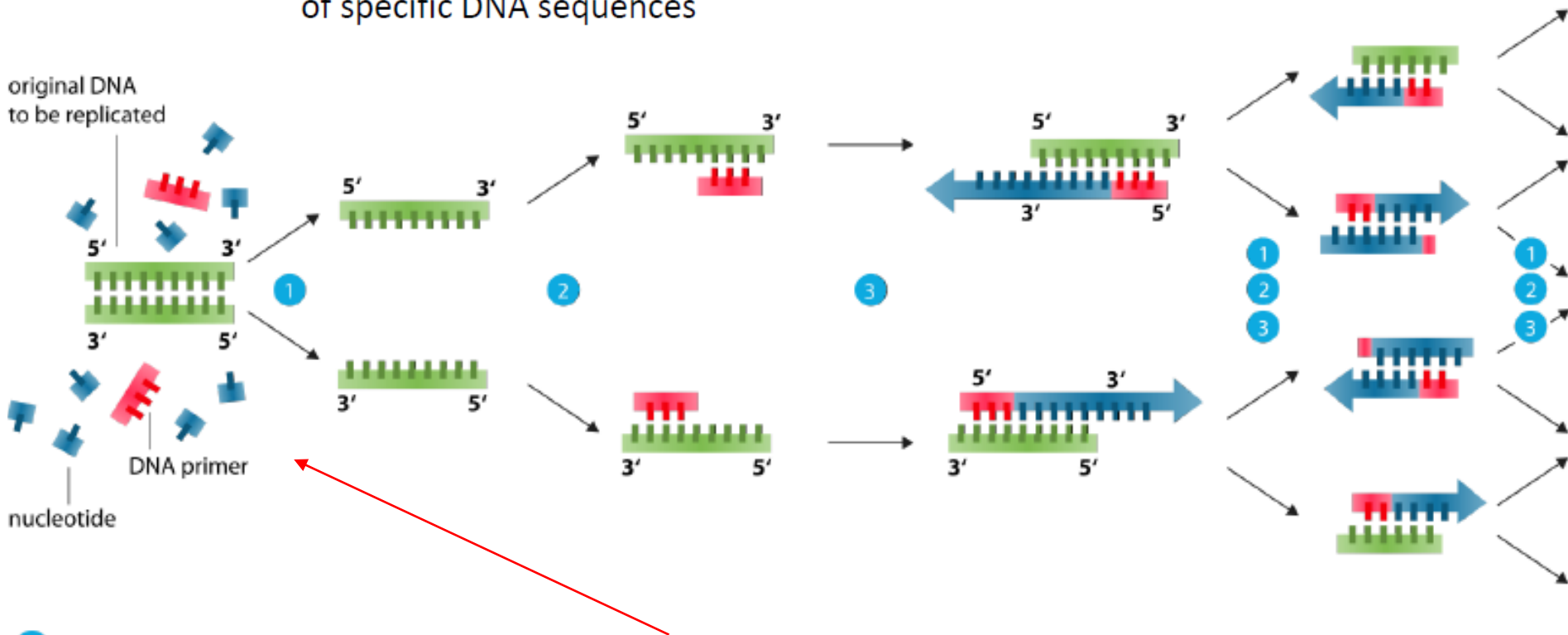
- PCR is a technology used mainly for amplification and/or isolation of specific DNA sequences
- The method relies on thermal cycling, consisting of cycles of repeated heating and cooling of the reaction for DNA melting and enzymatic replication of the DNA
- PCR employ a thermostable DNA polymerase, such as Taq polymerase (DNA polymerase from *Thermus aquaticus*)



Taq polymerase enzymatically assembles a new DNA by using single-stranded DNA as a template and DNA oligonucleotides (DNA primers), which are required for initiation of DNA synthesis

PCR - Polymerase Chain Reaction

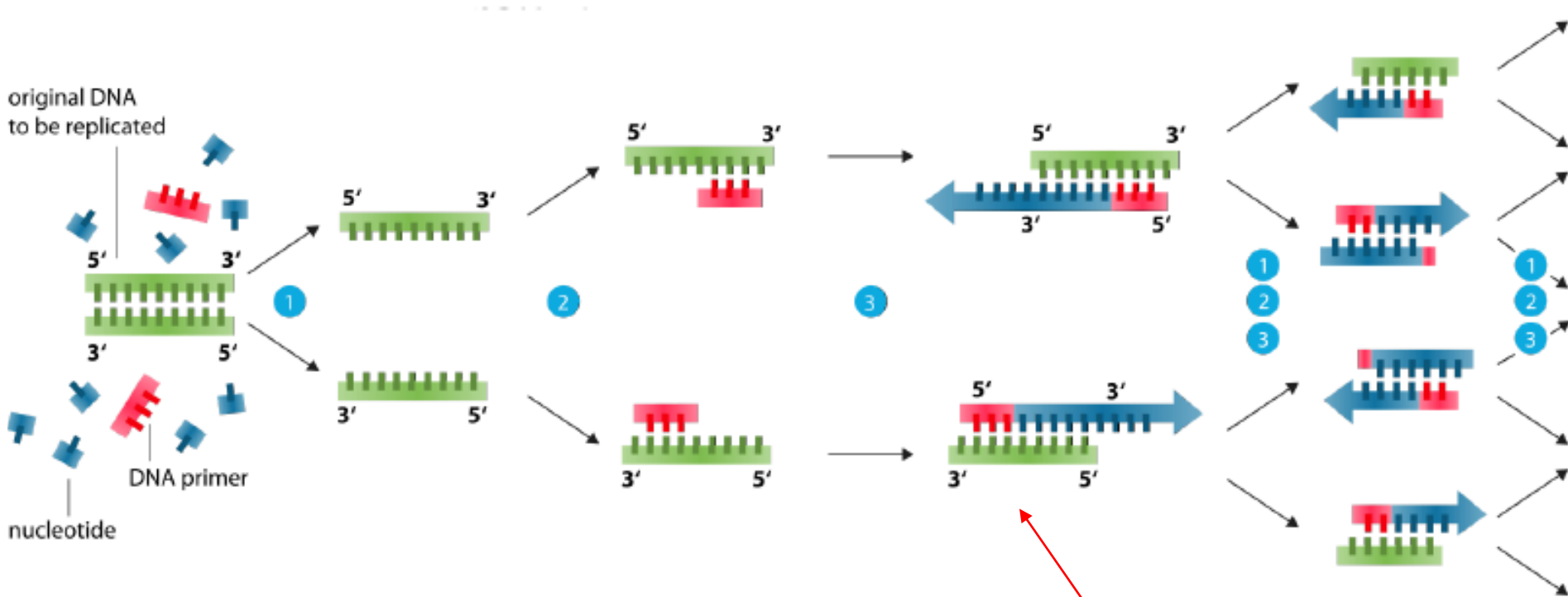
- PCR is a technology used mainly for amplification and/or isolation of specific DNA sequences



- 1 **Denaturation** at 94-96°C
- 2 **Annealing** at ~68°C
- 3 **Elongation** at ca. 72 °C

- The method relies on thermal cycling, consisting of cycles of repeated heating and cooling of the reaction for DNA melting and enzymatic replication of the DNA
- PCR employ a thermostable DNA polymerase, such as Taq polymerase (DNA polymerase from *Thermus aquaticus*)

PCR - Polymerase Chain Reaction



- 1 Denaturation at 94-96°C
- 2 Annealing at ~68°C
- 3 Elongation at ca. 72 °C

Taq polymerase enzymatically assembles a new DNA by using single-stranded DNA as a template and DNA oligonucleotides (DNA primers), which are required for initiation of DNA synthesis

Molecular cloning

1. Choice of host organism and cloning vector
2. Preparation of cloning DNA vector
3. Introduction of cloning vector into host organism
4. Screening for clones with desired DNA inserts

The great majority of molecular cloning experiments begin with a laboratory strain of the bacterium *Escherichia coli* and a *plasmid* cloning vector

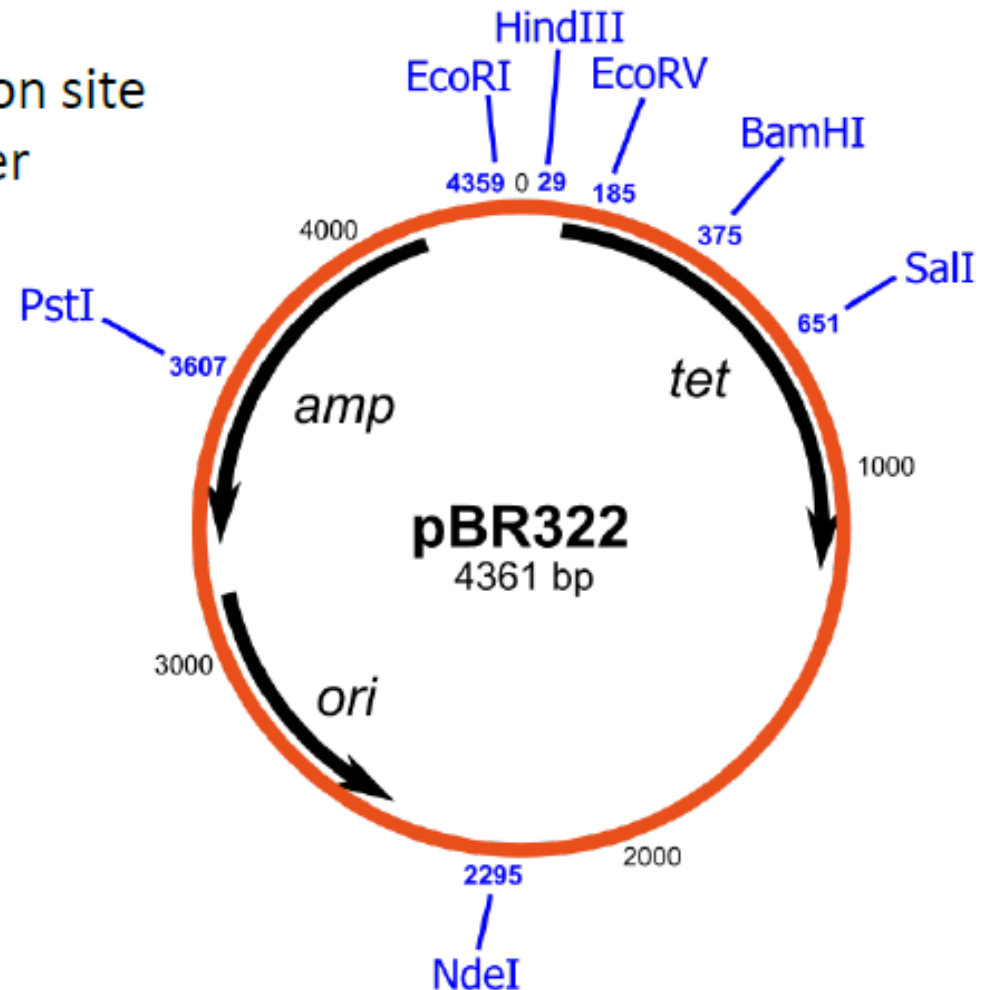


A plasmid is a small DNA molecule physically separated from a chromosomal DNA that can replicate independently

Plasmid vector

Essential plasmid elements:

- Origin of replication
- At least one cloning/restriction site
- At least one selectable marker

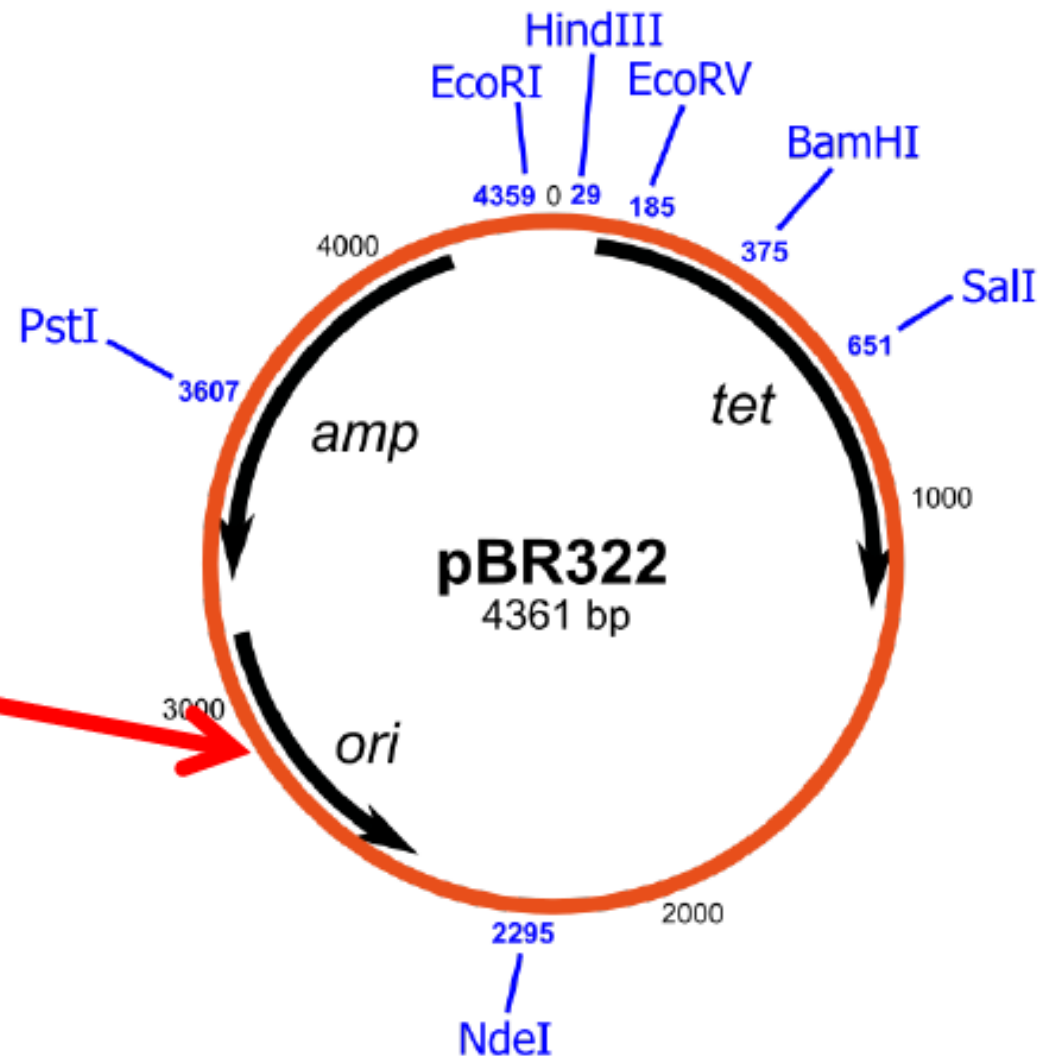


Plasmid vector

Essential plasmid elements:

- Origin of replication

The origin of replication (*ori*) is a particular sequence in a genome at which replication is initiated

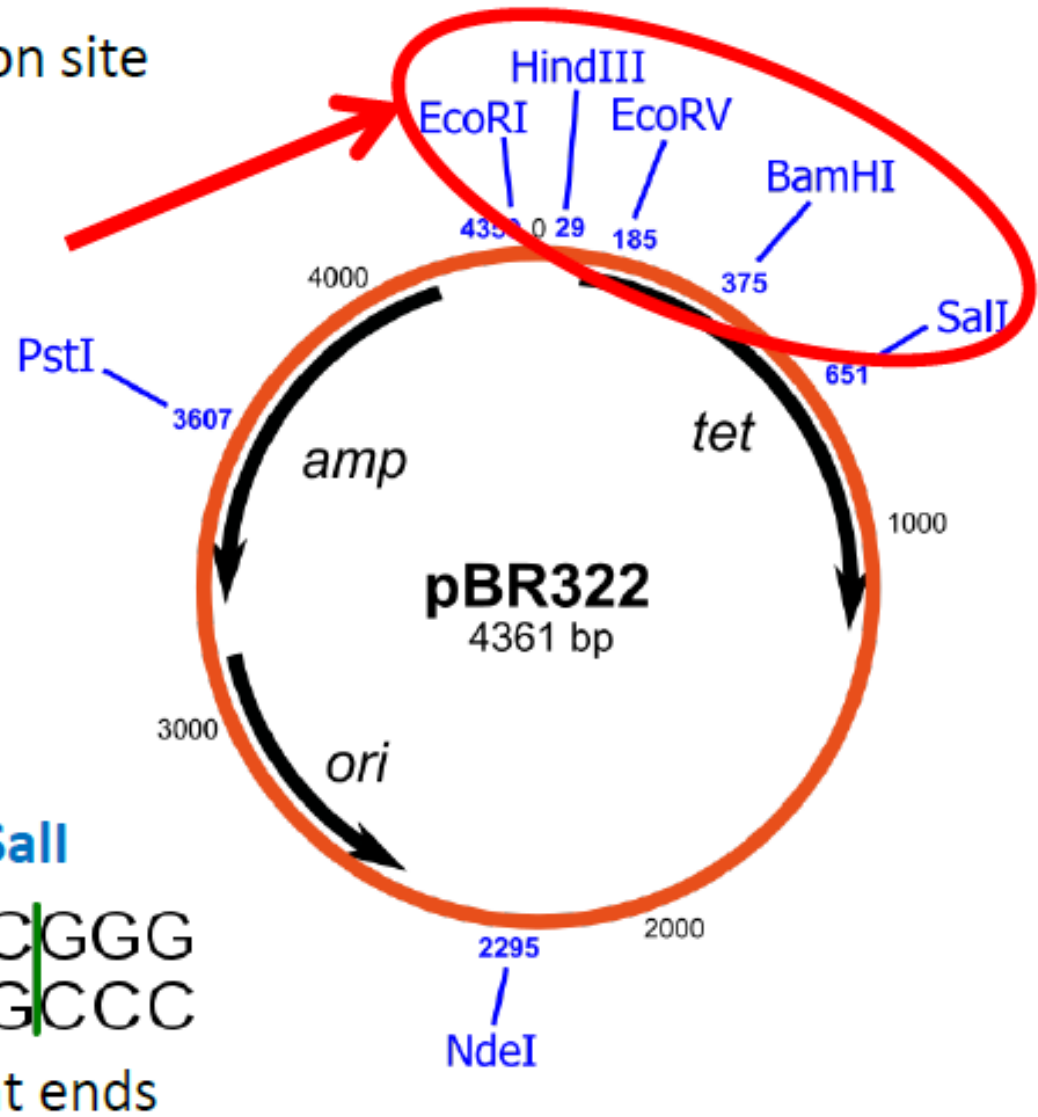


Plasmid vector

Essential plasmid elements:

- At least one cloning/restriction site

A restriction site is a nucleotide sequence specifically recognized by endonuclease enzymes (or restriction enzymes) that cuts DNA at or near the restriction sites



EcoRI



Sticky ends

SalI



Blunt ends

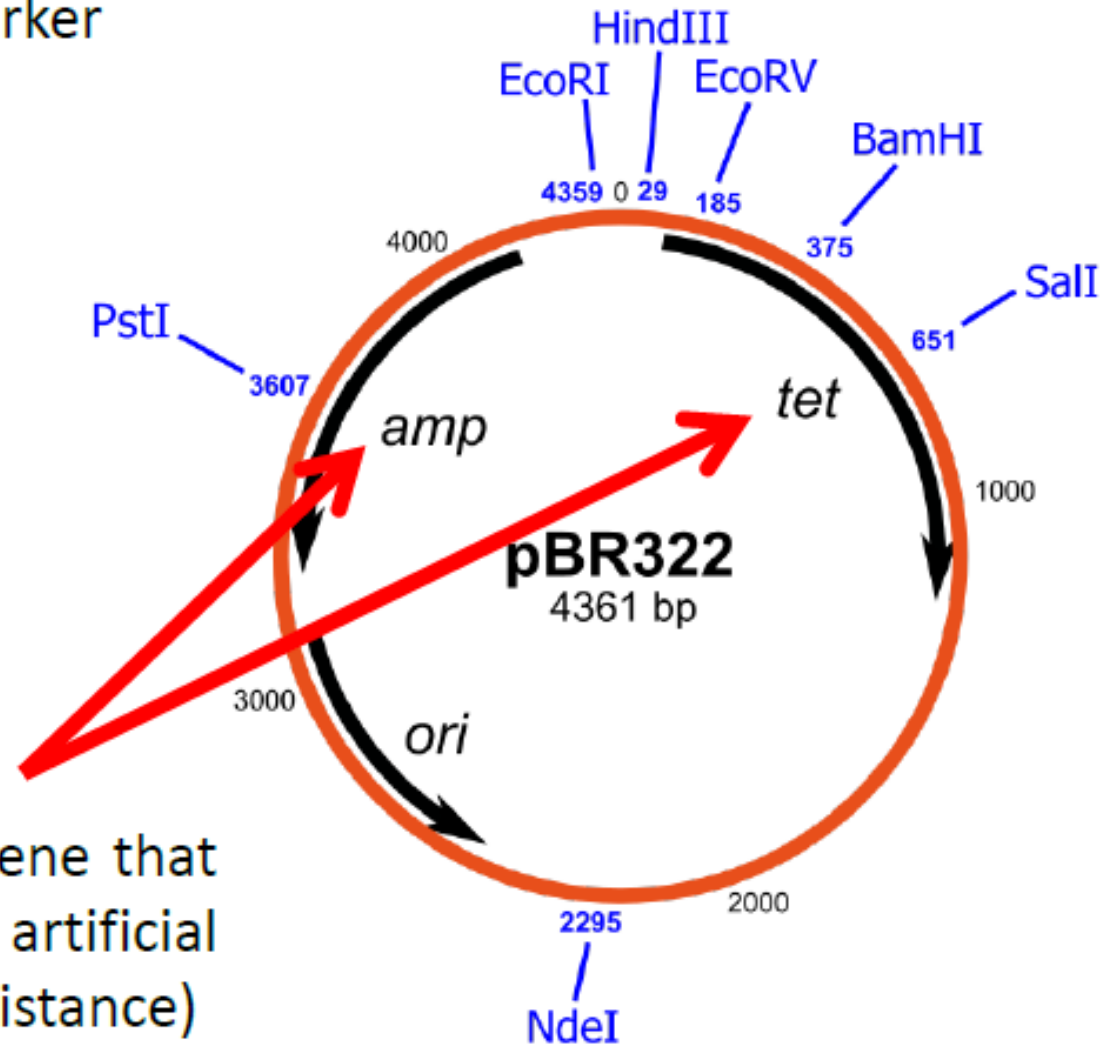
Plasmid vector

Essential plasmid elements:

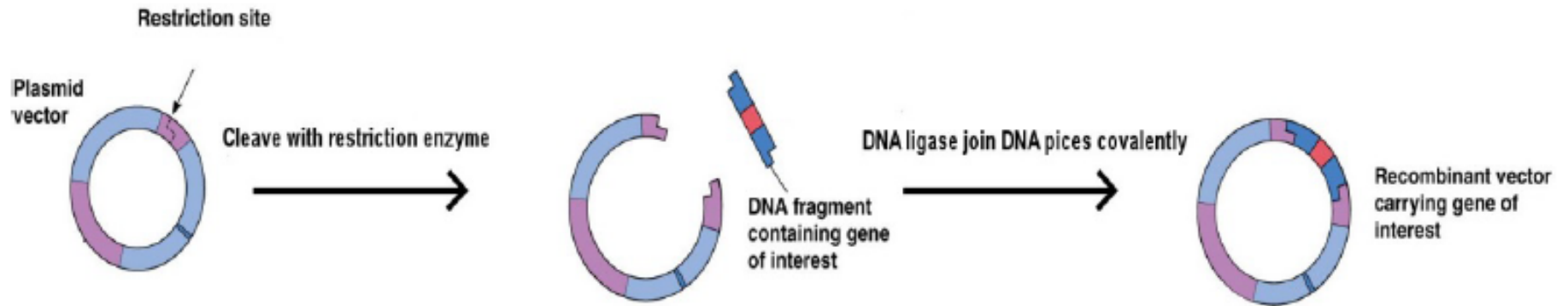
- At least one selectable marker

tet = tetracycline resistance
amp = ampicillin resistance

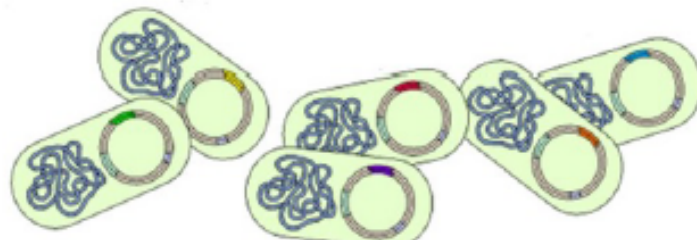
A selectable marker is a gene that confers a trait suitable for artificial selection (i.e. antibiotic resistance)



DNA insertion



Plasmid is inserted into the cloning host (i.e. *E. coli*)



Cell transformation

Insert exogenous DNA in host microorganism

Main techniques:

- Heat shock
- Divalent cations (CaCl_2)
- Electroporation (electric field of 10-20 kV/cm)
- Enzyme treatment



Increase cell permeation

