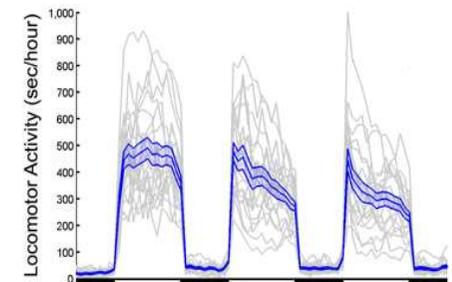
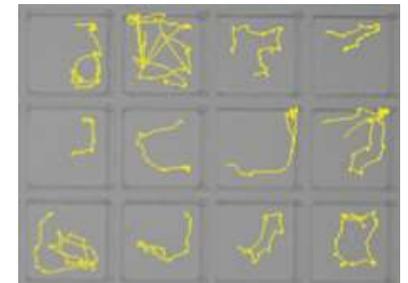


# MOLECULAR NEUROPHYSIOLOGY *-lesson 6-*



**Prof. G.Cellot**





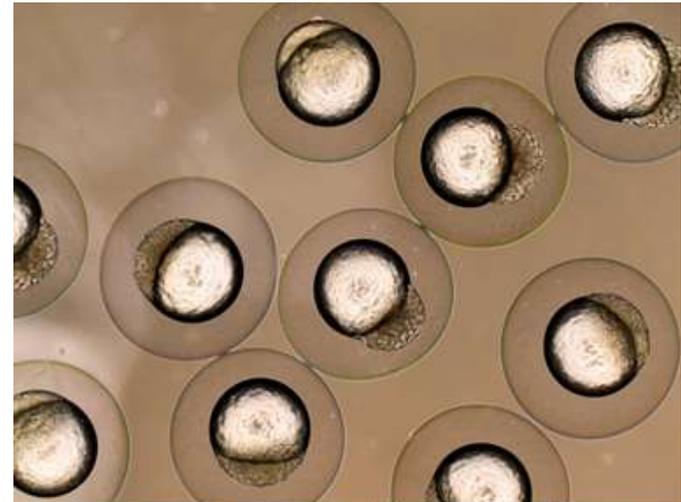
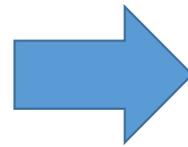
Zebrafish as alternative model for the study of synaptic plasticity and its behavioral correlates

# Is Zebrafish (*Danio Rerio*) a good alternative research model for biomedical applications?

- 70% of genetic homology with humans
- Ethics in using animal for research purpose

Principle of the 3Rs (Replacement, Reduction and Refinement) **OK**

- Cost effective



# Zebrafish (*Danio Rerio*) as alternative research model for biomedical applications

- Extra-uterine fertilization

- Quick development

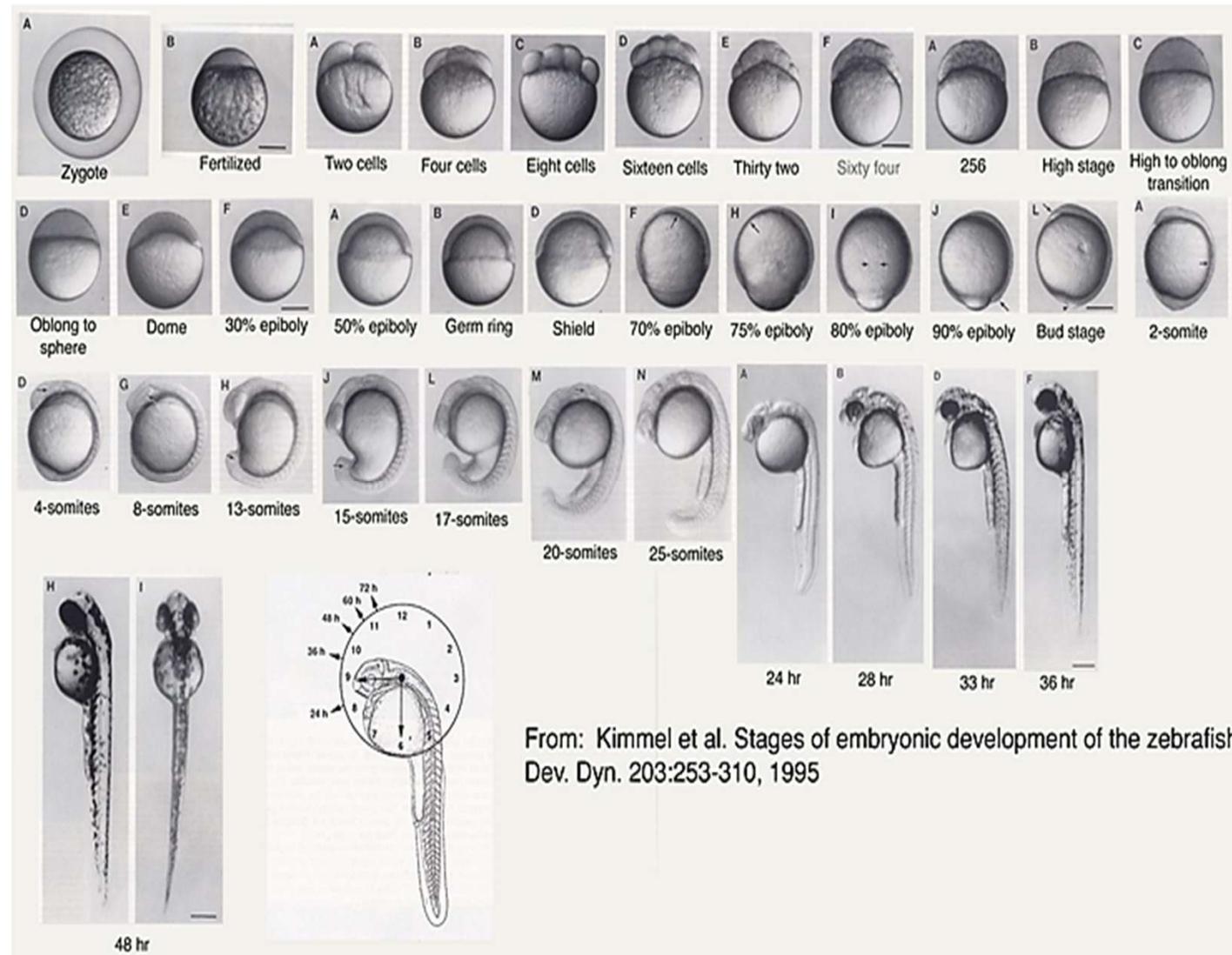
➔ *Genetic manipulation for transgenic lines*

- 2-3 years of lifespan

➔ *Studying aging, neurodegeneration,...*

- Optical transparency

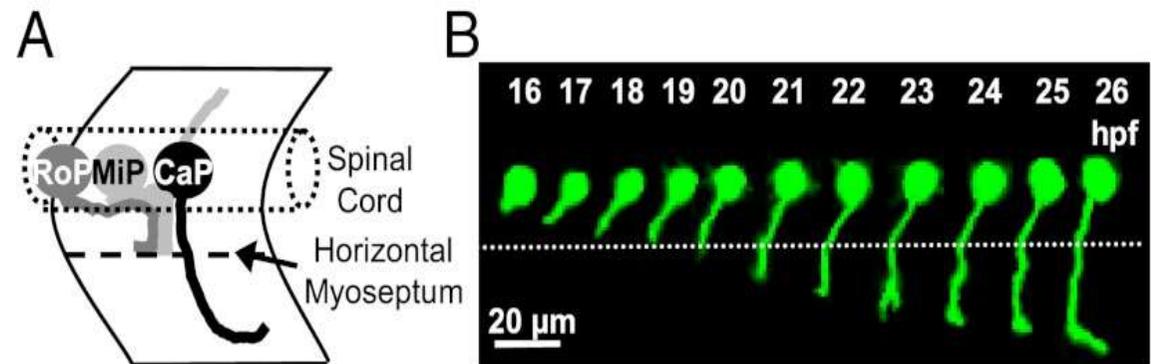
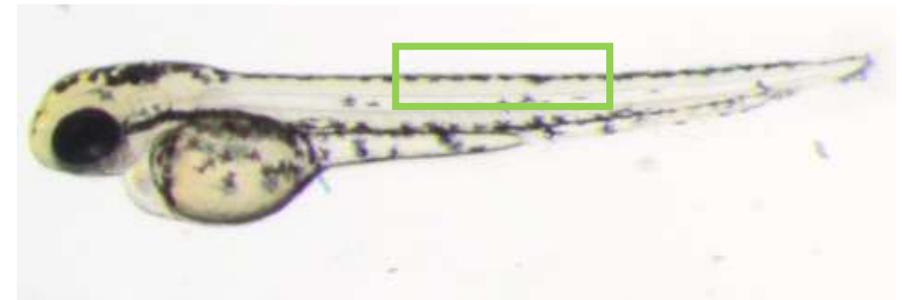
➔ *Accessibility of the nervous system for functional techniques*



From: Kimmel et al. Stages of embryonic development of the zebrafish *Dev. Dyn.* 203:253-310, 1995

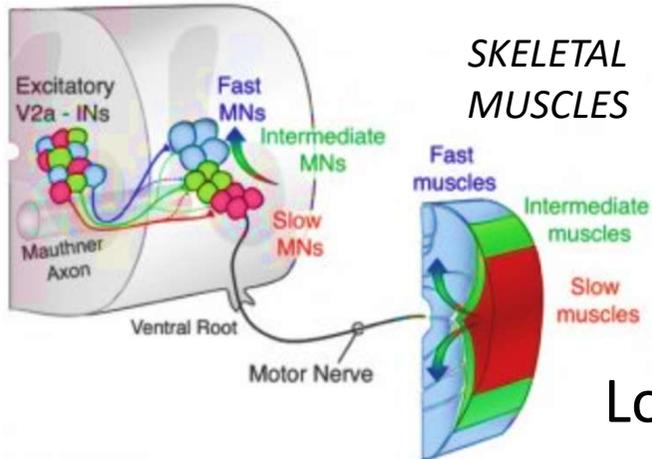
## Using zebrafish in neurophysiology to correlate behavior with neuronal function

- Spontaneous behavior
- Independent on sensory stimulation
- Correlating with neuro-muscular synapses



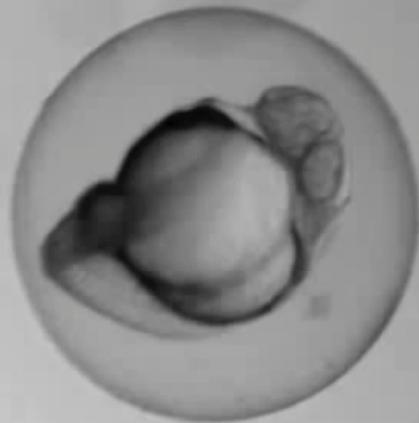
Plazas et al, PNAS 2013, DOI: [10.1073/pnas.1213048110](https://doi.org/10.1073/pnas.1213048110)

### SPINAL CORD



Locomotor activity

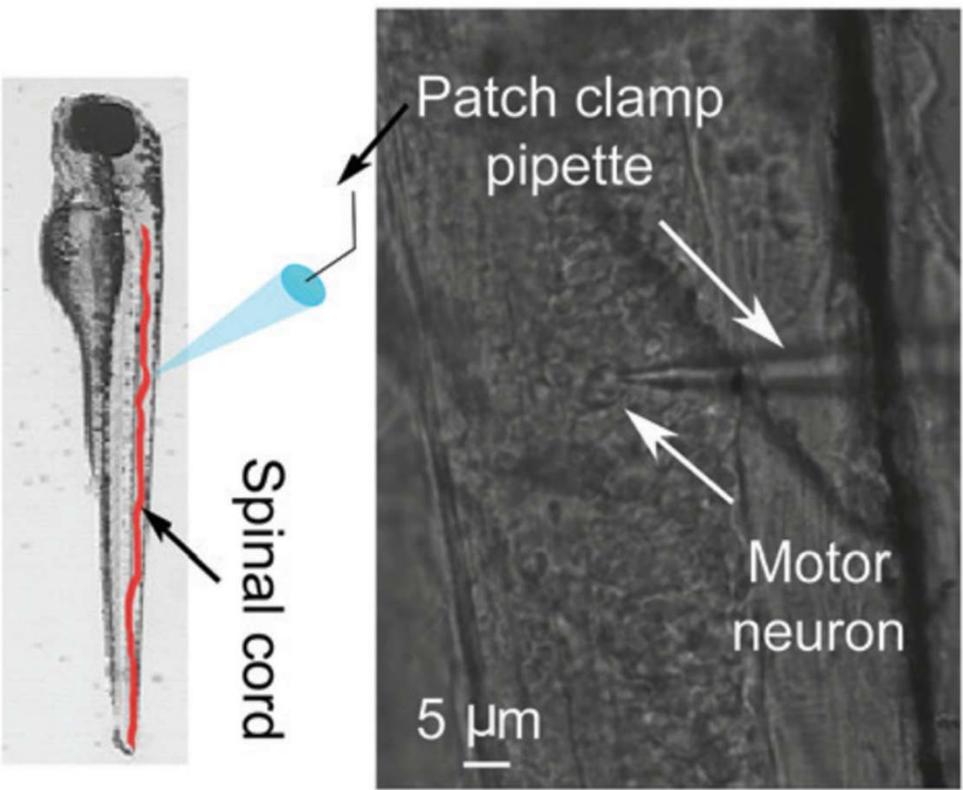
Modified from Ampatzis et al, Neuron, 2014



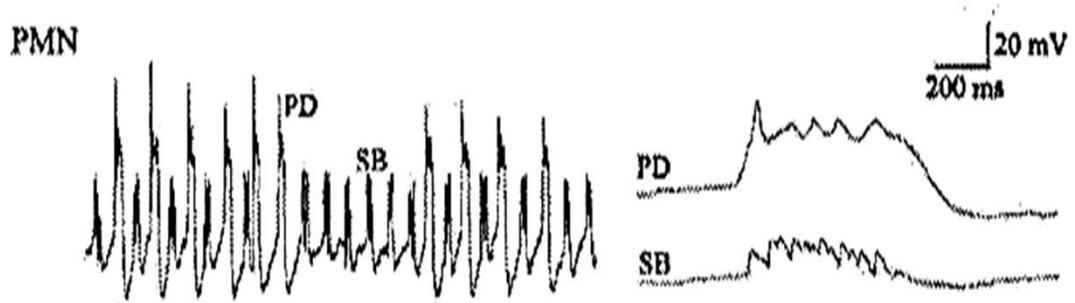
**24 hours:**  
**Coiling behavior**

# Using zebrafish in neurophysiology to correlate behavior with neuronal function

## *Electrical correlates of coiling behavior*



- Based on GJ dependent depolarizations
- Glycine depolarizing action

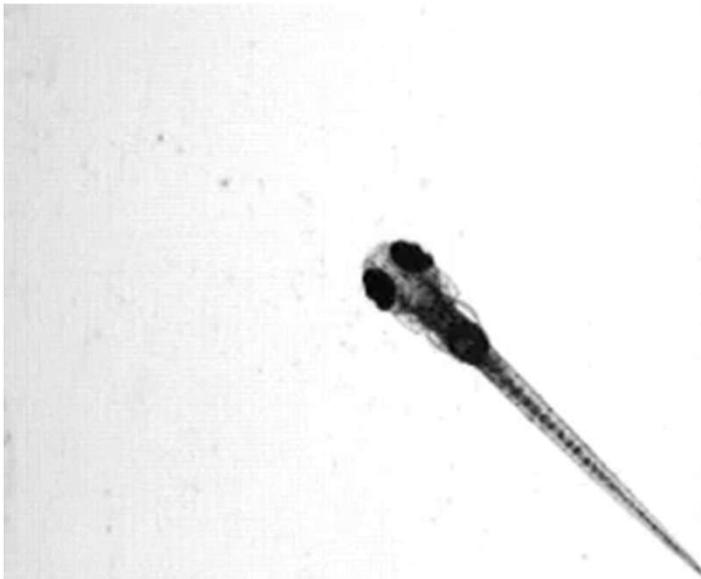


Cellot et al, Nanoscale Horizons, 2020

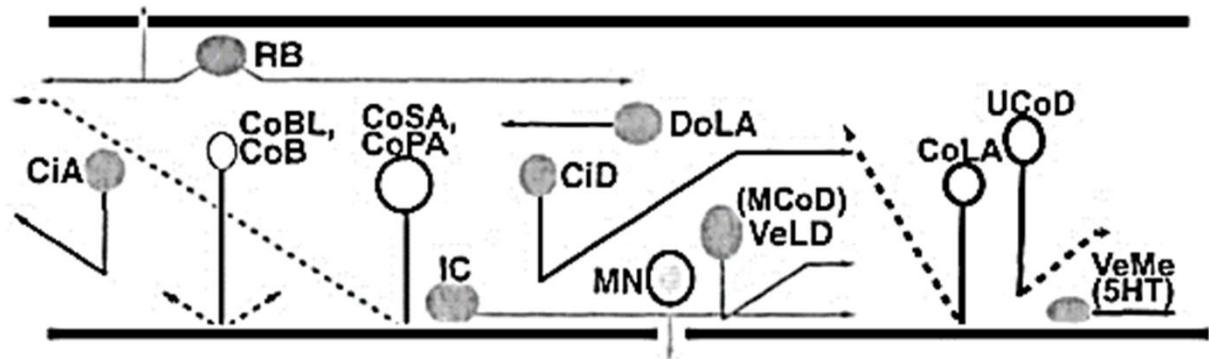
Drapeau et al, Progress in Neurobiology, 2002

# Using zebrafish in neurophysiology to correlate behavior with neuronal function

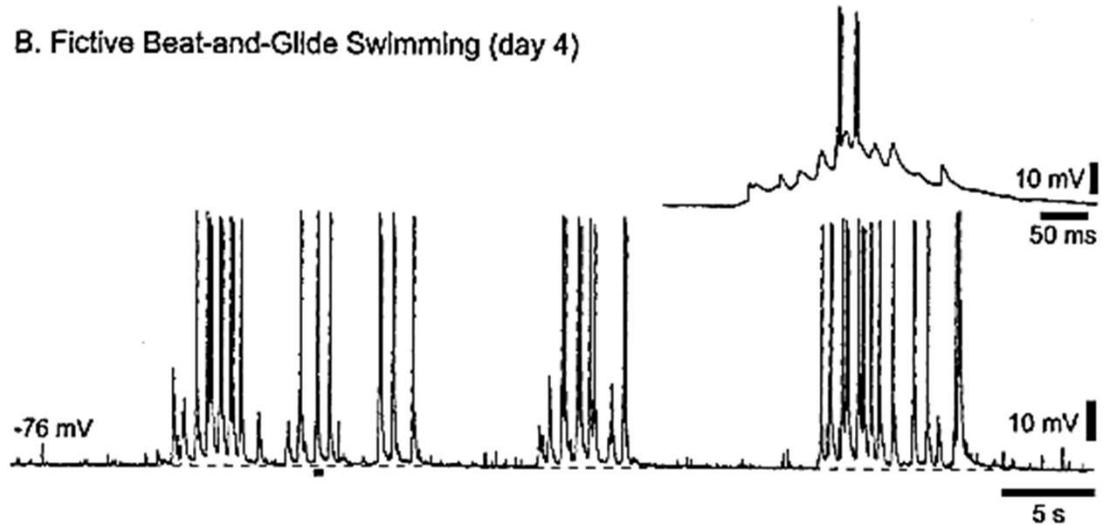
*4 days:  
Beat and glide  
swimming*



Maturation of inhibitory inputs to contralateral spinal network

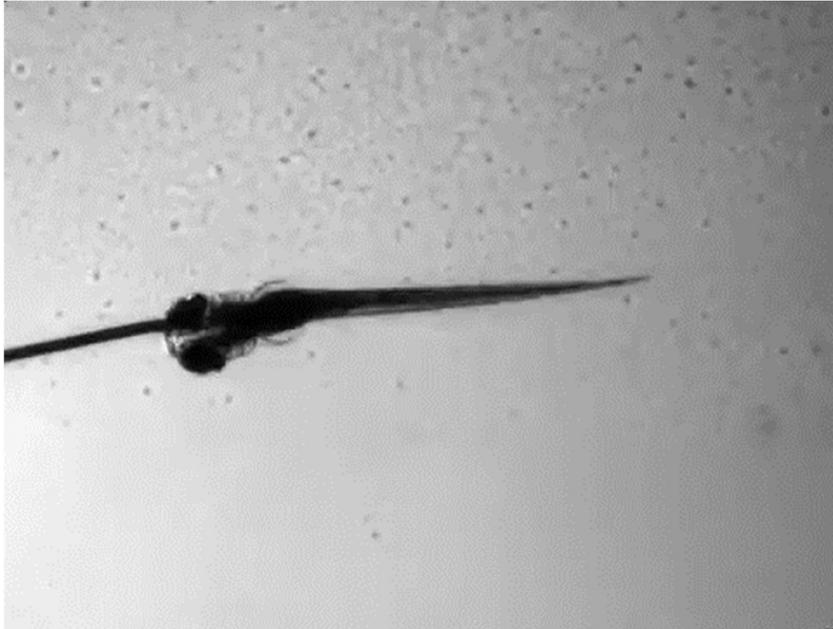


B. Fictive Beat-and-Glide Swimming (day 4)



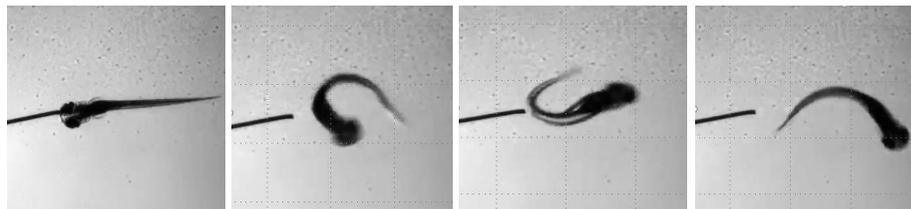
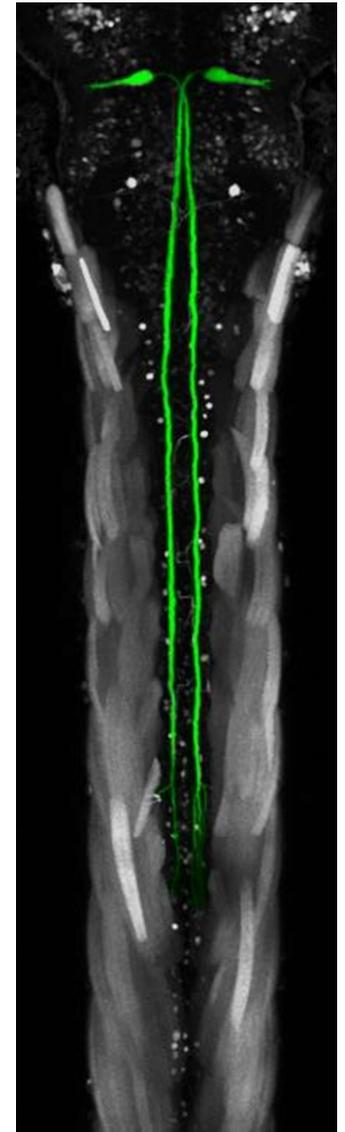
Drapeau et al, Progress in Neurobiology, 2002

# Using zebrafish in neurophysiology to correlate behavior with neuronal function

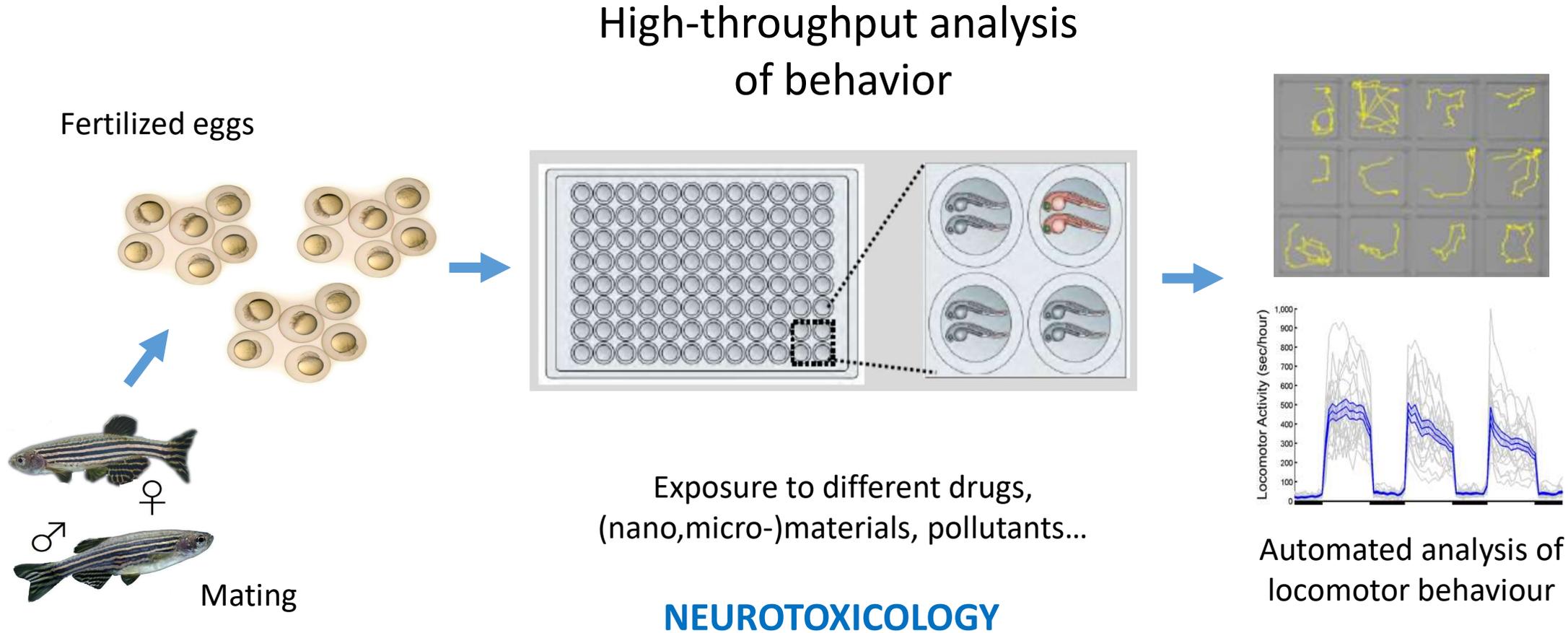


## *Touch response (C-start) = ESCAPE REFLEX*

- Sensory modulation of the behavior via Mauthner cells (M-cells, reticulo-spinal cells in the hindbrain)
- Activated by visual, tactile or acoustic stimuli
- Laser ablation of M-cells decreases and delays escape response

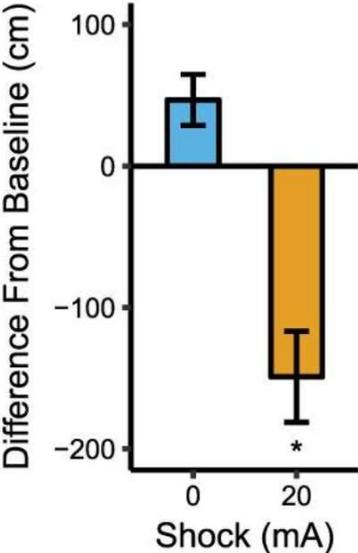
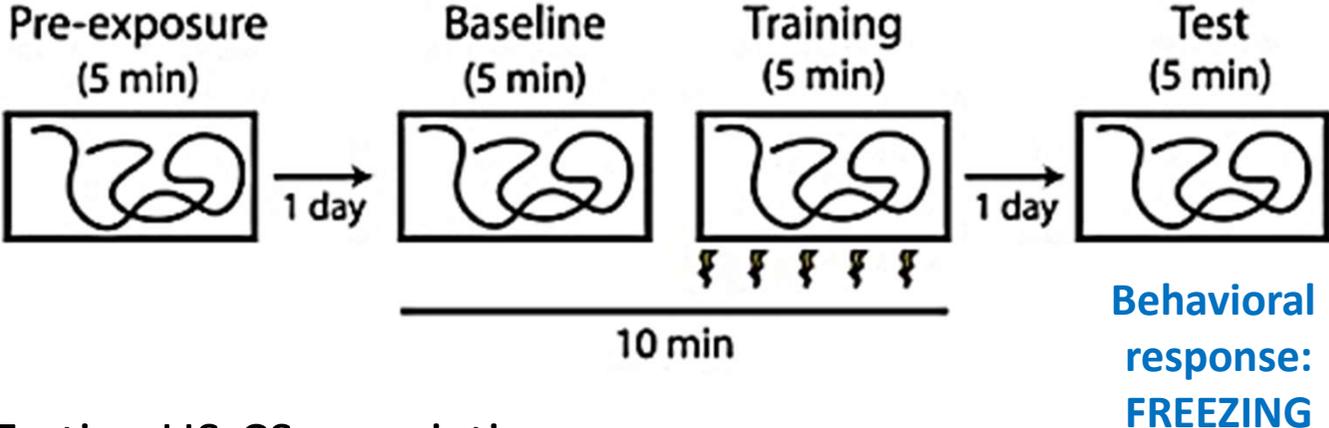


# Behavior can be used as a macroscopic readout of the neuronal function



# Studying complex behaviors in zebrafish: Contextual fear conditioning

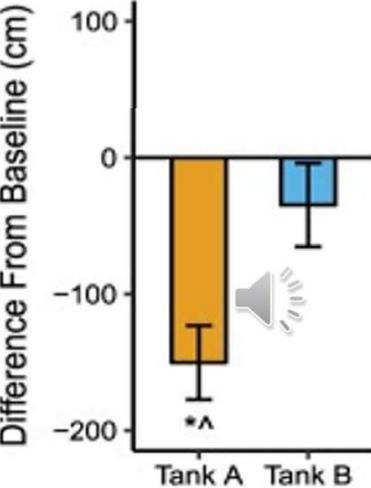
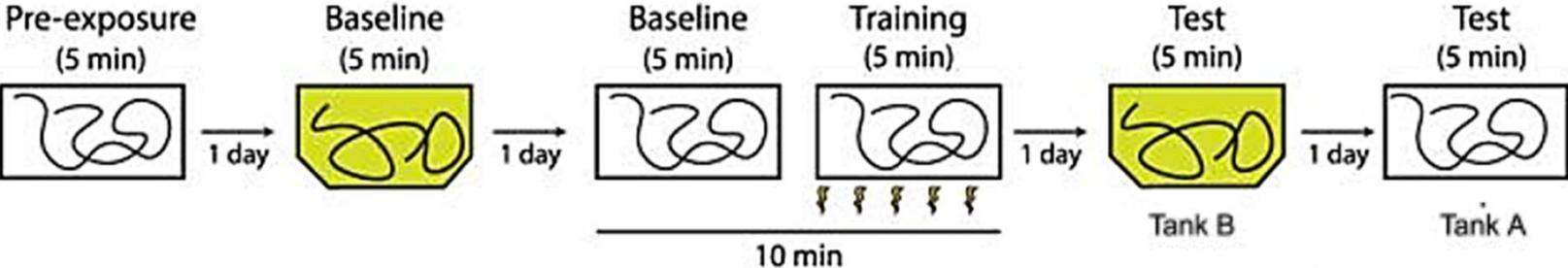
Testing US (electrical shock)



Testing US-CS association

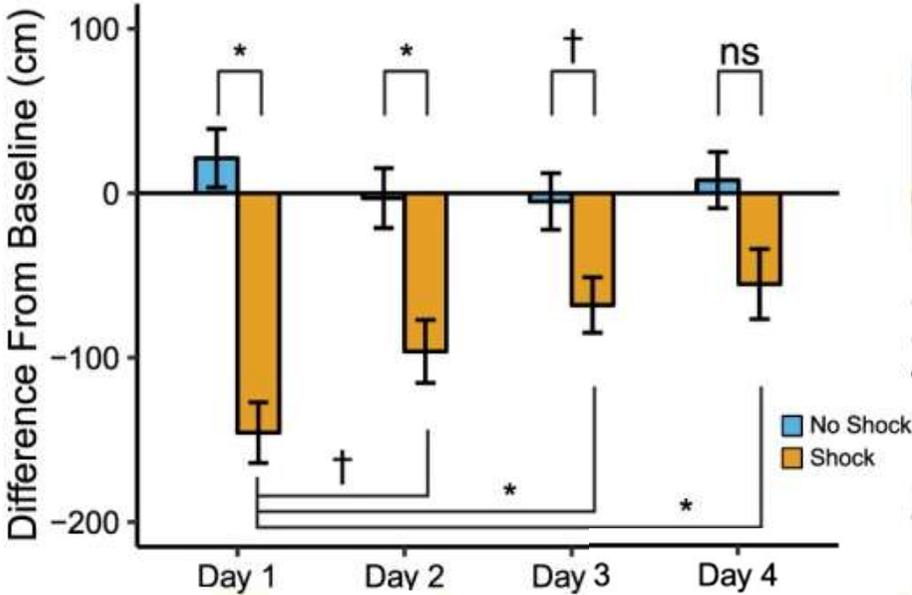
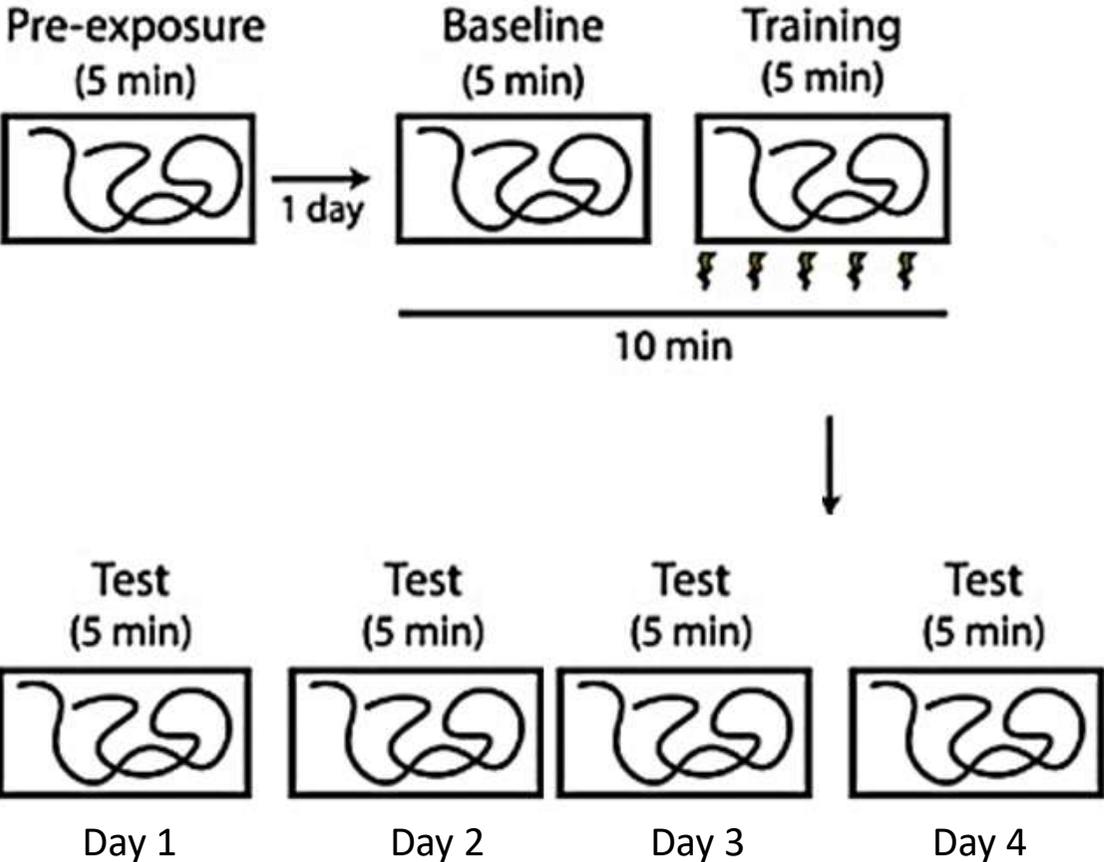
US = electrical shock

CS = context



Kenney et al, Learn Mem. 2017; doi: [10.1101/lm.045690.117](https://doi.org/10.1101/lm.045690.117)

# Studying complex behaviors in zebrafish: fear extinction



# How to identify neurons involved in complex neuronal functions?

## Traditional rodent model

Behavior



STAINING of  
genetic markers of neural activity  
(e.g. immediate early genes, c-fos)

- ✓ Only strongly activated cells can induce IEGs
- ✓ Neurons that are switched off are not detected

## Zebrafish

Behavior

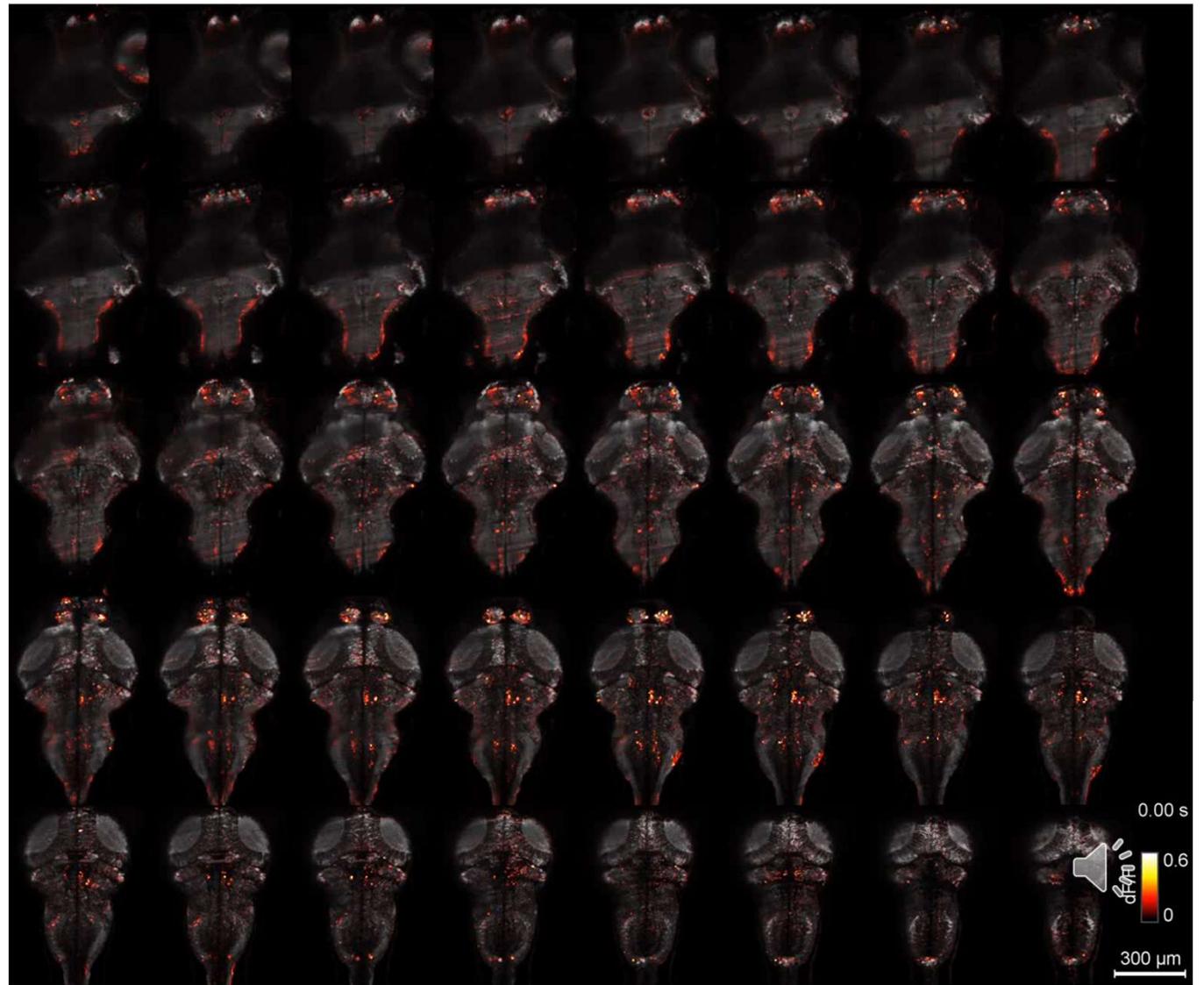
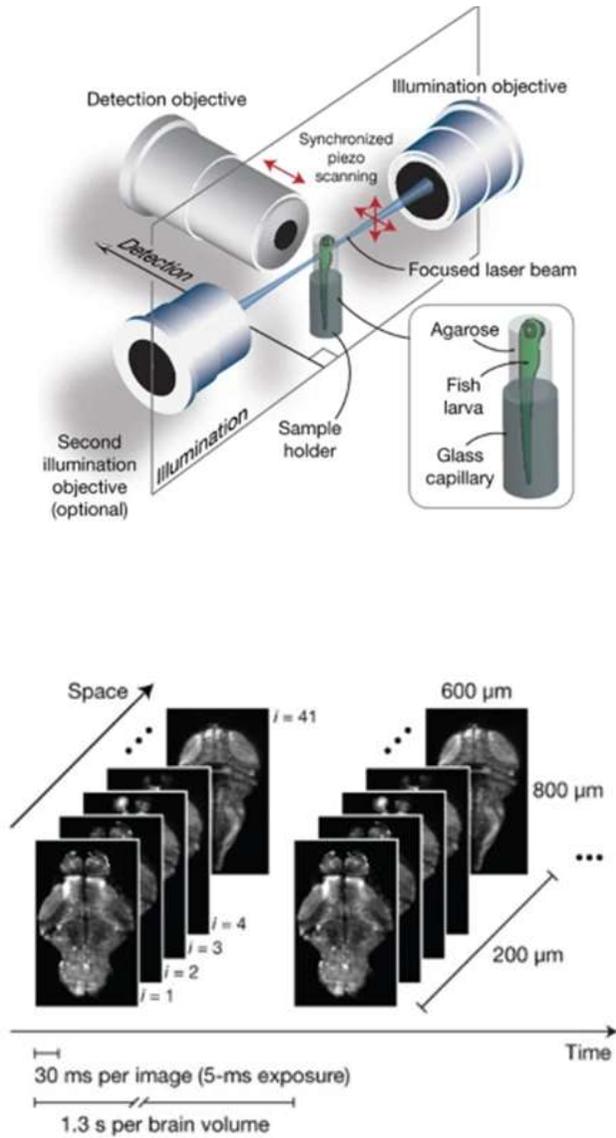


Monitoring live cell activity through  
genetically encoded Ca<sup>2+</sup> indicators

- ✓ Not invasive
- ✓ Small brain size and transparency allow simultaneous recording from hundreds of cells
- ✓ AP resolution



# Whole-brain functional imaging at cellular resolution using light-sheet microscopy.



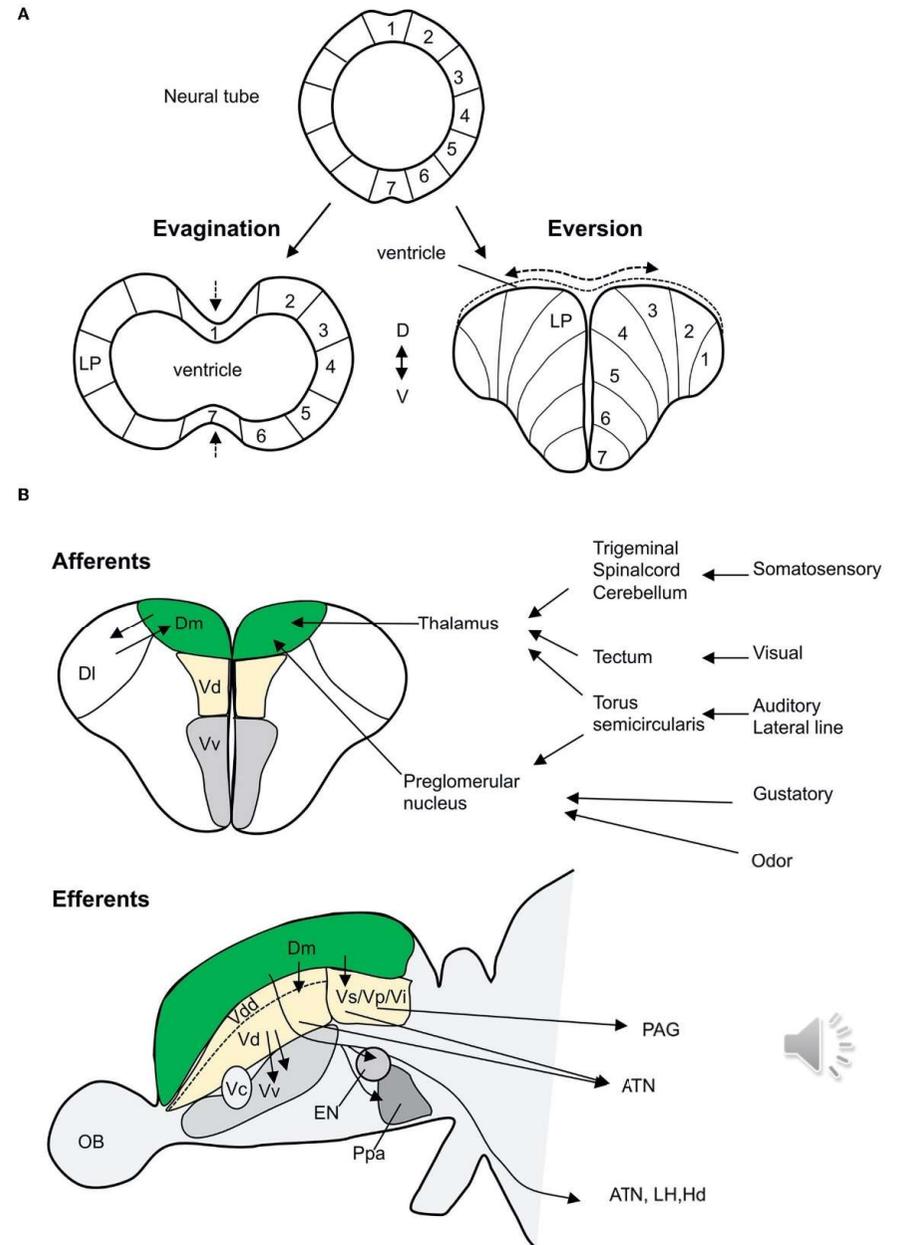
Ahrens, M. B., and Keller, P. J. (2013). Nat. Methods doi: 10.1038/nmeth.2434

# Functional Anatomy of Zebrafish Amygdala

Basolateral complex of Amygdala  
 = Medial part of dorsal telencephalon  
 (Dm) (also called dorsal pallium)

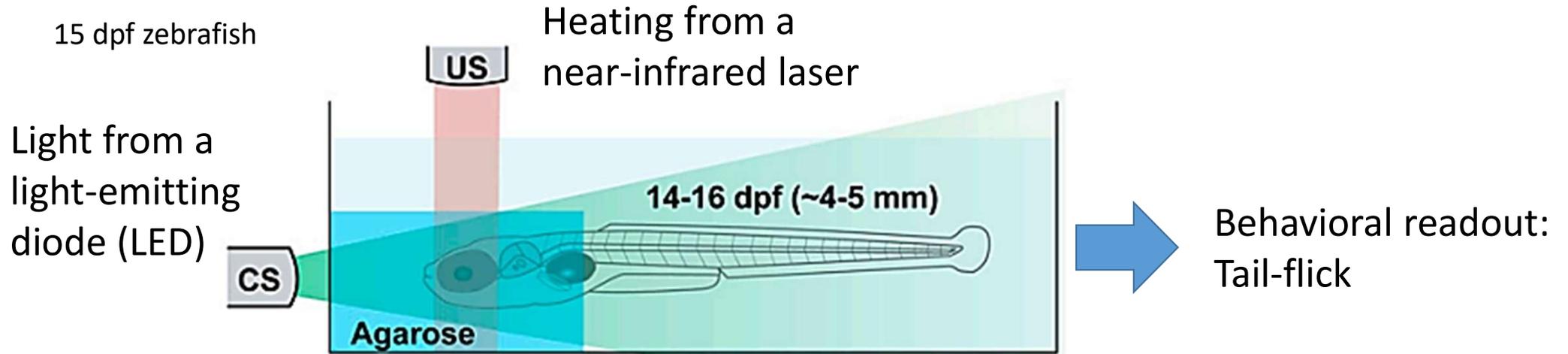
Central complex of the Amygdala  
 = Ventral telencephalon (dorsal nucleus  
 (Vd), supracommissural nucleus (Vs),  
 and postcommissural nucleus (Vp))

Hippocampus = lateral  
 telencephalon (DI)

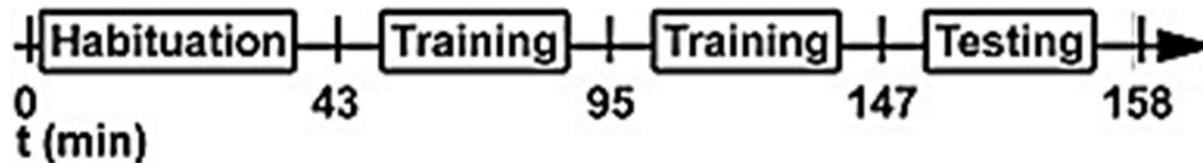


# Investigating associative learning induced brain changes in in vivo zebrafish

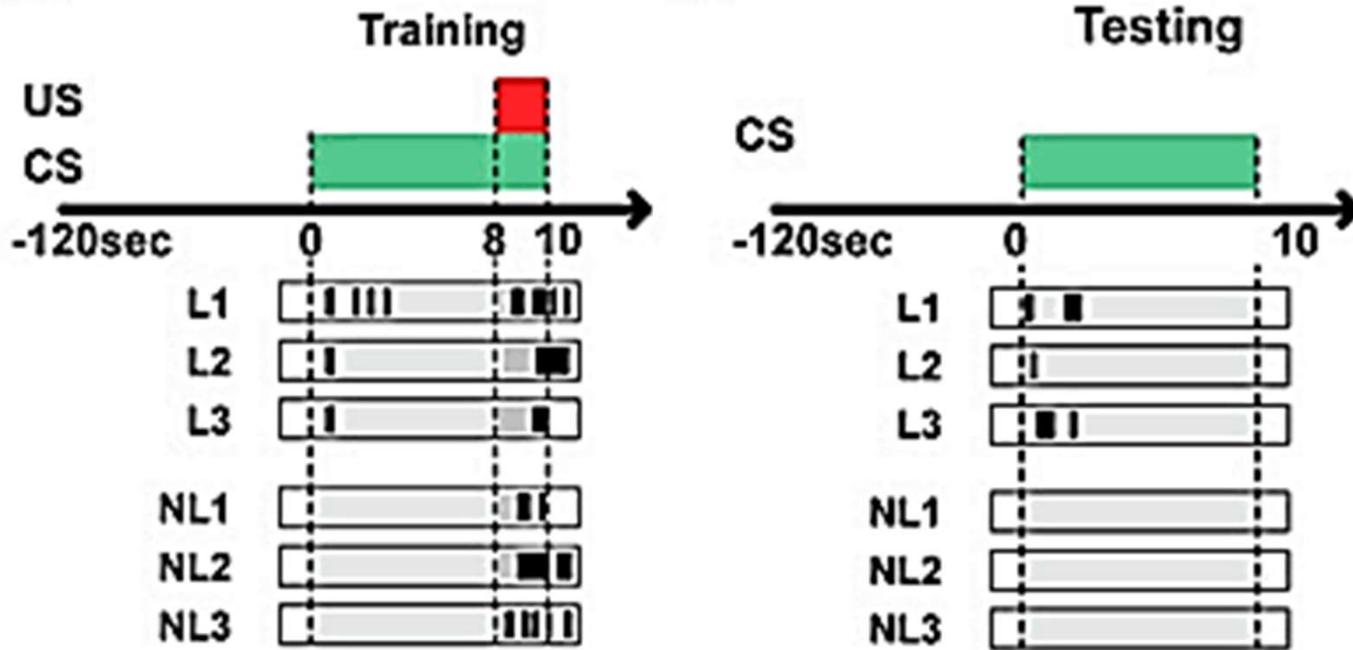
## Cued aversive conditioning



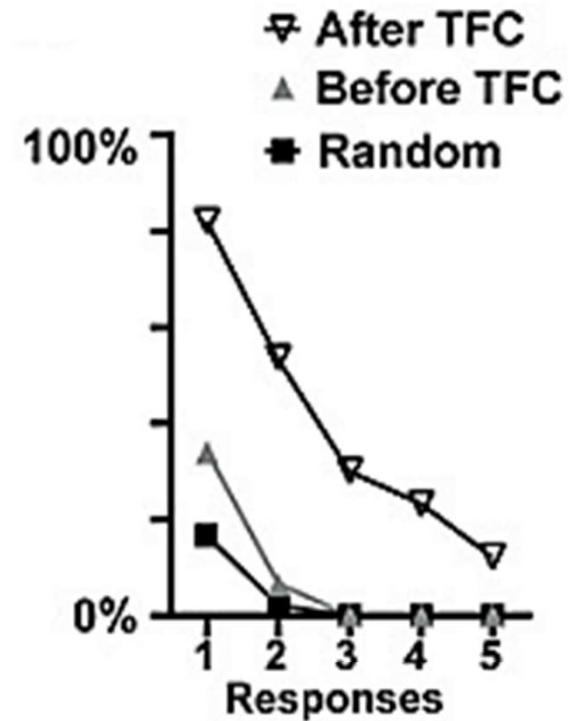
## Tail Flick Conditioning



# Investigating associative learning induced brain changes in in vivo zebrafish



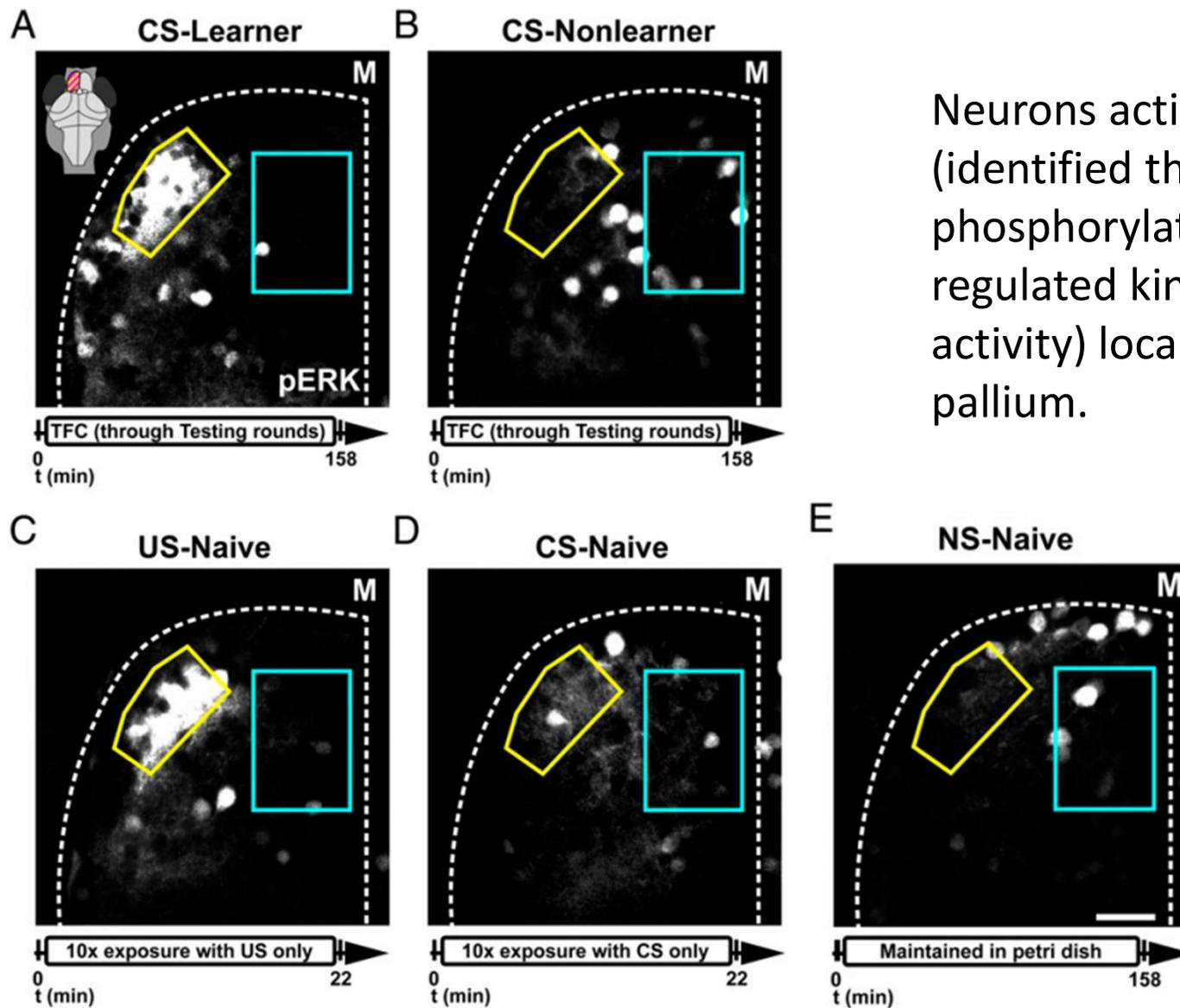
L = Learners  
 NL = Nonlearners



Tail flick conditioning is:

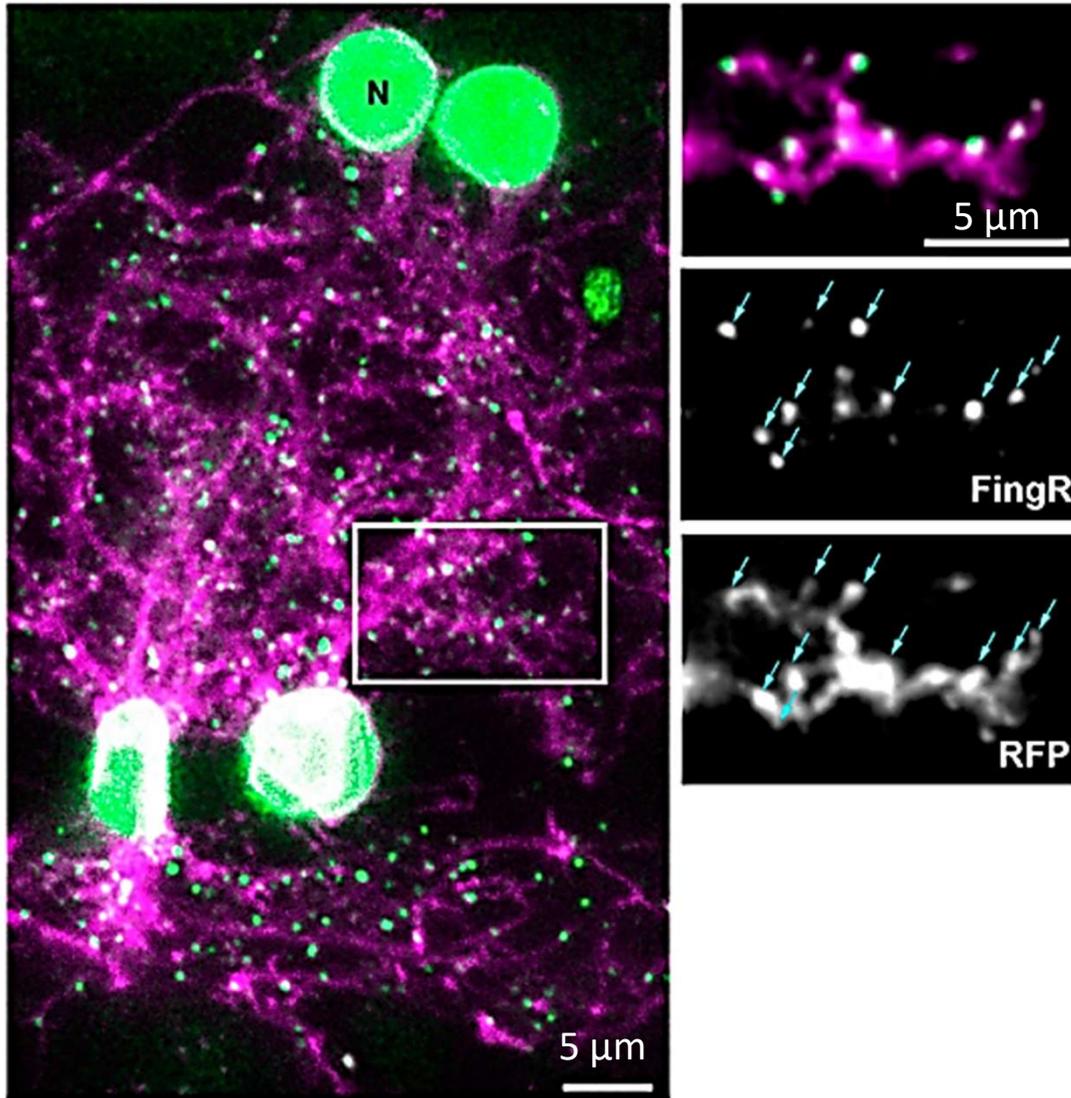
- ✓ NMDA receptor dependent
- ✓ Sensitive to extinction





Neurons active during conditioning (identified through immunostaining for the phosphorylated extracellular signal-regulated kinase, pERK, marker of neuronal activity) localize in the antero-lateral dorsal pallidum.





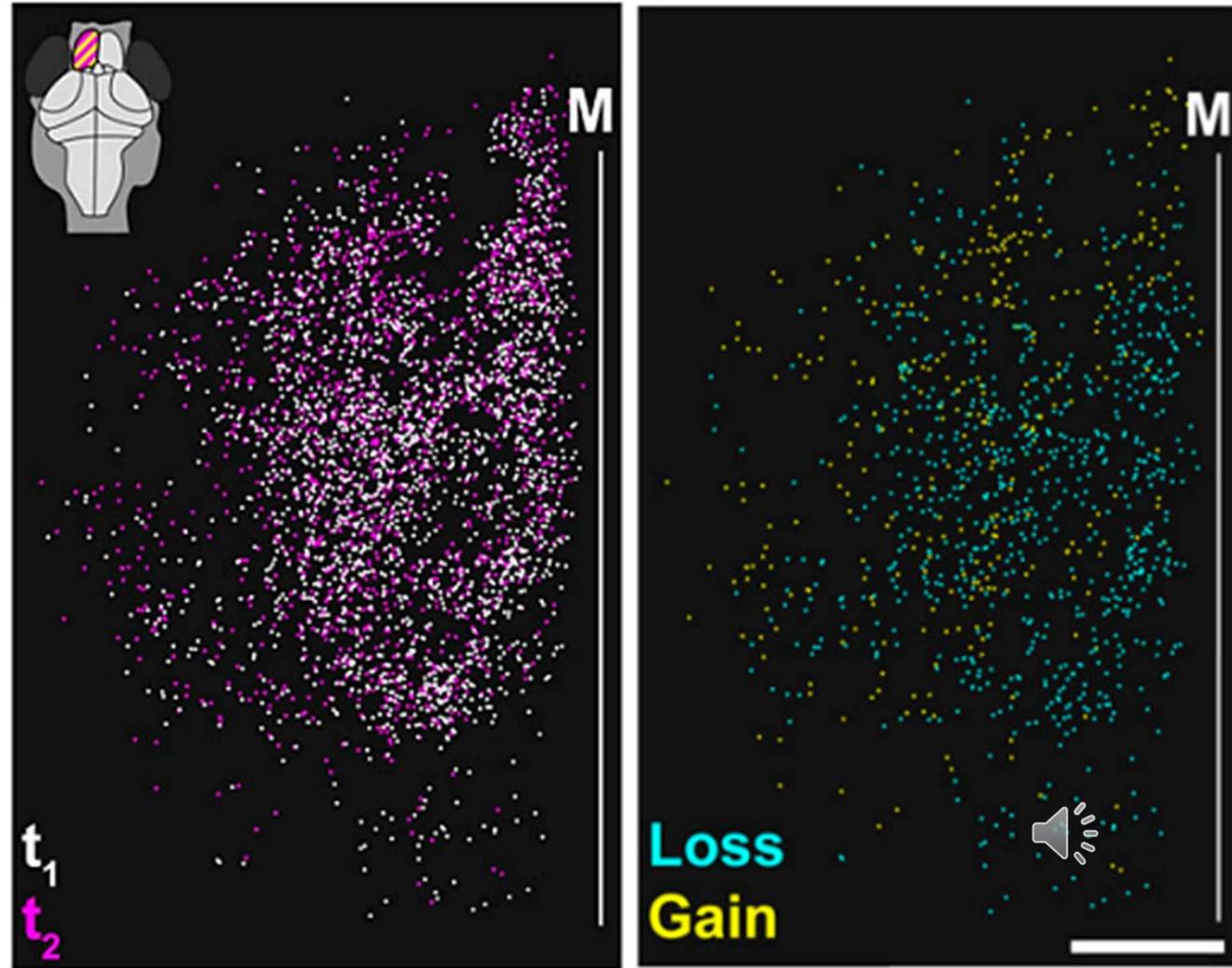
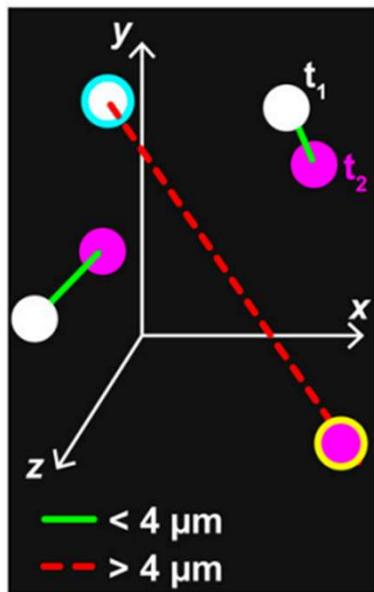
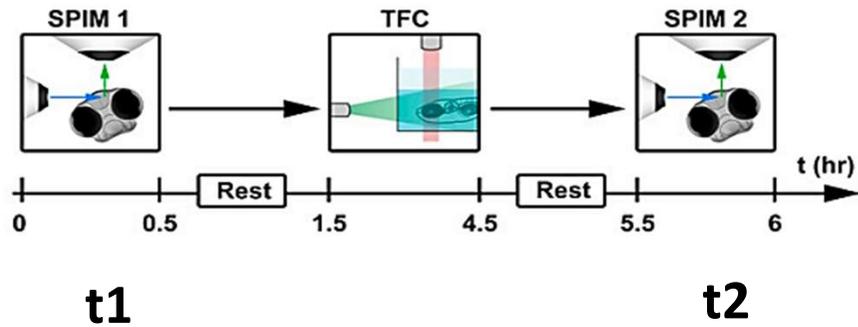
Live visualization of excitatory synapses in dorsal pallium by using transgenic zebrafish co-expressing:

- PSD-95.FingR-GFP (green)
- membrane-targeted mScarlet (magenta)

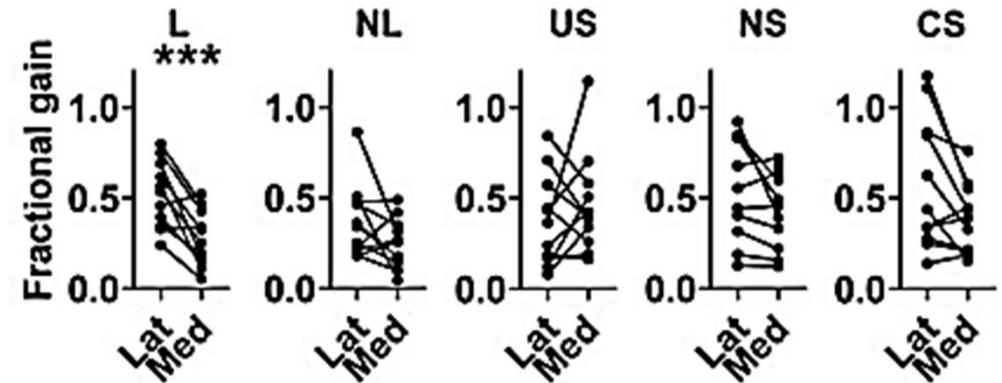
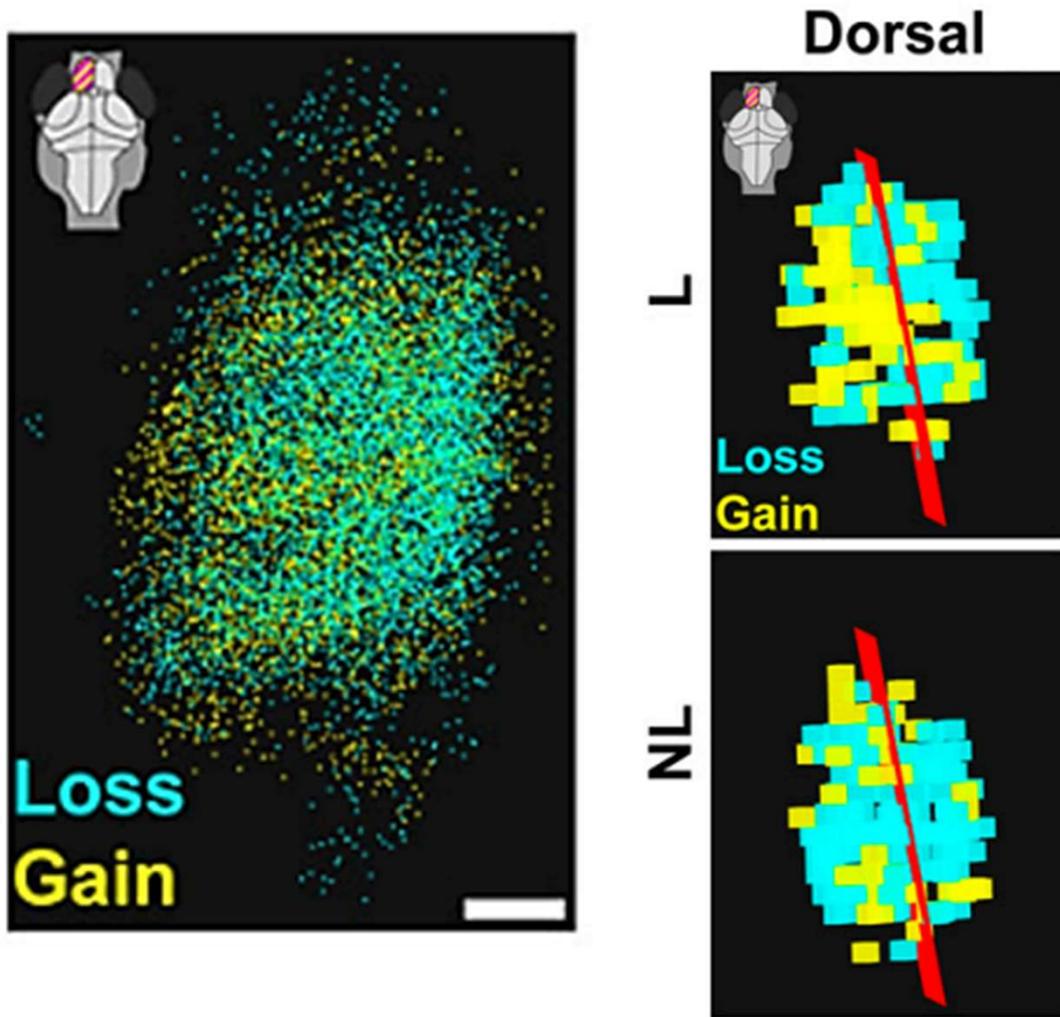


Selective plane illumination microscopy (SPIM)

# A computational approach applied to in vivo observation of zebrafish dorsal pallium during conditioning



...however, their distribution across the dorsal pallium is modified



Regional differences in synapse formation in the pallium of learner fish: new synapses are formed only in the lateral pallium



## Conclusions



- ✓ Zebrafish present several advantages for neurophysiological studies
  - Multiple techniques for multiple level of resolution of neuronal functions
- ✓ Studying complex behavior and neuronal correlates
  - genetically encoded calcium indicators
  - Synaptic resolution

