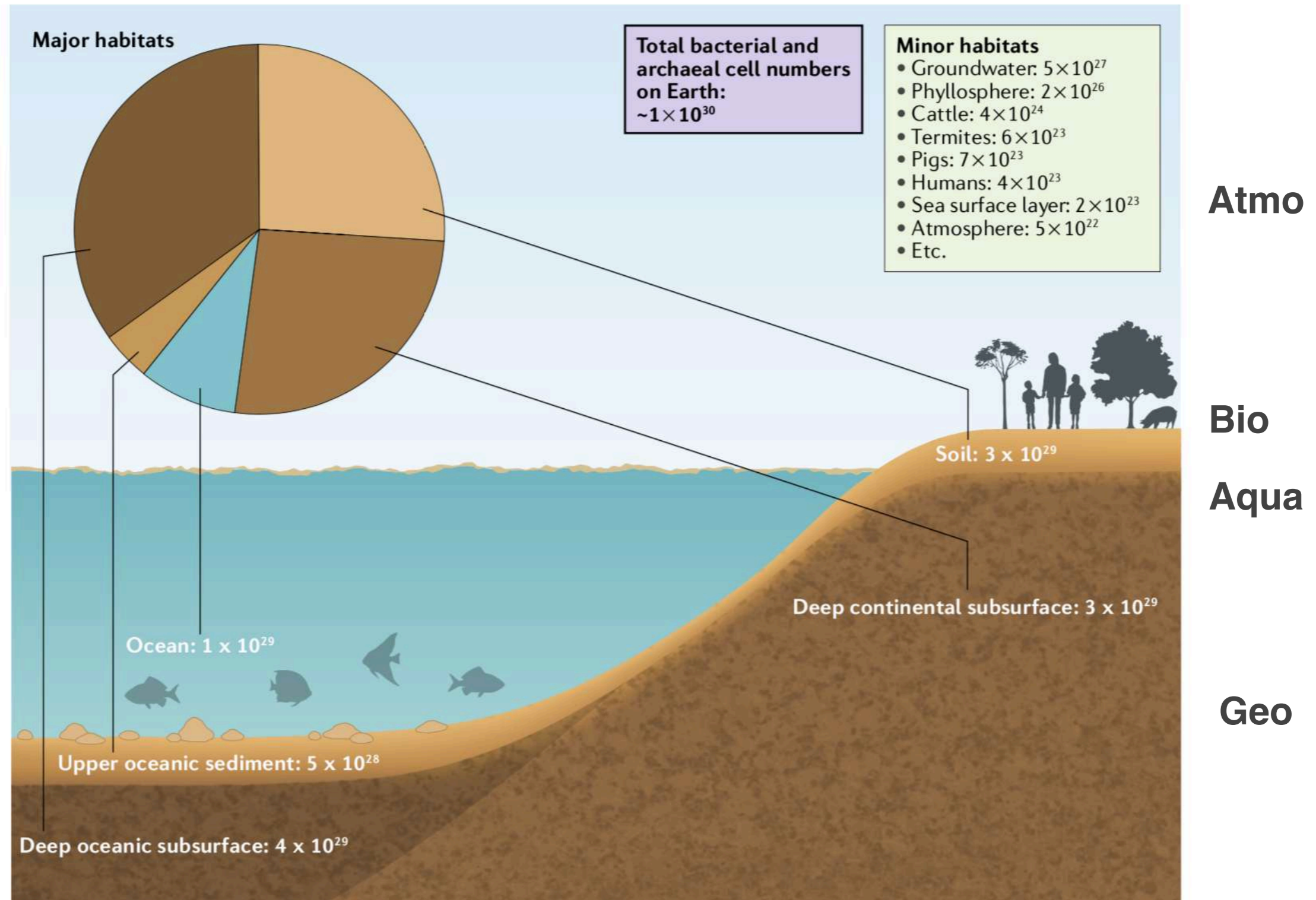


L07c

Microbial Ecology

- Study of the **interactions** of microorganisms with their **environment (including organic matter), each other**, and plant and animal species (**other organisms**)
—> symbioses, biogeochemical cycles, climate change
- **Marine sediment ecosystem**
- **Ocean ecosystem**
- **Freshwater ecosystem**

Microbial abundance



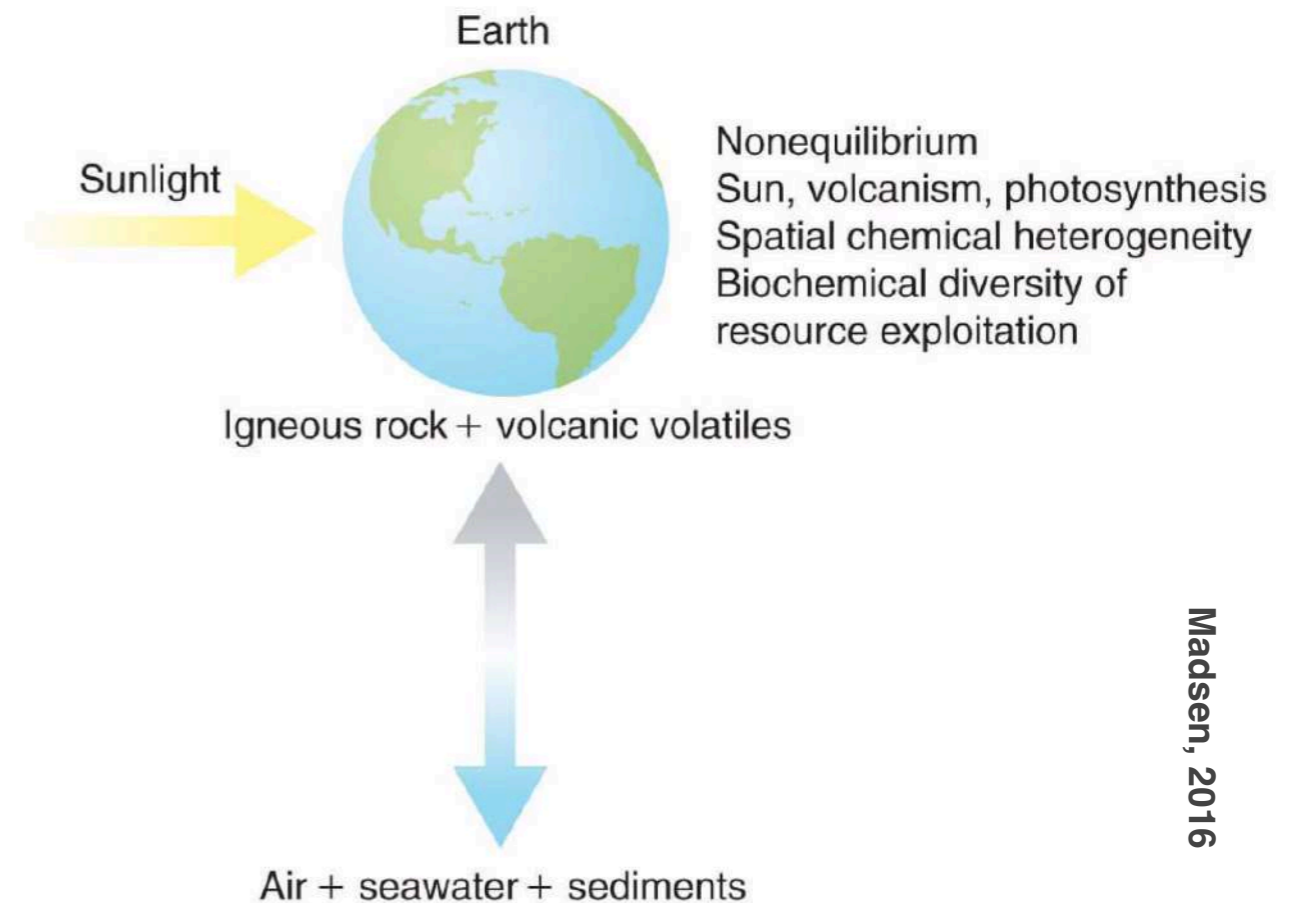
Flemming & Wuertz, 2019

MARINE SEDIMENTS

OCEAN FLOOR

Marine sediments as a microbial ecosystem

- Global view of Earth: a heterogeneous mixture of rocks, water, gases, organisms is maintained in a non equilibrium state by sunlight and volcanism
- Hydrothermal vents, cold seeps are oasis of primary production and microbial and biochemical diversity



Madsen, 2016

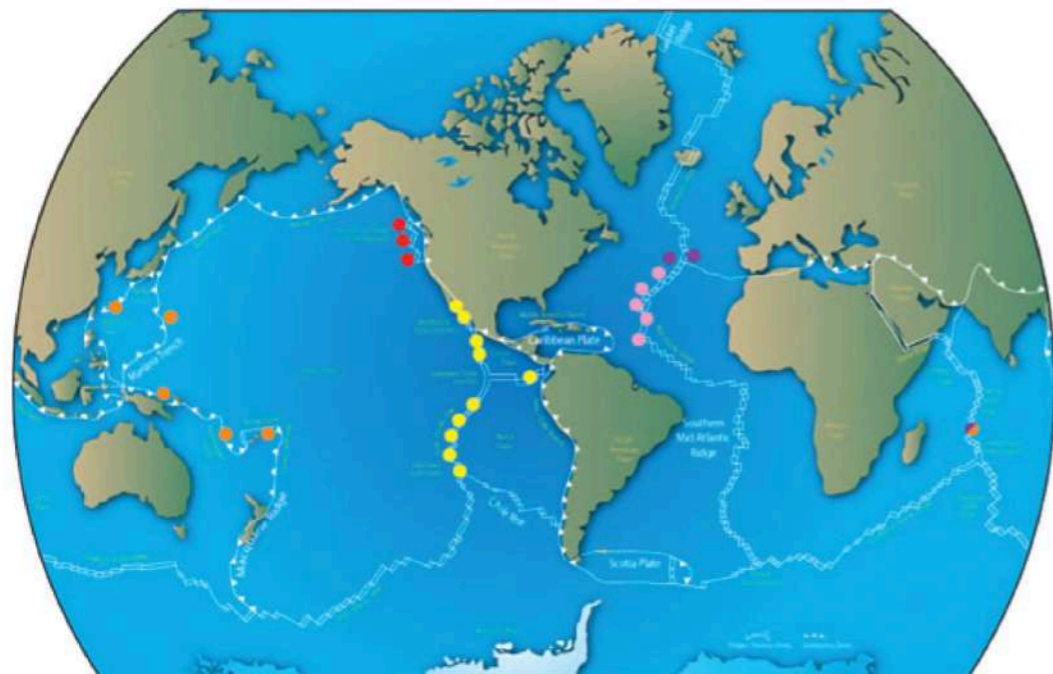


Figure 4.10 Plate tectonic map of the globe showing major plate boundaries, mid-ocean ridges, and distribution of hydrothermal vent sites. Colored circles show vents with similar animal communities. (Courtesy of E. Paul Oberlander, with permission from Woods Hole Oceanographic Institute.)

- Approximately 1×10^9 microbes in 1 g of upper sediment
- Biofilm

Microbial metabolic pathways shaping Earth ecosystem

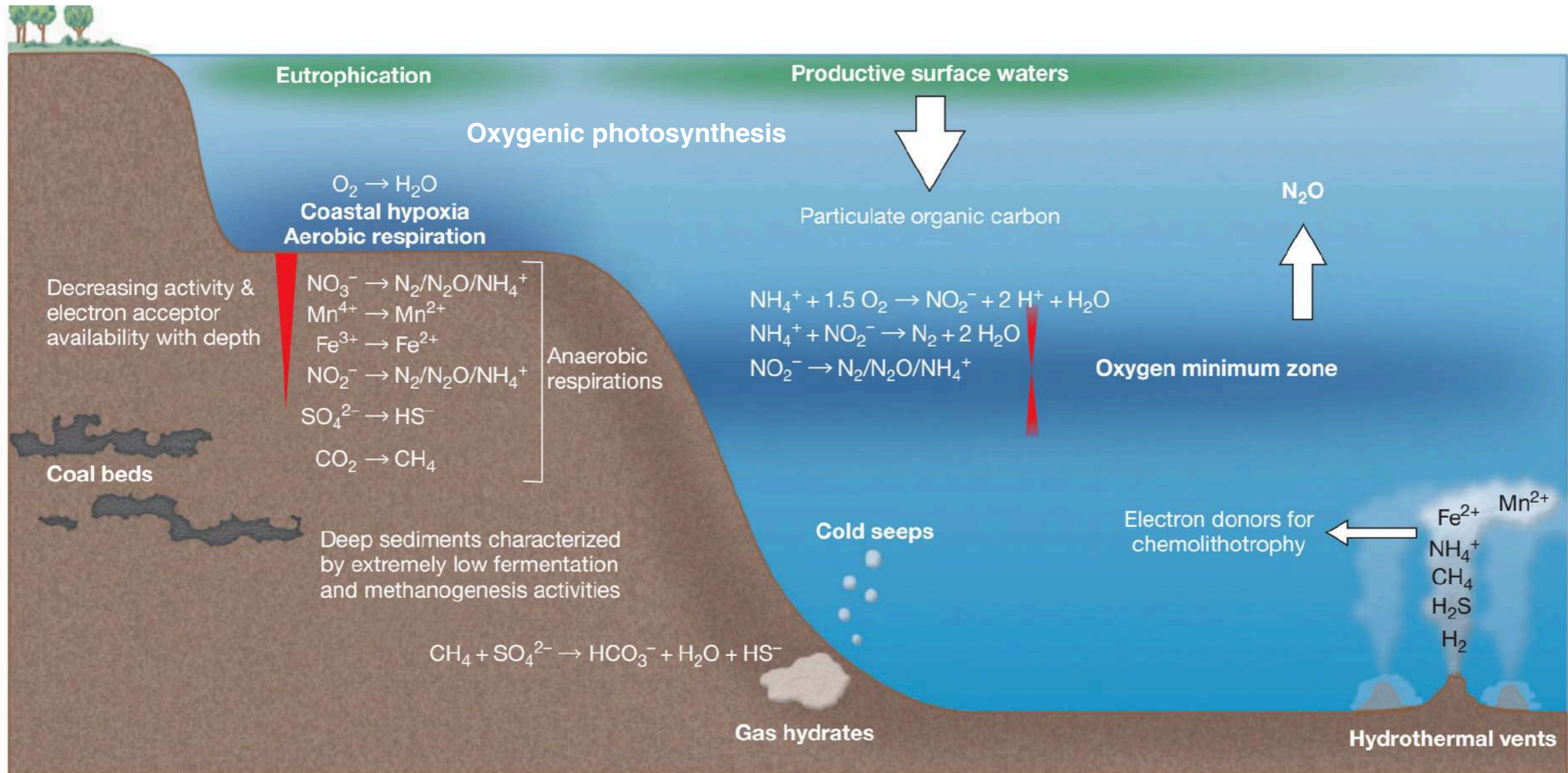


Figure 20.20 Diversity of marine systems and associated microbial metabolic processes. Decreasing electron acceptor availability with depth into the sediment or with increasing distance into an oxygen minimum zone is indicated by red wedges. Sulfate becomes limiting only at greater depths in marine sediments. The indicated metabolic diversity is covered in Chapter 14.

Madigan et al. 2018

Terrestrial vs marine fingerprint

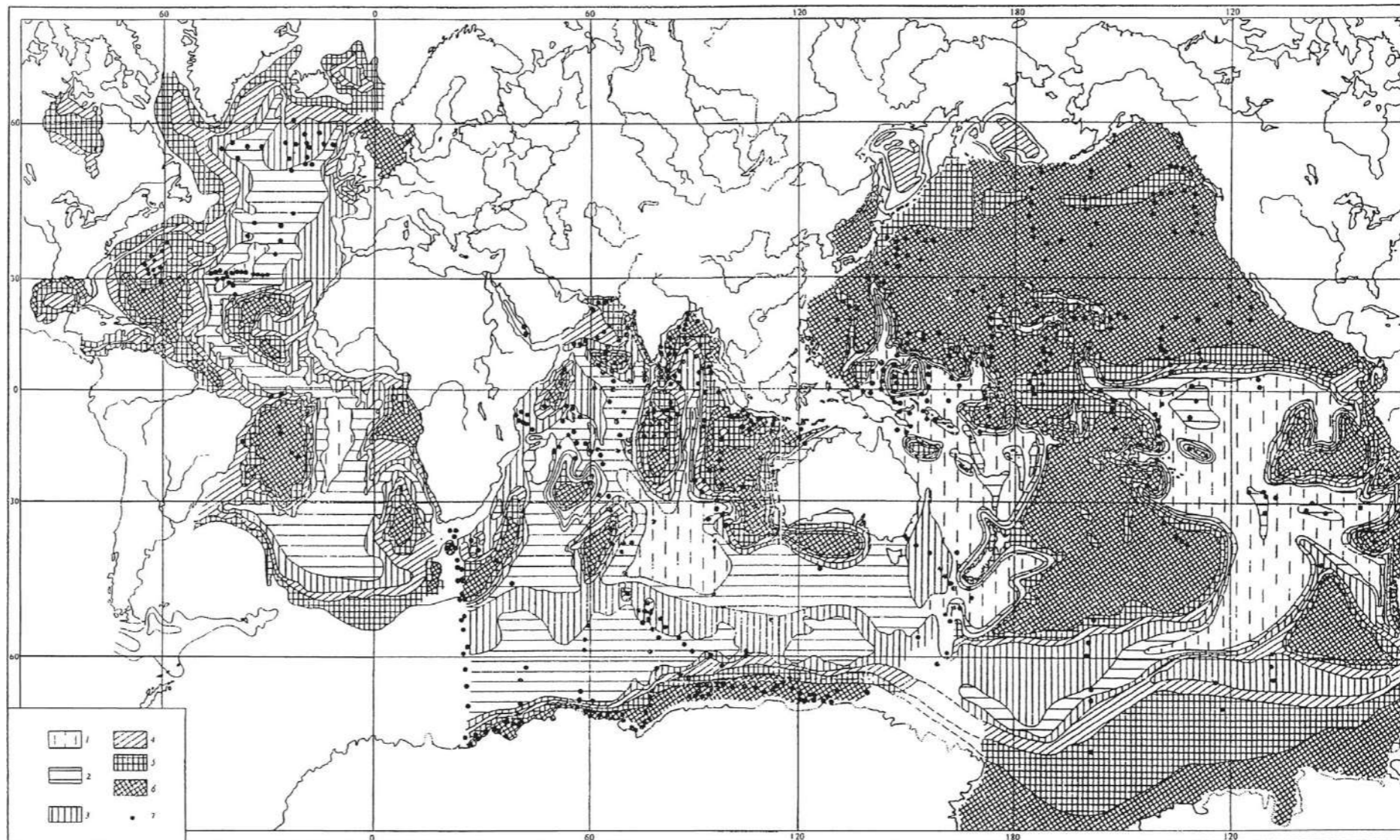
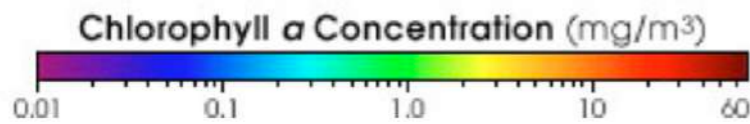
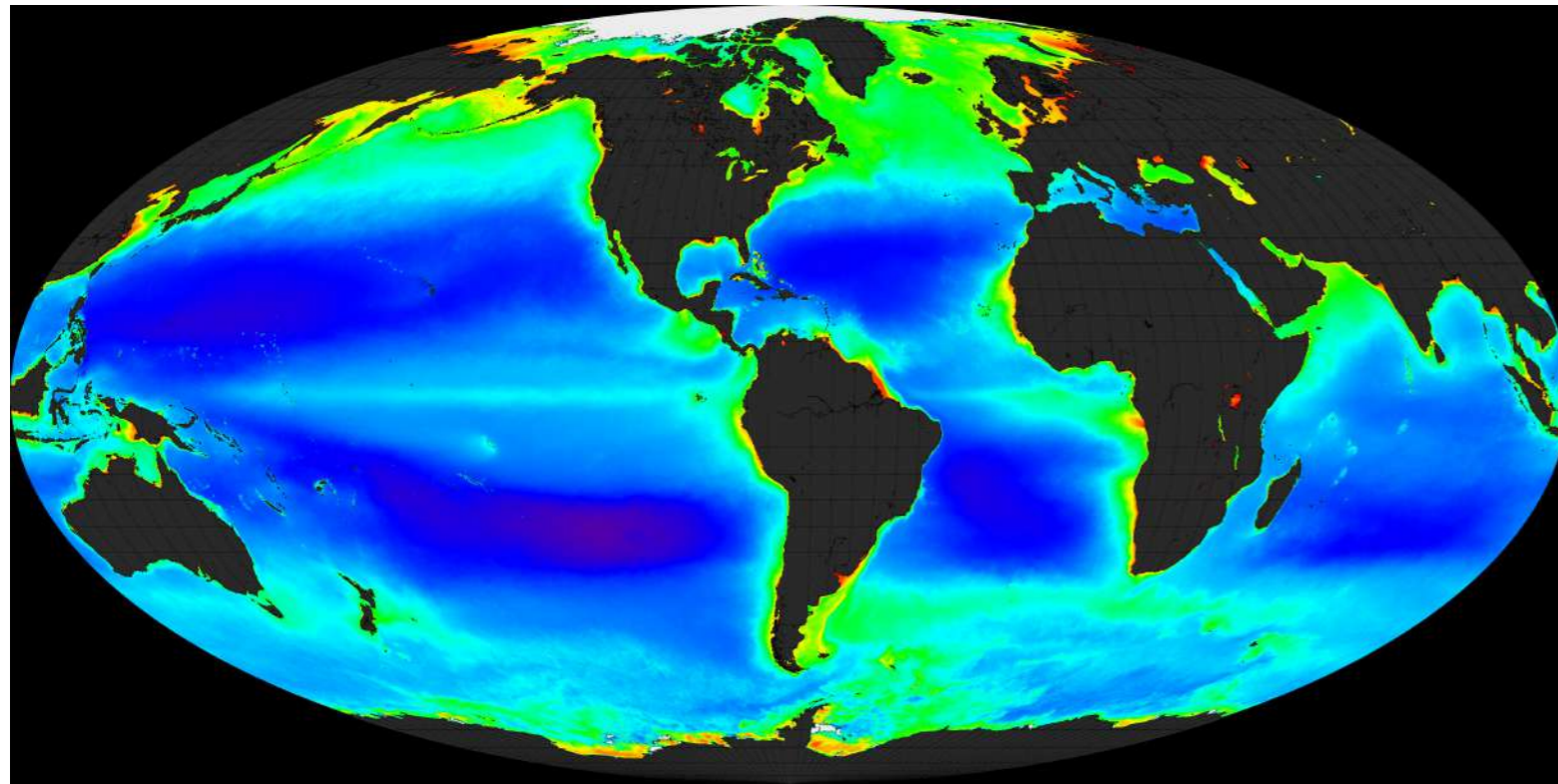


Figure 4.11 Composition of ocean sediments: percent derived from continents (terrigenous) versus autochthonous biotic sources. The six boxes in the key (numbered 1–6) show values from <10% (1, lightest shading) to >90% (6, darkest shading) terrigenous sediment. (From Lisitzin, A.P. 1996. *Oceanic Sedimentation*, pp. 28–29. American Geophysical Union, Washington, DC. Copyright 1996, American Geophysical Union. Reproduced by permission of American Geophysical Union.)

- Marine sediments are influenced by terrestrial ecosystem and water column ecosystem
- Marine sediments: clay-silt-sand
- Marine sediments are very dynamic

Ocean surface photosynthesis dictates sedimentation



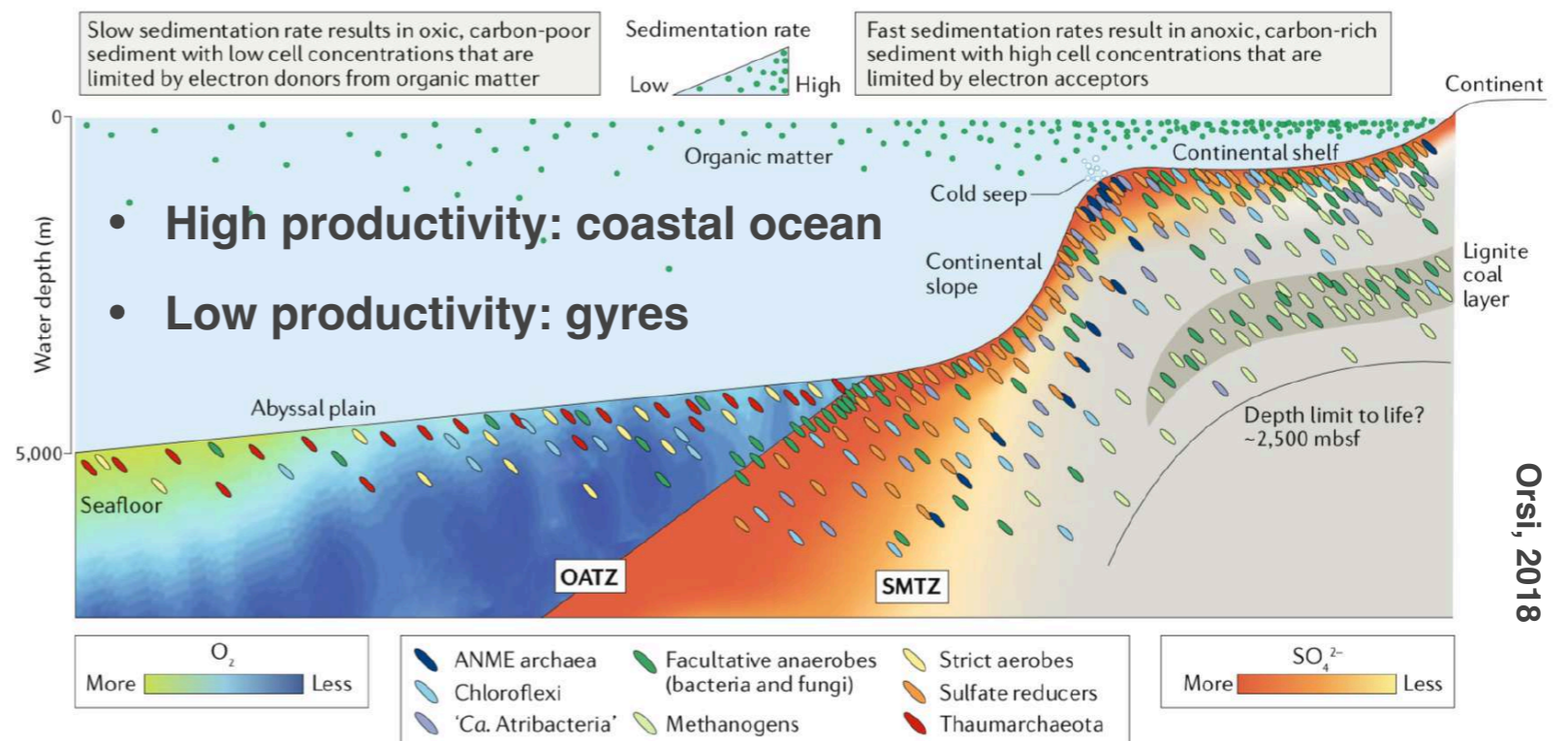
NOAA

- Chlorophyll data from satellite to inform on photosynthesis → organic matter production by microscopic algae (phytoplankton) and cyanobacteria
- Life and death in the upper ocean create a down-ward flux of organic matter and associated minerals

Dead organisms
Fecal pellets

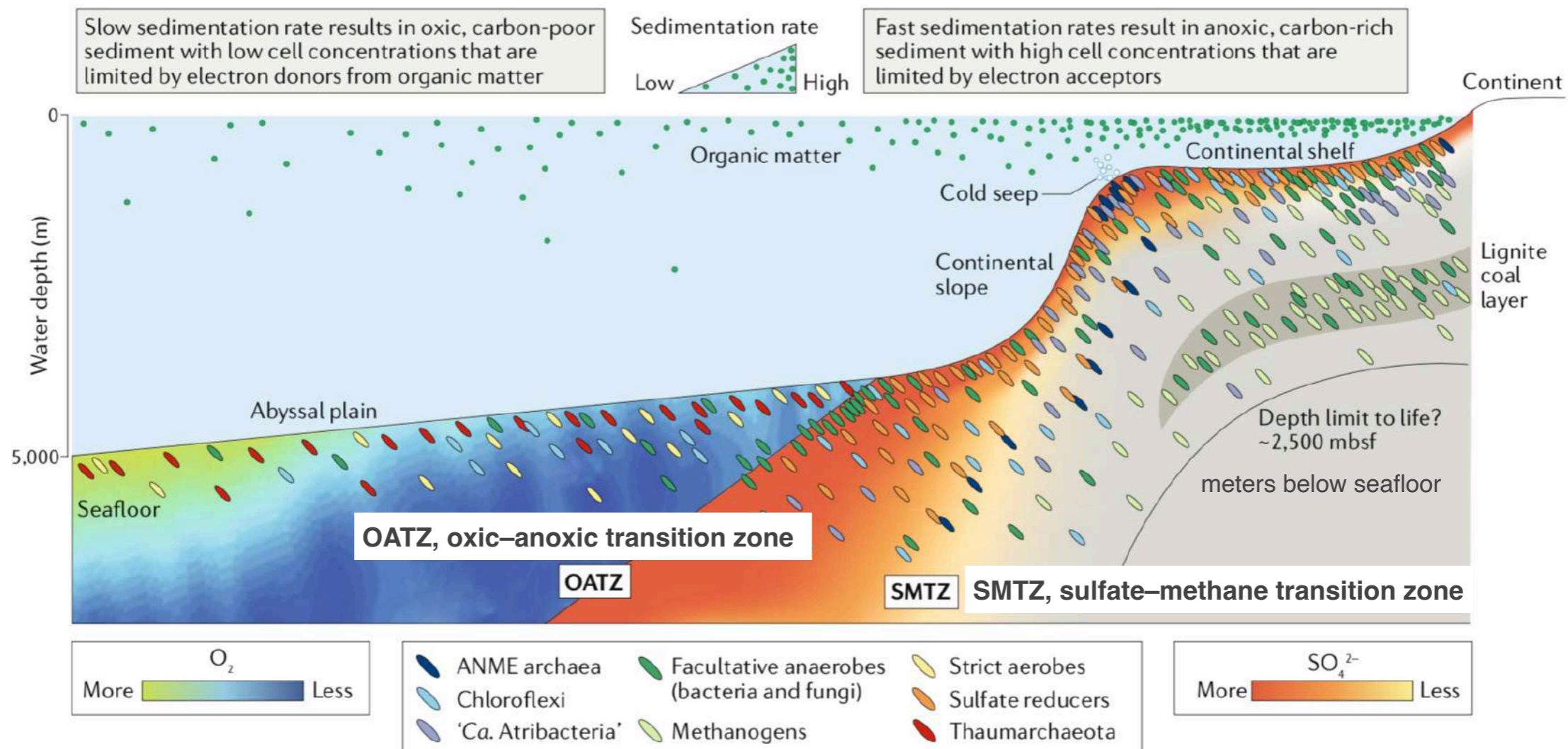


Marine snow



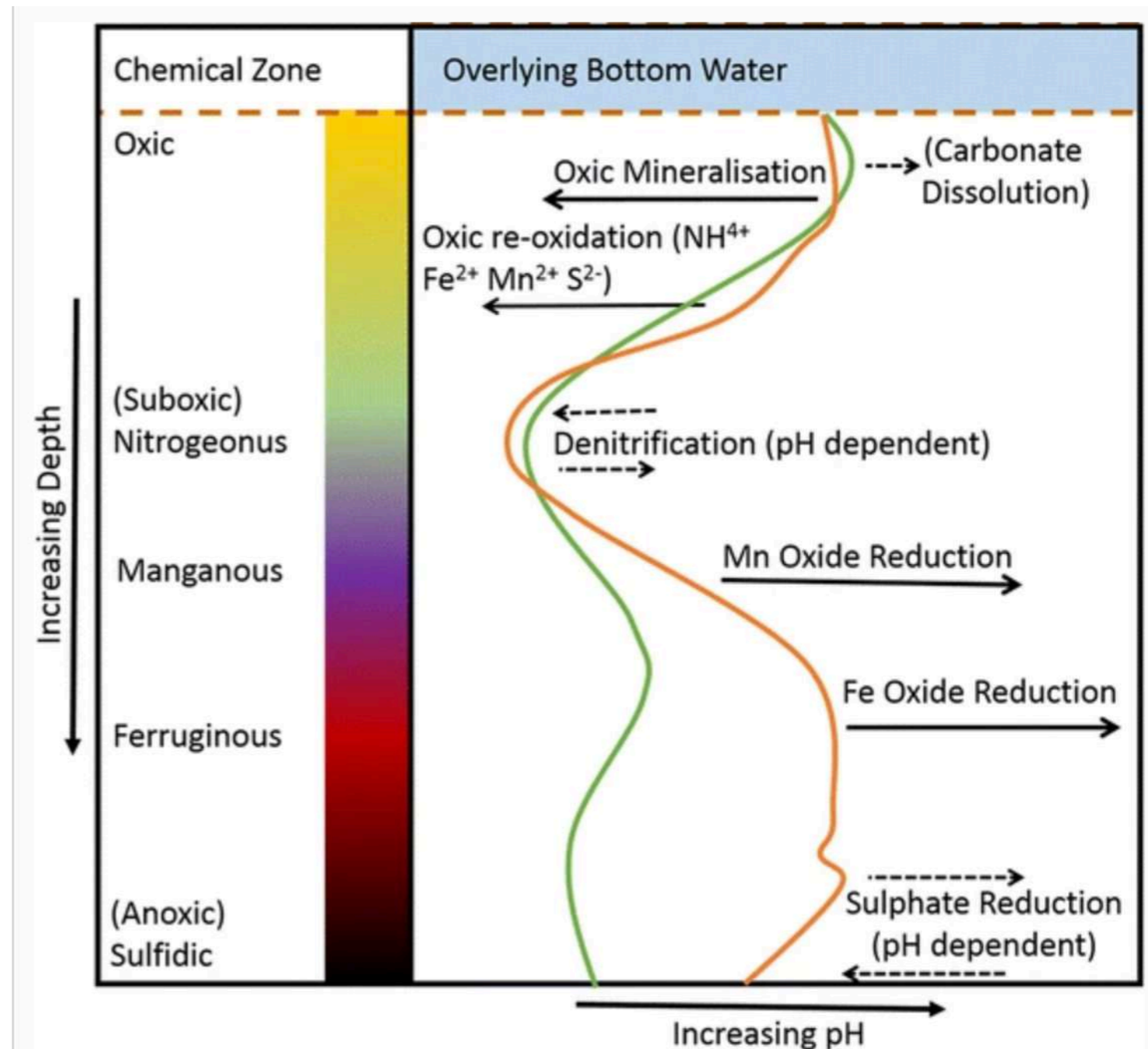
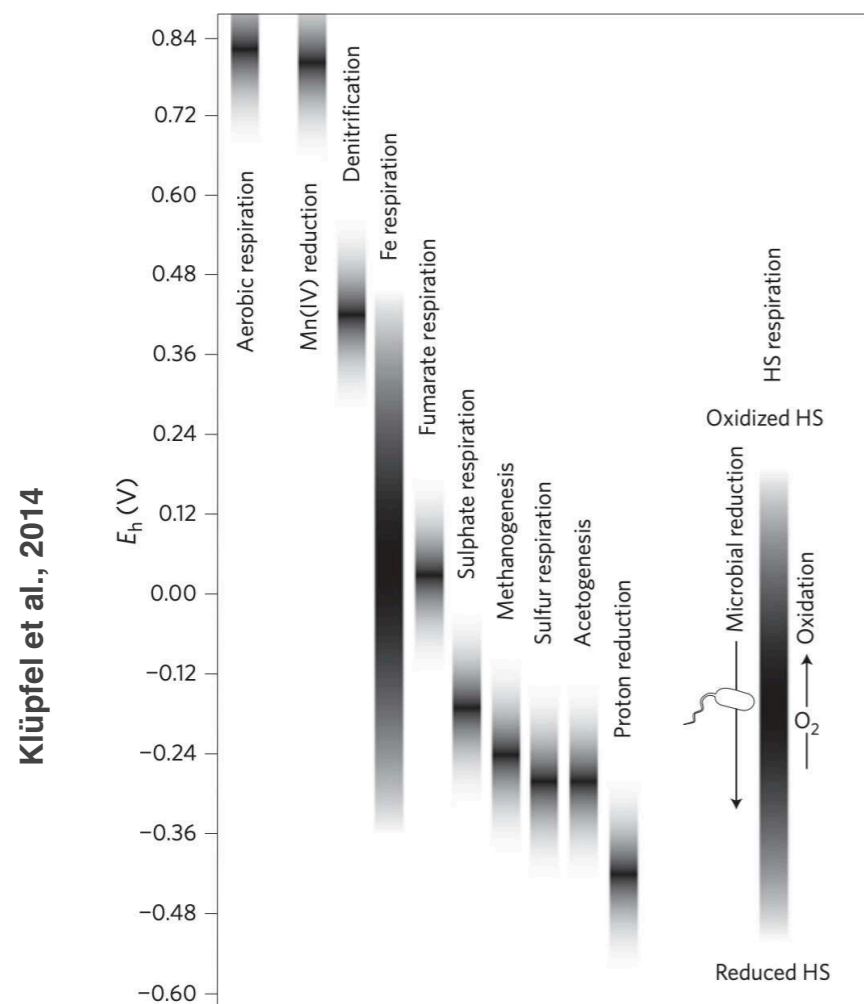
Sedimentation rate

- Redox conditions in seafloor and subseafloor ecosystems are linked to sedimentation rate and distance from shore
- Ultra-low rates of organic matter sedimentation under oligotrophic gyres lead to deep O₂ penetration (left), and consequently, the survival of microbial communities is limited by availability of electron donors from organic matter
- Faster sedimentation rates closer to shore cause anoxic conditions shortly below the seafloor owing to the rapid consumption of oxygen for organic matter oxidation at the sediment surface (right), terminal electron acceptors—> sulfate



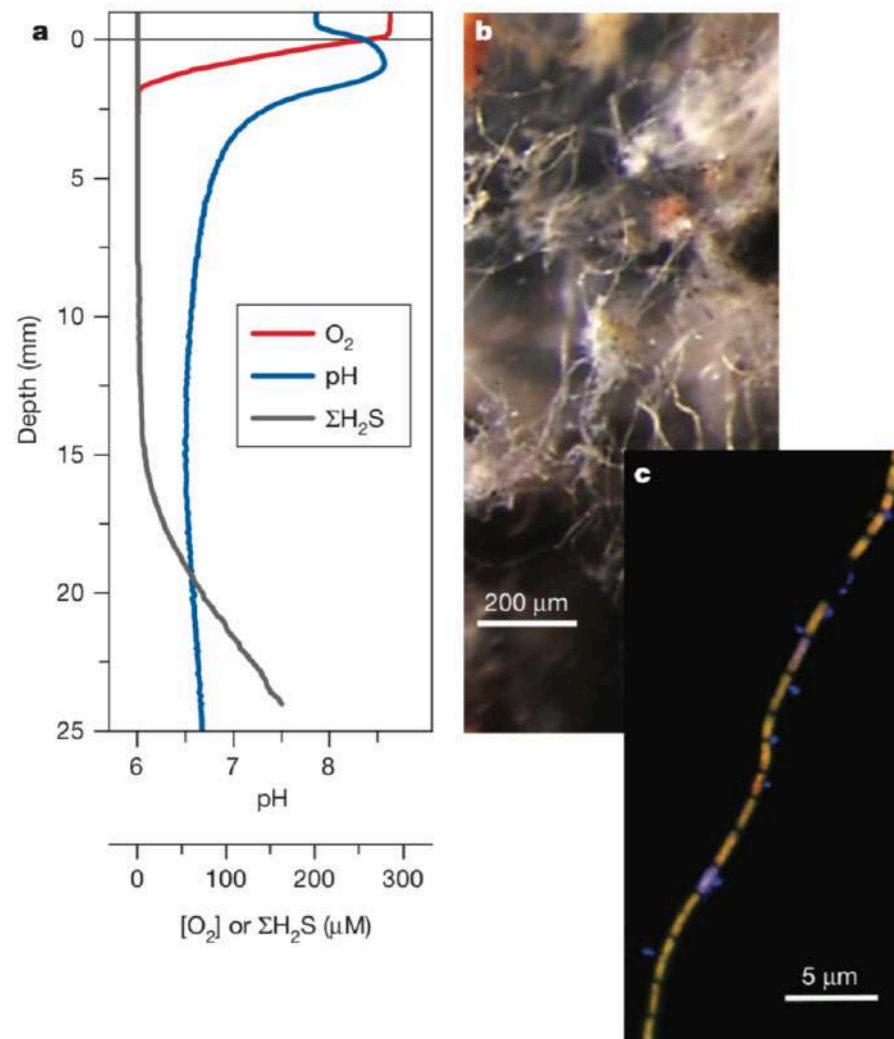
Sediment: Organic matter decomposition/respiration → nutrient recycling

- Microbes able to respire in multiple ways will always choose available acceptors with the biggest potential difference to the donor

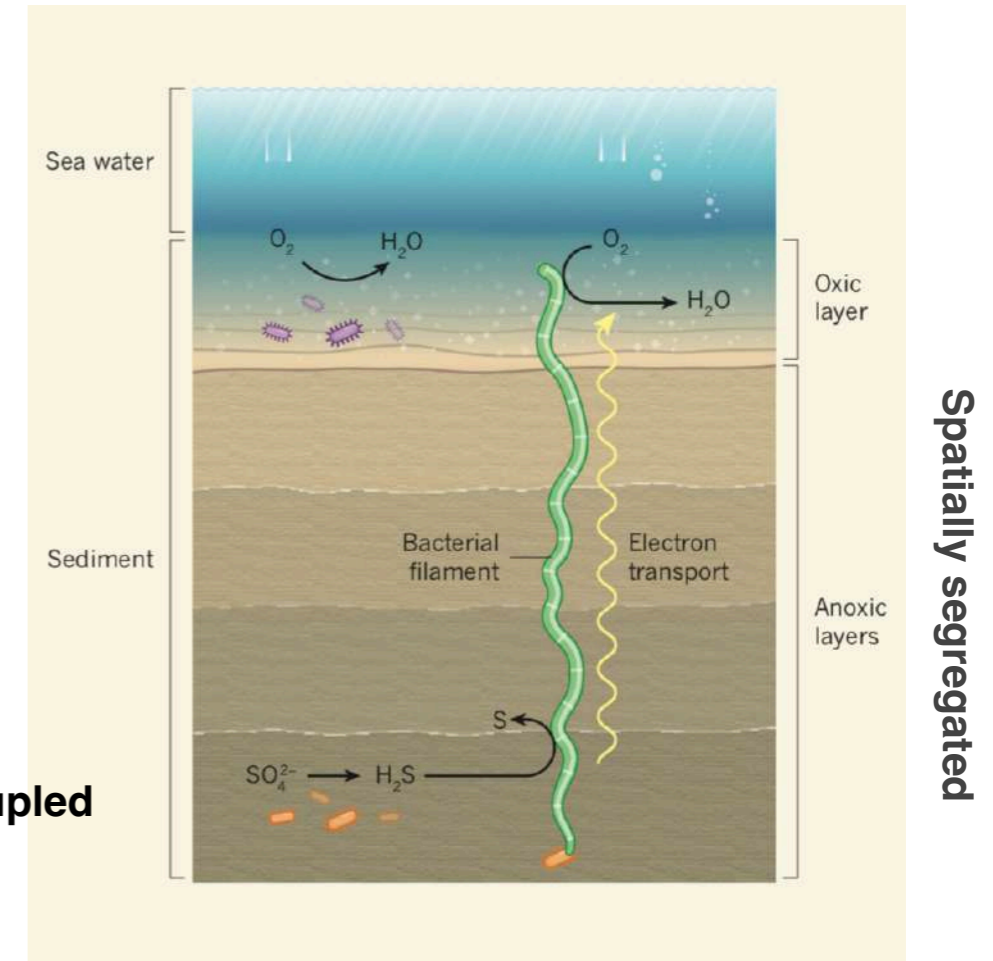


Sharing the burden in the sediment

Pfeffer et al., 2012



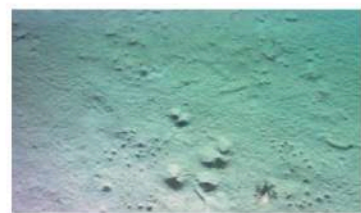
Reguera, 2012



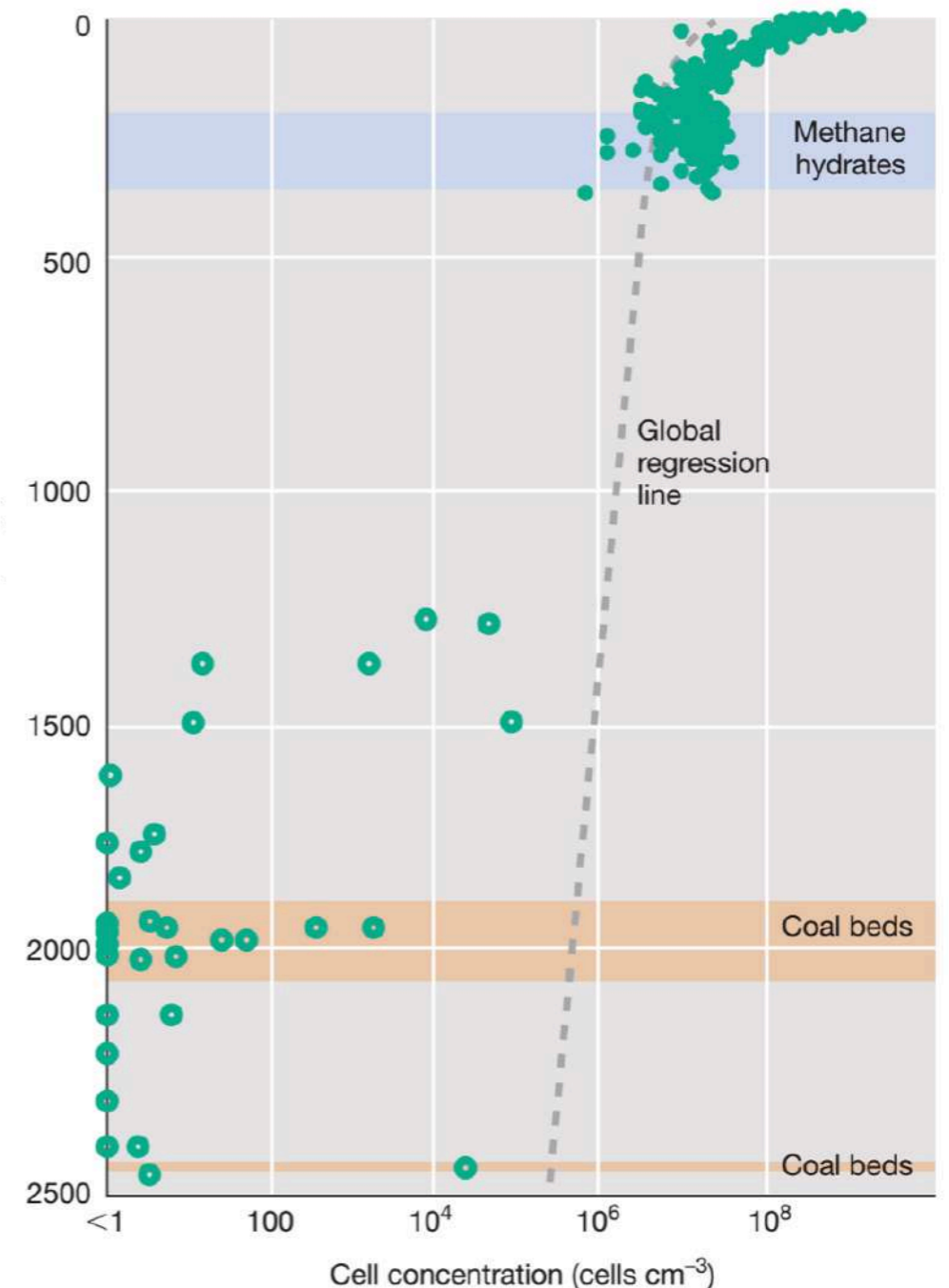
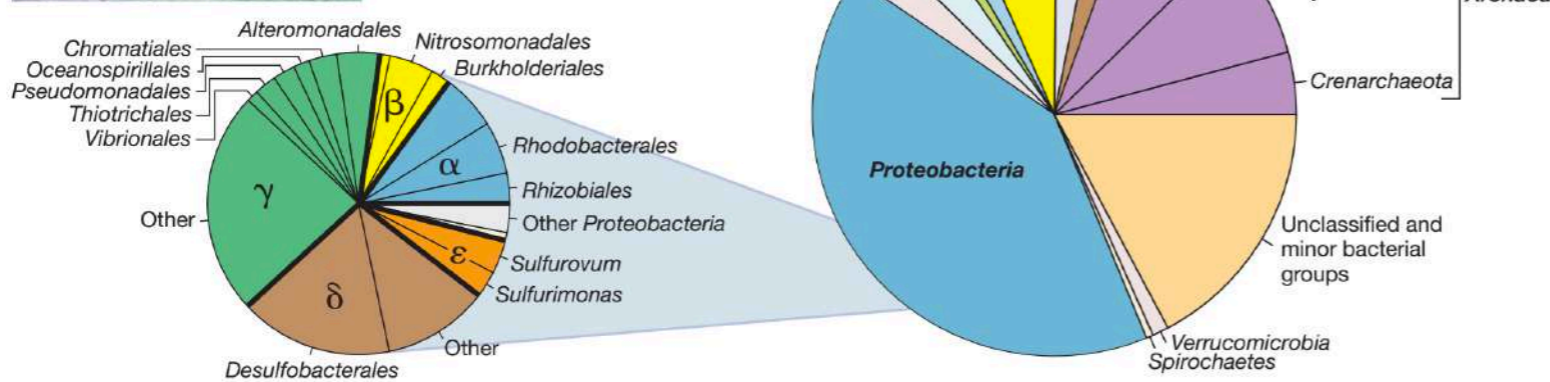
- Microorganisms (purple) in the upper layers of marine sediments use oxygen (O₂) that diffuses from sea water as an acceptor of electrons, which they produce in energy-generating metabolic reactions. **As a result, other microbes (orange) in deeper, anoxic layers (where oxygen is scarce or absent) have to use other electron acceptors such as sulphate (SO₄²⁻) for growth.** Transfer of electrons to oxygen results in the formation of water, whereas electron transfer to sulphate produces hydrogen sulphide (H₂S), which is poisonous to many organisms
- Long bacterial filaments could transport electrons generated when hydrogen **sulphide is converted into sulphur (S)** at the bottom of the sediments and use them to consume oxygen in the upper layers
- During classical sulfur (S) oxidation, bacteria consume both O₂ and H₂S, and, as a result, both half-reactions of the overall redox reaction occur in the same location
- During electrogenic sulfur oxidation, **long-distance electron transport (LDET)** allows multicellular cable bacteria to harvest H₂S in deeper horizons of the seafloor, while still keeping access to O₂, which is available only in the top millimetres of the sediment. **By separating the two half-reactions in space, cable bacteria can outcompete classical sulfide oxidizing bacteria**

Microbial abundance and diversity in the sediment

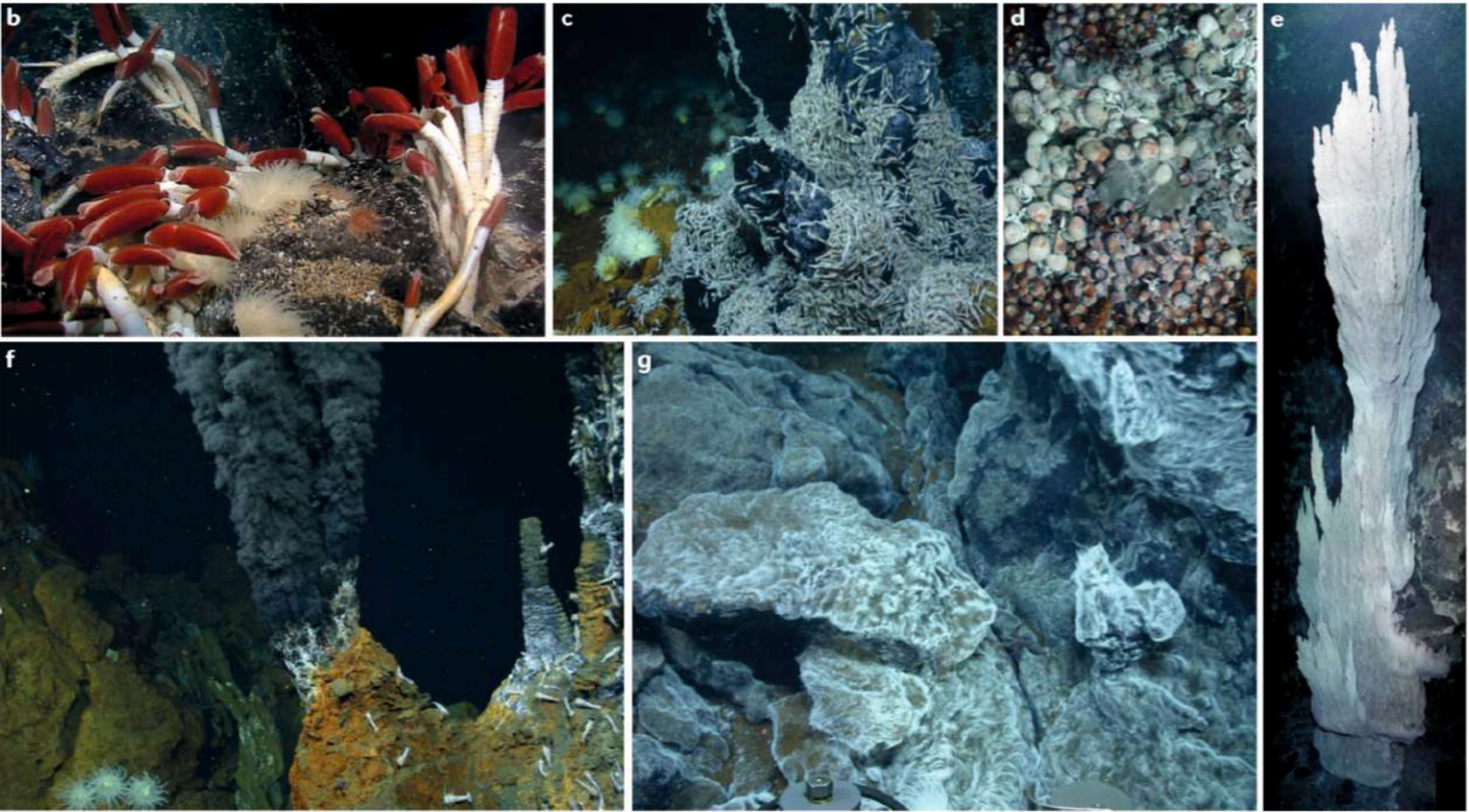
- Structured 3D environment at high pressure at low temperature
- Hydrothermal vents, cold seeps, brines, carcasses as oasis of metabolic diversity (e.g. symbiosis, temperature, relatively fresh organic matter)



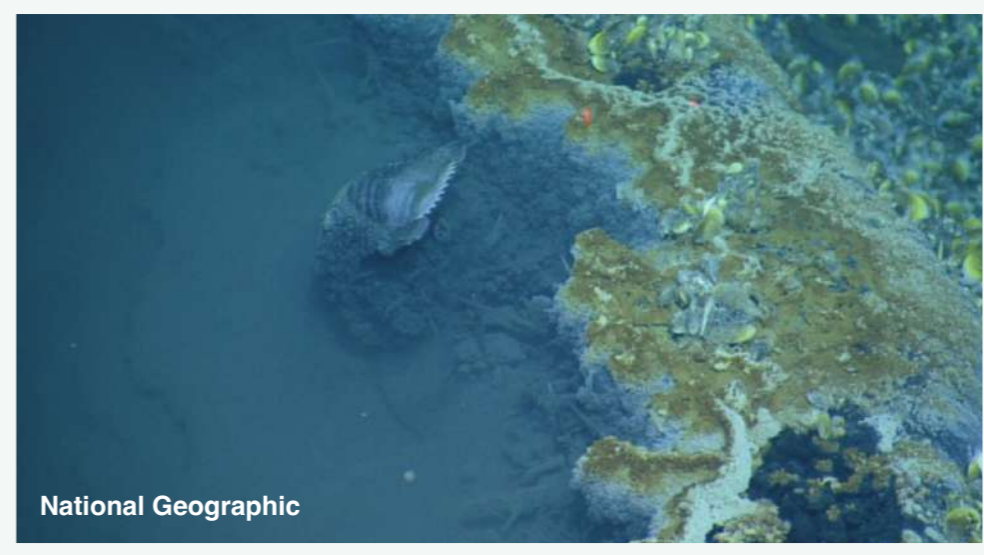
1. Extract sediment microbial community DNA
2. Isolate, sequence, and analyze 16S rRNA genes



- 16SrRNA gene based diversity



Oasis in the deep ocean



[STAGES OF A WHALE-FALL COMMUNITY]

SCAVENGER STAGE

Hagfish—primordial relatives of vertebrates that are virtually blind and live on the muddy seafloor—eat much of the blubber and muscle tissue, helped by other scavengers, including sleeper sharks and some crabs.

DURATION: UP TO 2 YEARS

OPPORTUNIST STAGE

Animals feed on leftover scraps of meat and blubber and on whale oil that has soaked the surrounding sediment. This second wave of scavengers includes snails, bristle worms and hooded shrimp. Meanwhile “zombie worms” (see illustration on page 84) begin to spread their roots into the bones and feed on their lipid content.

DURATION: UP TO 2 YEARS

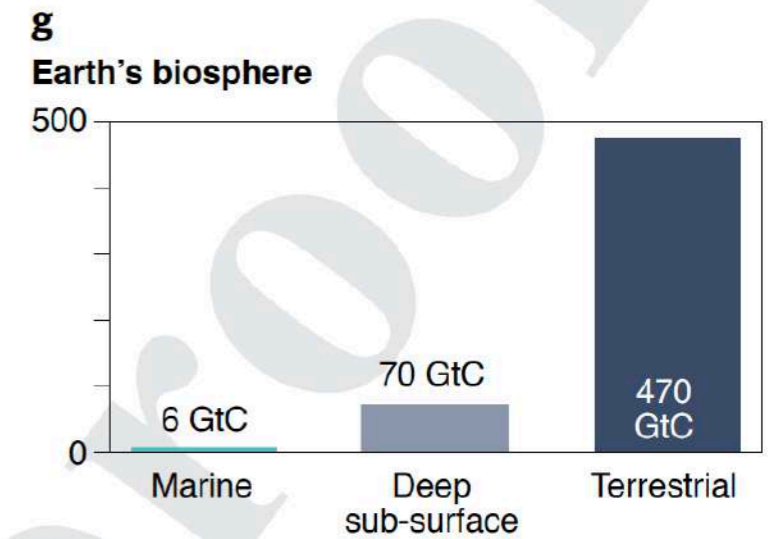
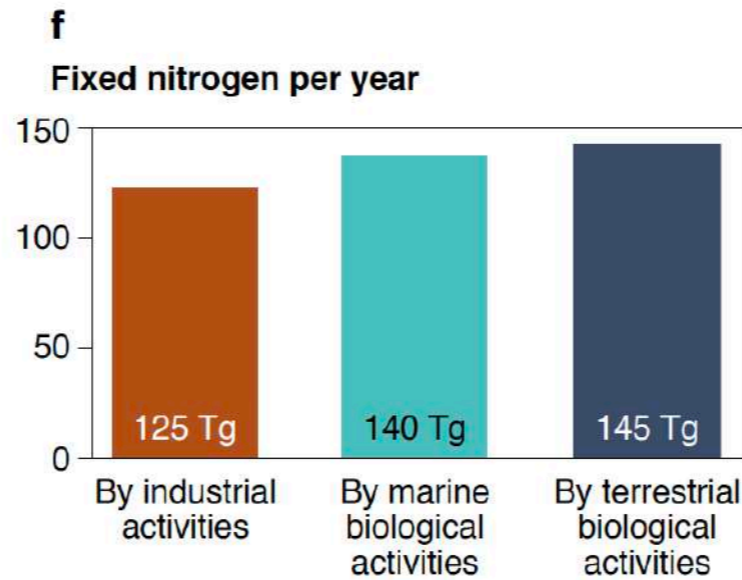
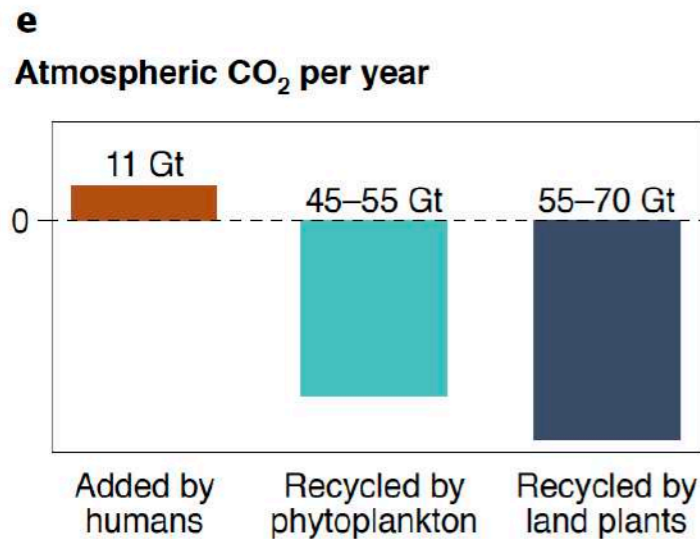
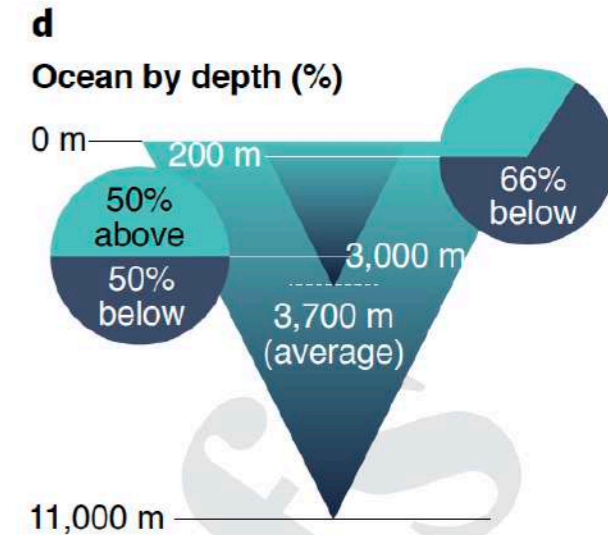
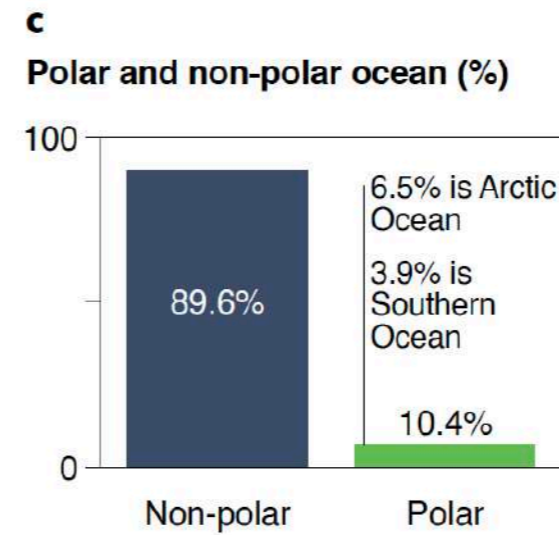
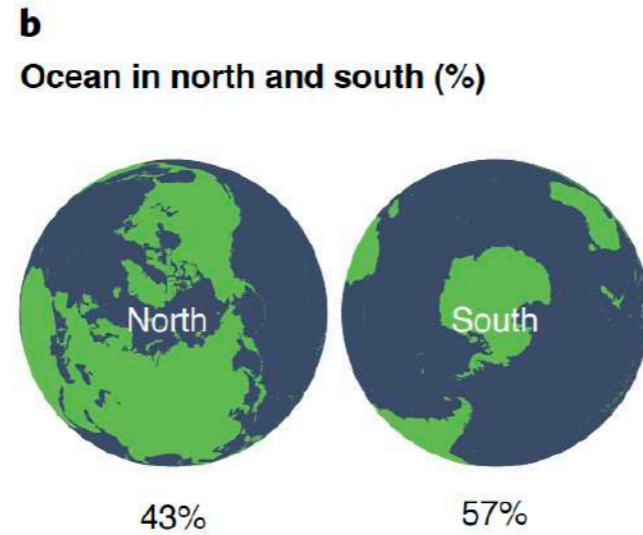
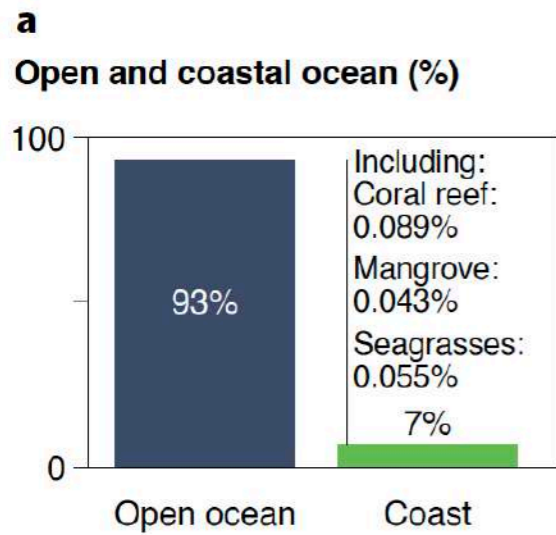
SULFOPHILIC STAGE

Anaerobic bacteria produce hydrogen sulfide, which other, “sulfophilic,” bacteria use for energy. The sulfophilic bacteria, in turn, support all other organisms (inset at bottom). Mussels, tube worms and clams derive energy from sulfophilic bacteria that live symbiotically within them. Bristle worms and limpets feed on mats of such microbes. Crustaceans such as squat lobsters prey on other animals.

DURATION: UP TO 50 YEARS

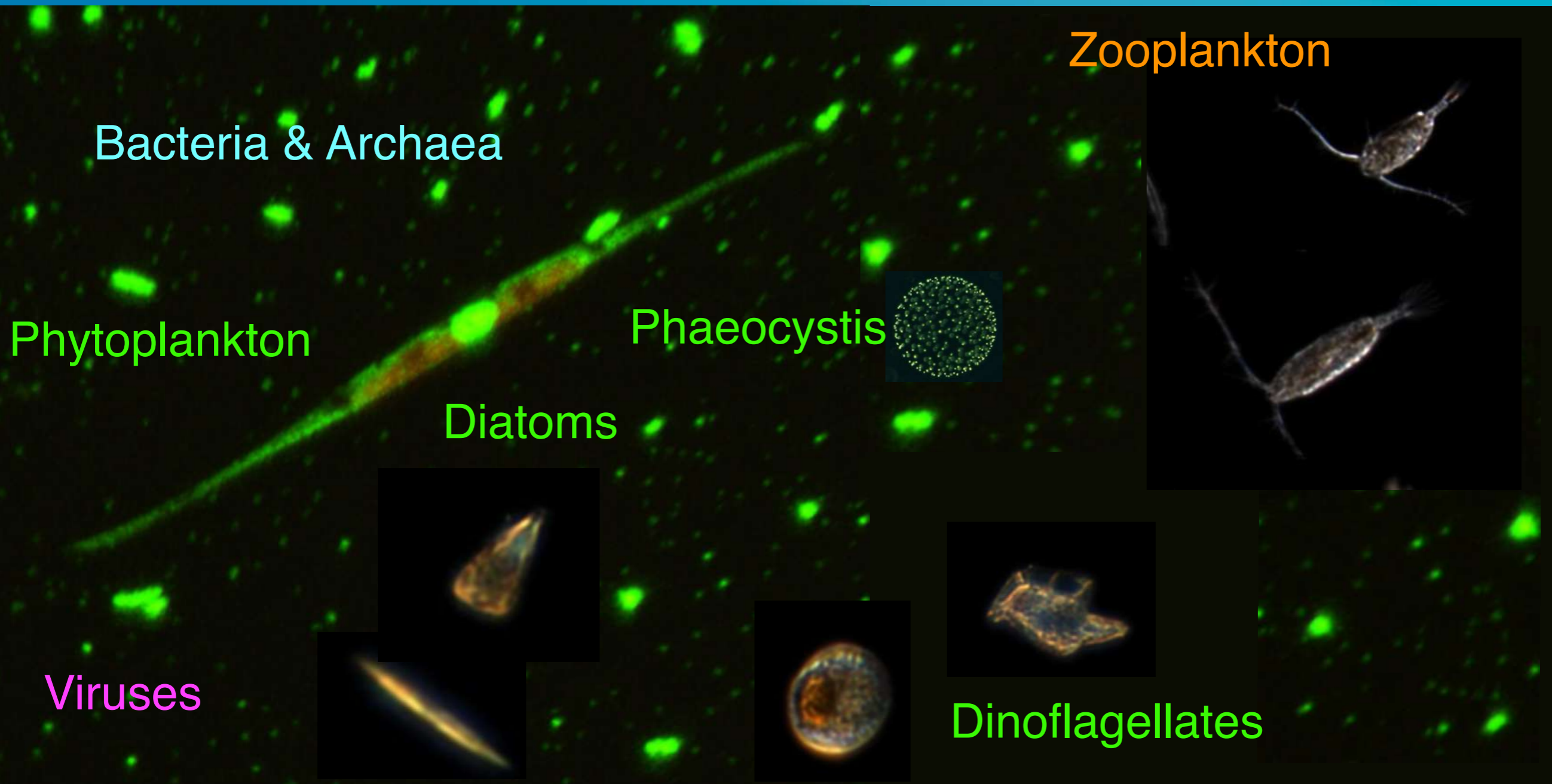
**SEAWATER:
COASTAL
And
OPEN
OCEAN**

Key features related to the ocean microbiome

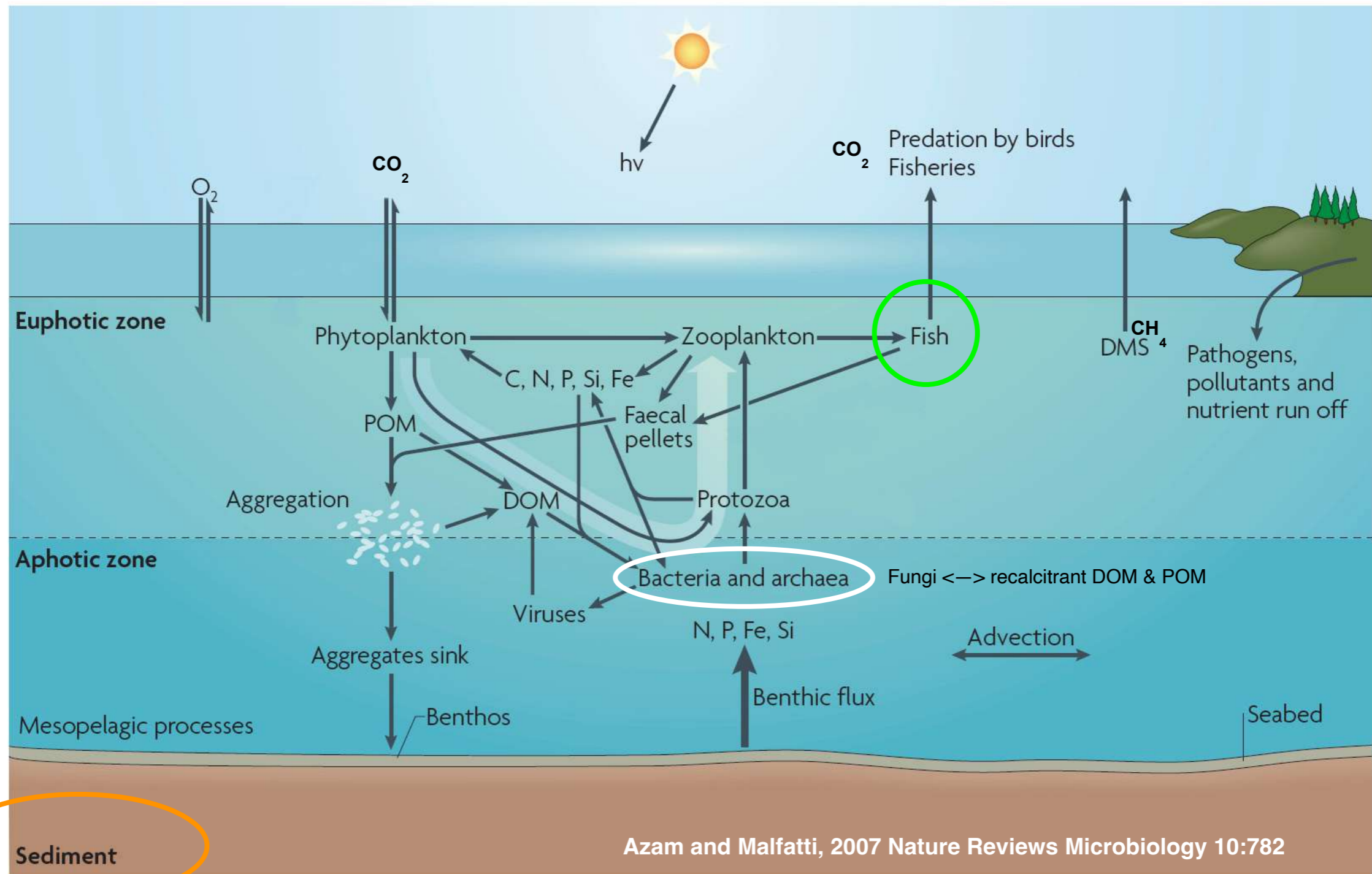


Upper ocean water column

SeaSCAPES microbial dynamics

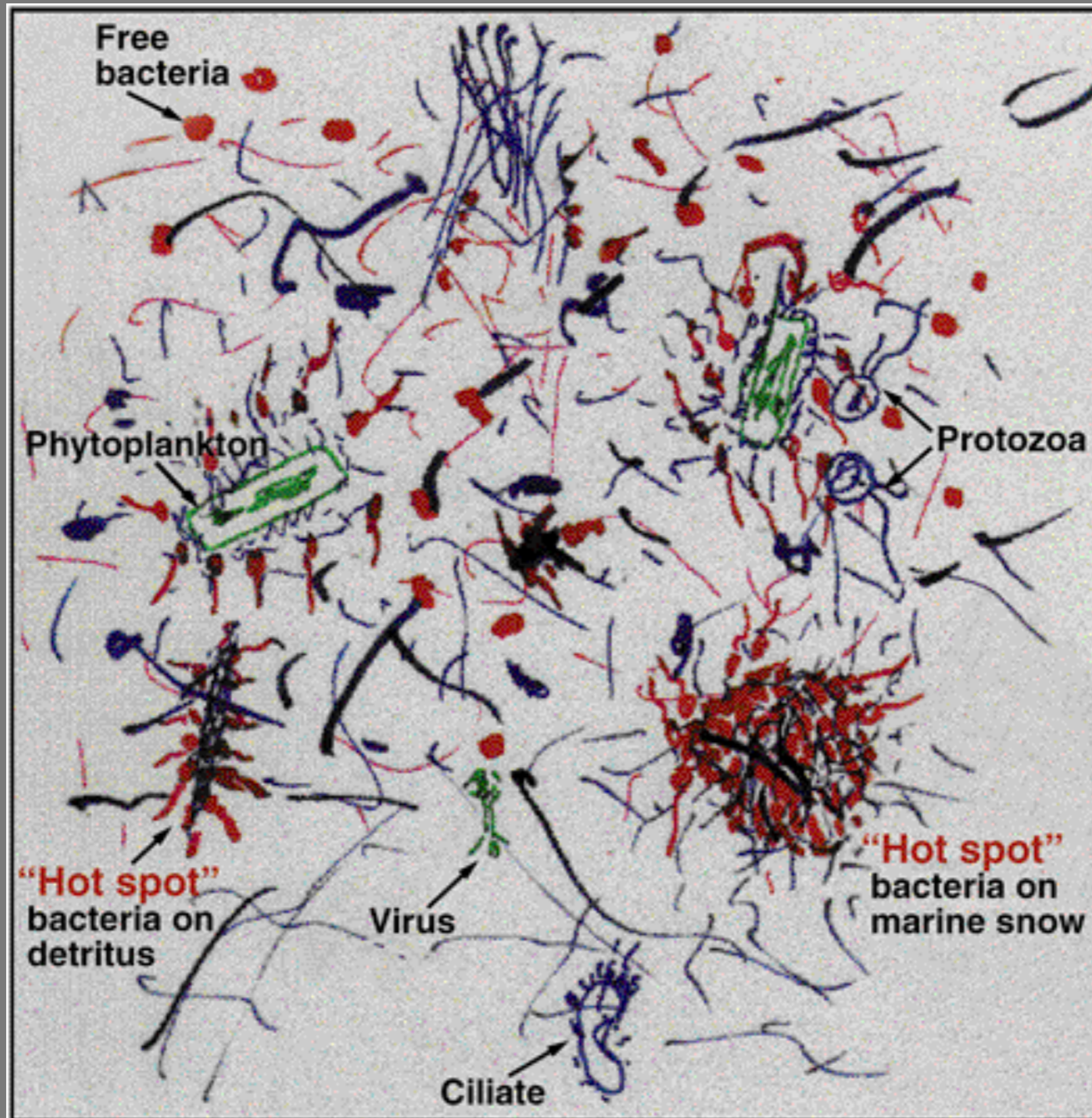


Fate of fixed carbon in ocean ecosystem

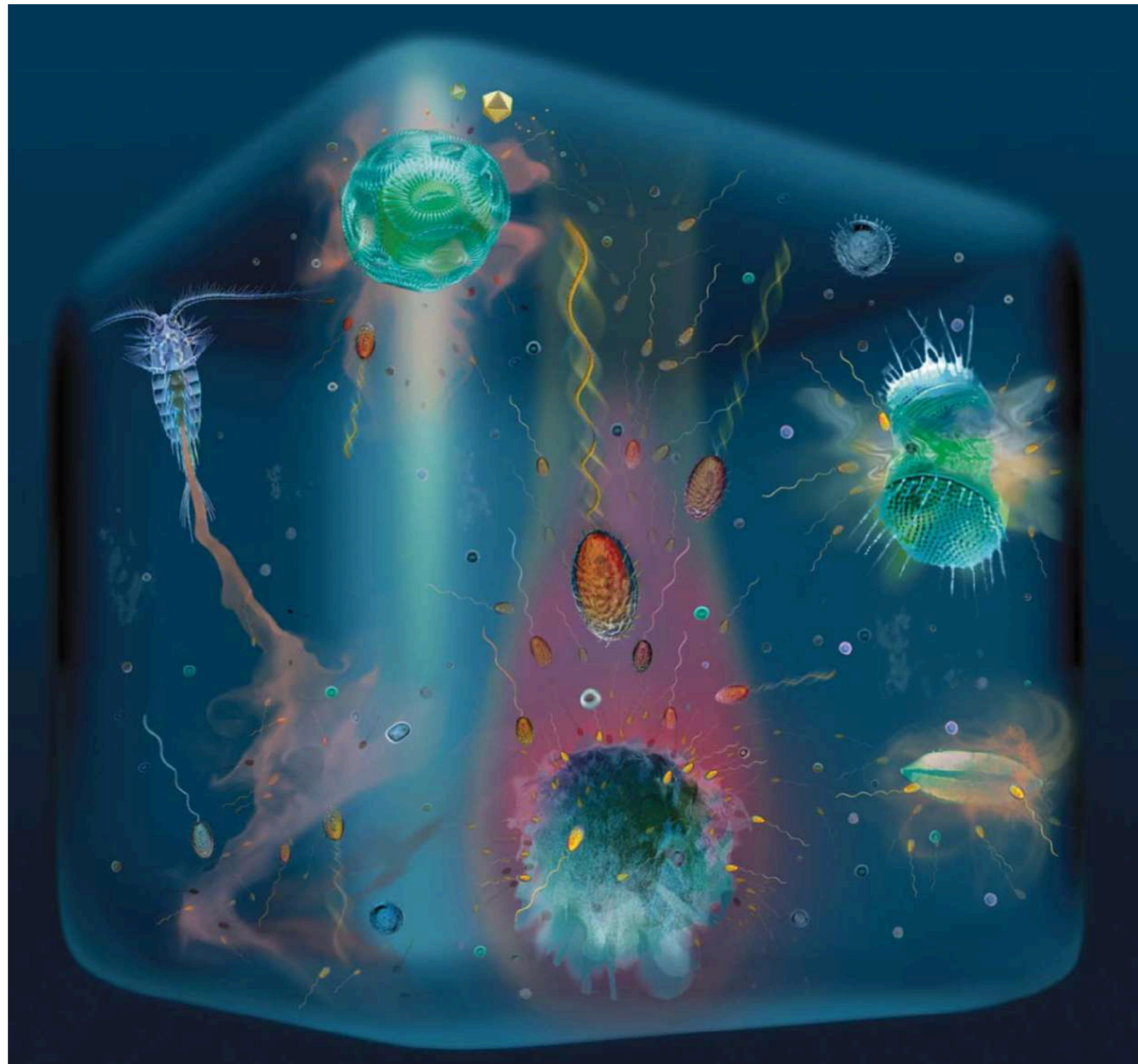


Seasonality → high and low production

Marine microbial environment: impressionistic painting



Marine microbial microenvironment

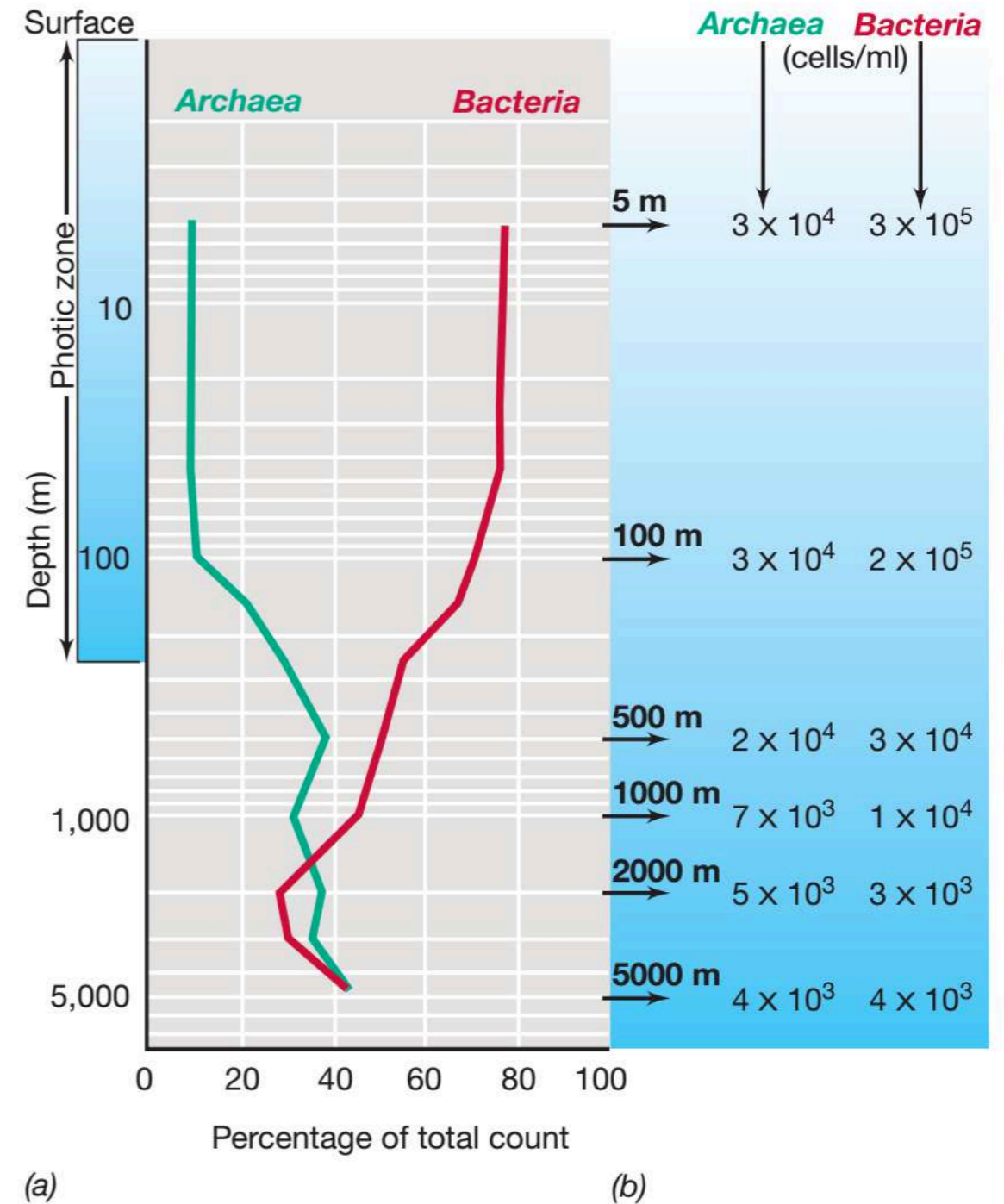
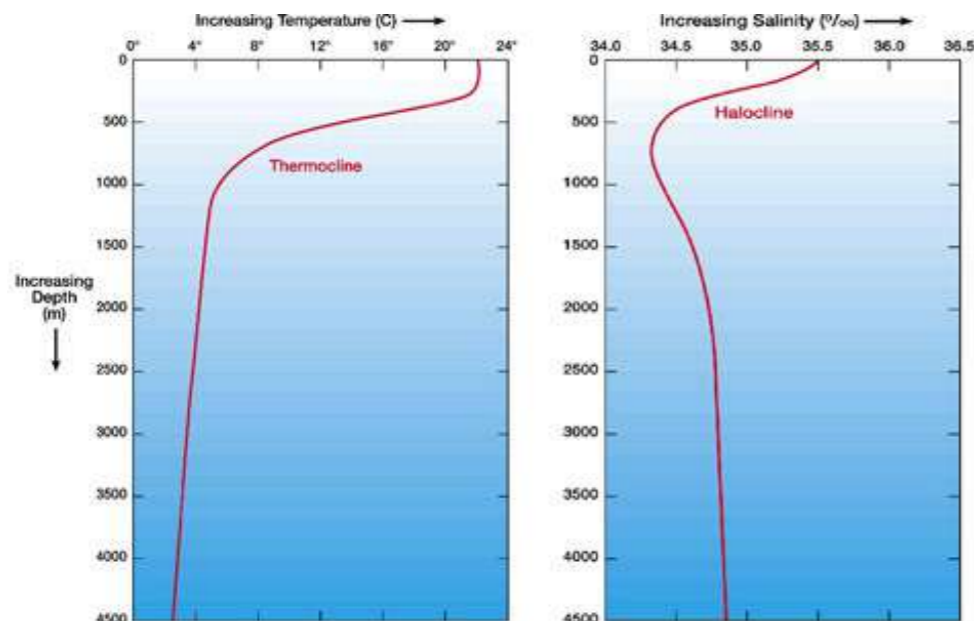


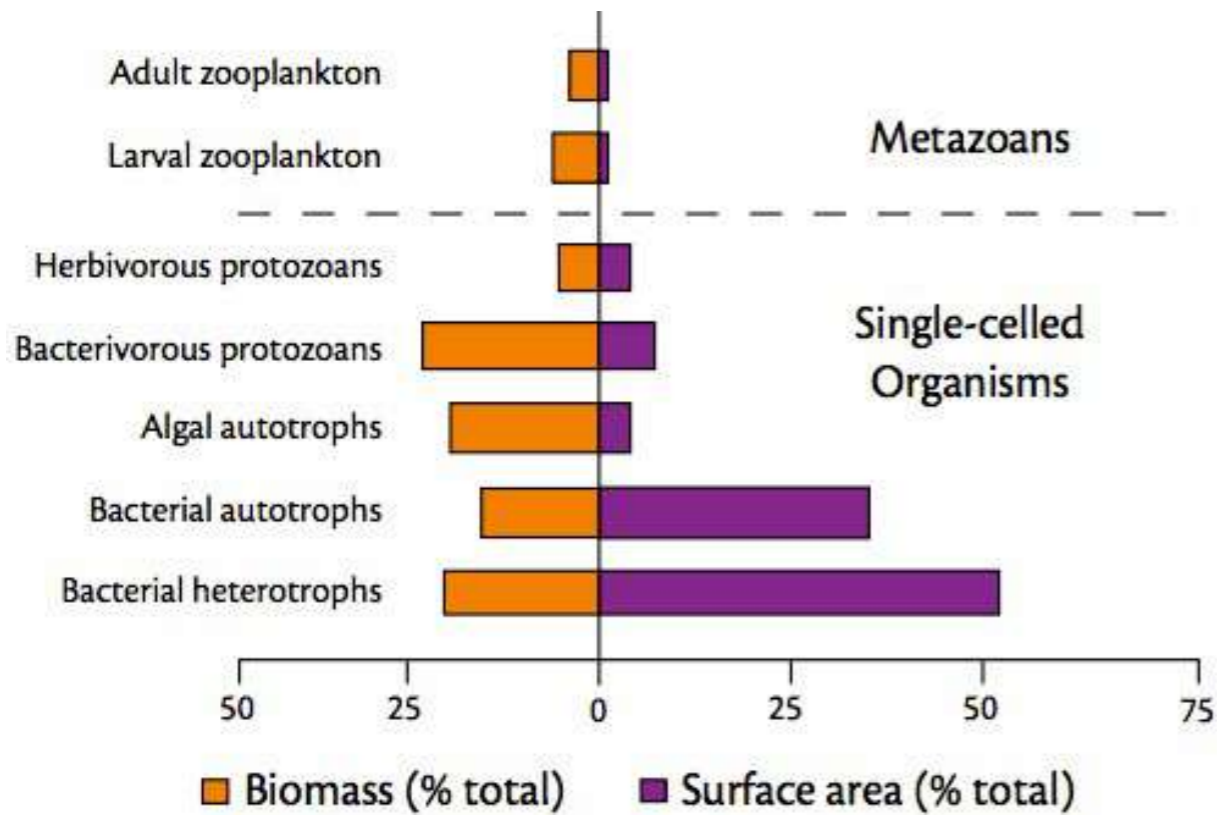
Stocker, 2012

- Disparate processes contribute to make the ocean a sea of gradients, from the vantage point of microorganisms

Microbial abundance in ocean

- Photic zone 0-200 m
- More than 96% of ocean is dark and with a constant temperature ($\sim 4^{\circ}\text{C}$)
- Microbes in coastal ocean 1×10^6 cells/mL
- Archaea dominate at depth
- Primary production sustain ocean interior — > organic matter degradation and sediment too!





Bacteria/Archaea represent 90% the total biomass in the ocean
Bacteria/Archaea are the most productive organisms in the ocean and contribute up to 90% to biota respiration

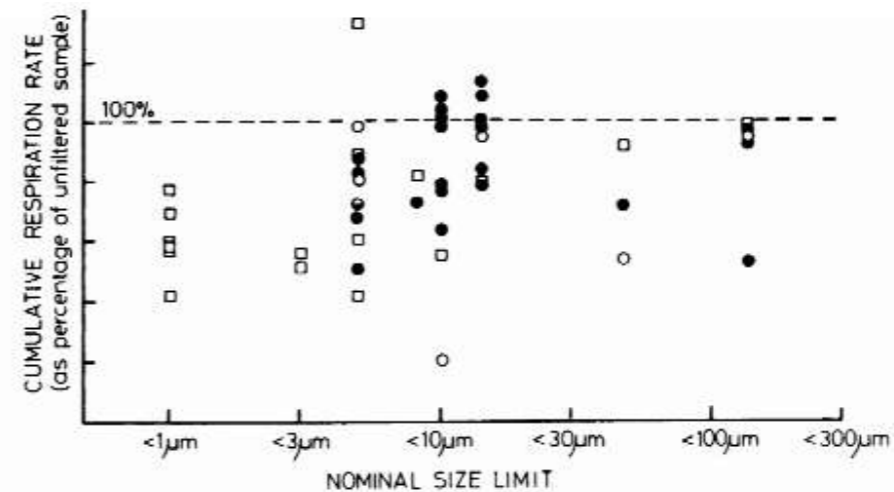
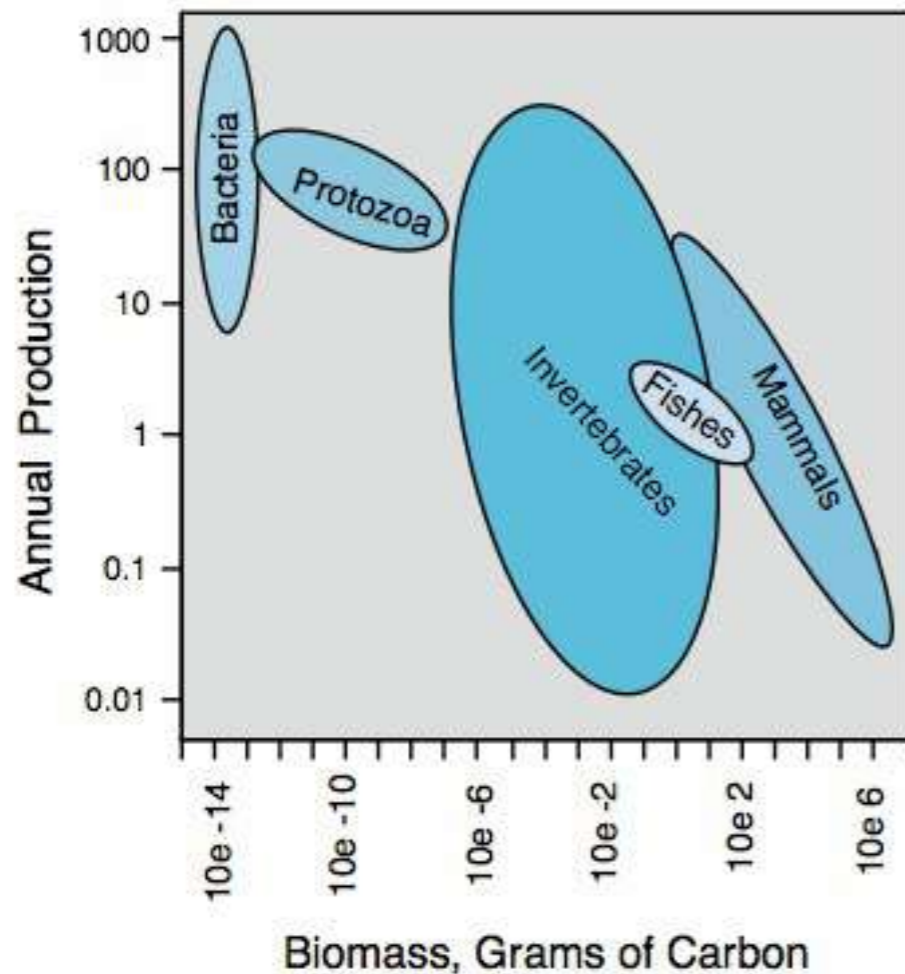
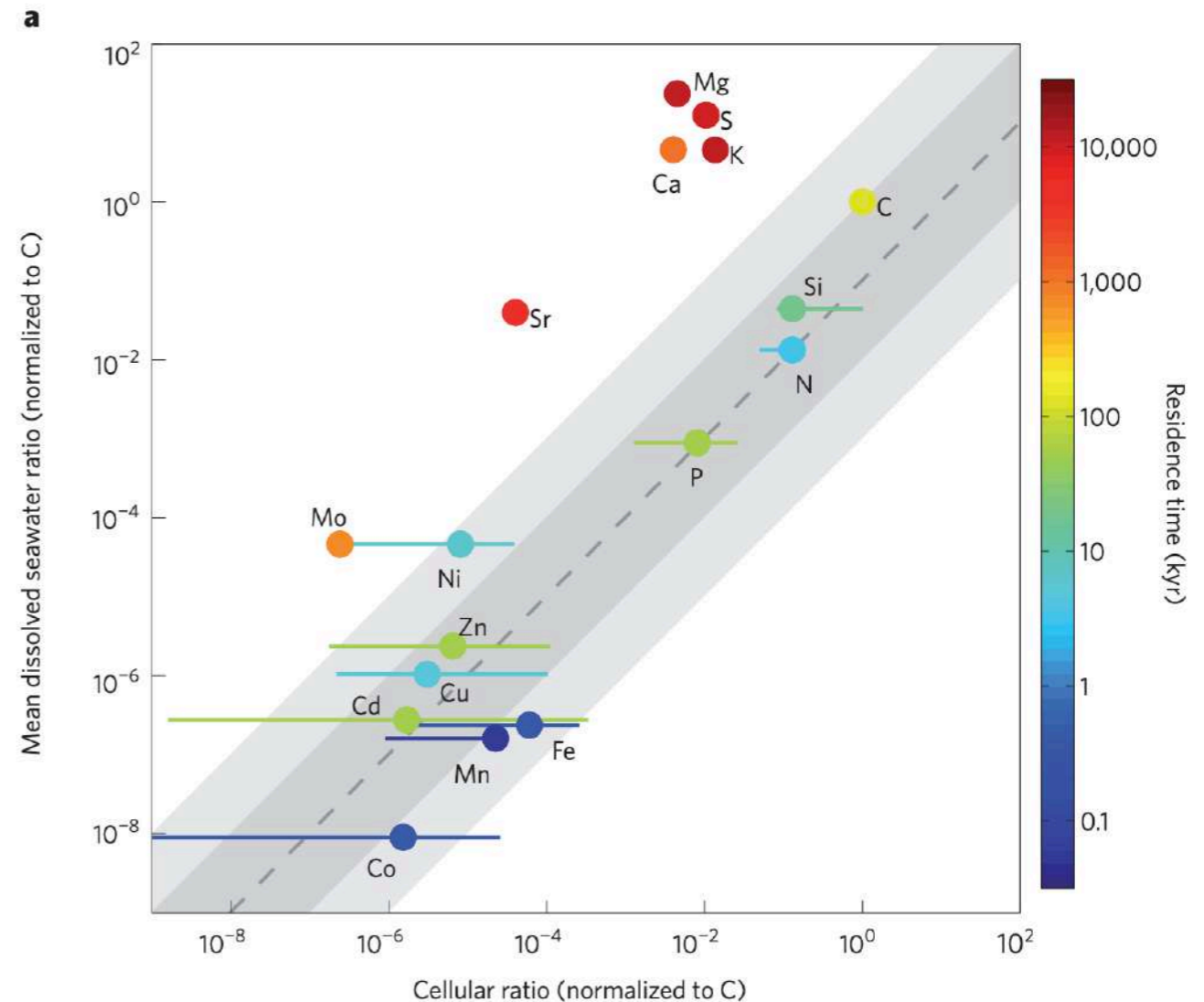
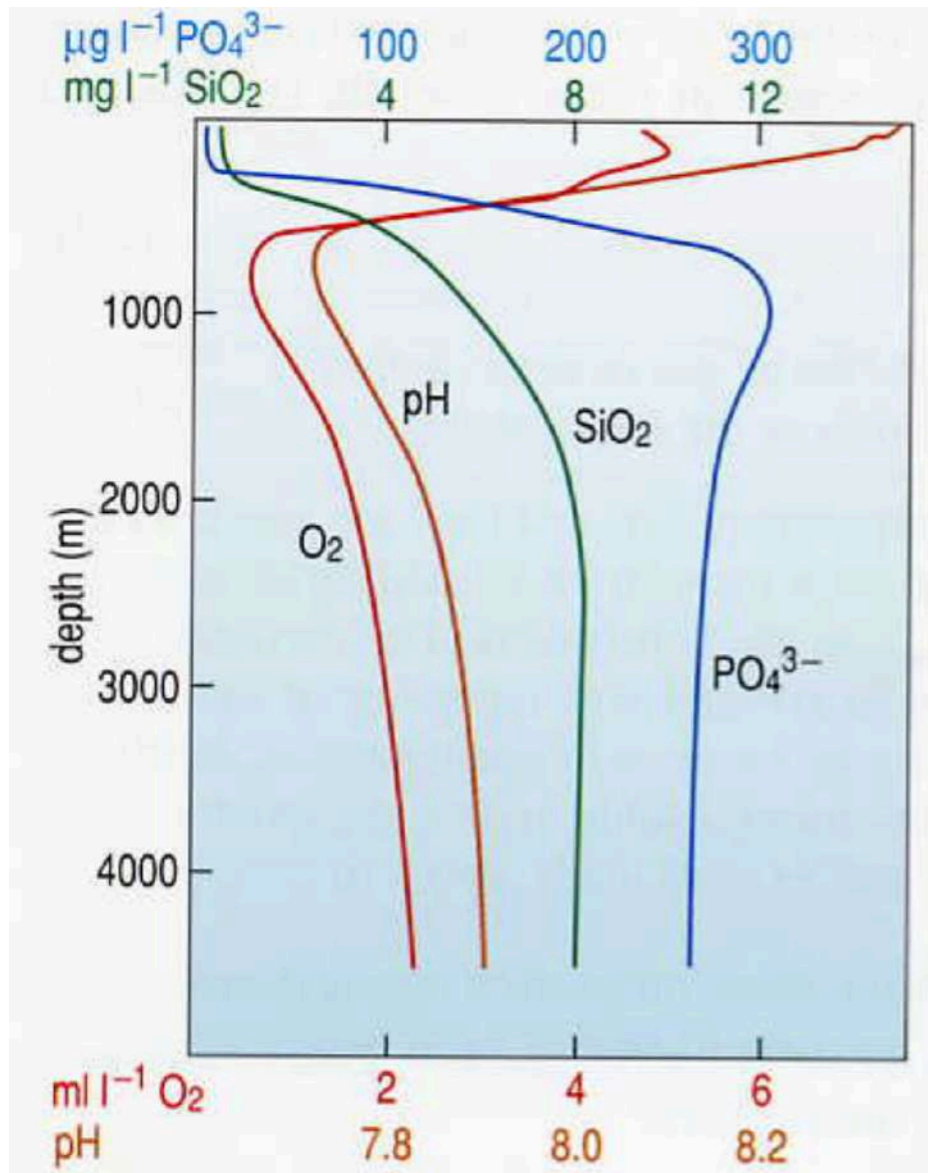


Figure 6. Distribution of respiratory activity with size. (□) CEPEX, samples from bag; (○) Loch Ewe, samples from bag; (●) Loch Ewe samples from outside bag. Data are expressed as cumulative respiration up to various size limits, normalized against the rate in the unfiltered sample. All the data points are for a single size horizon and are not replicates.

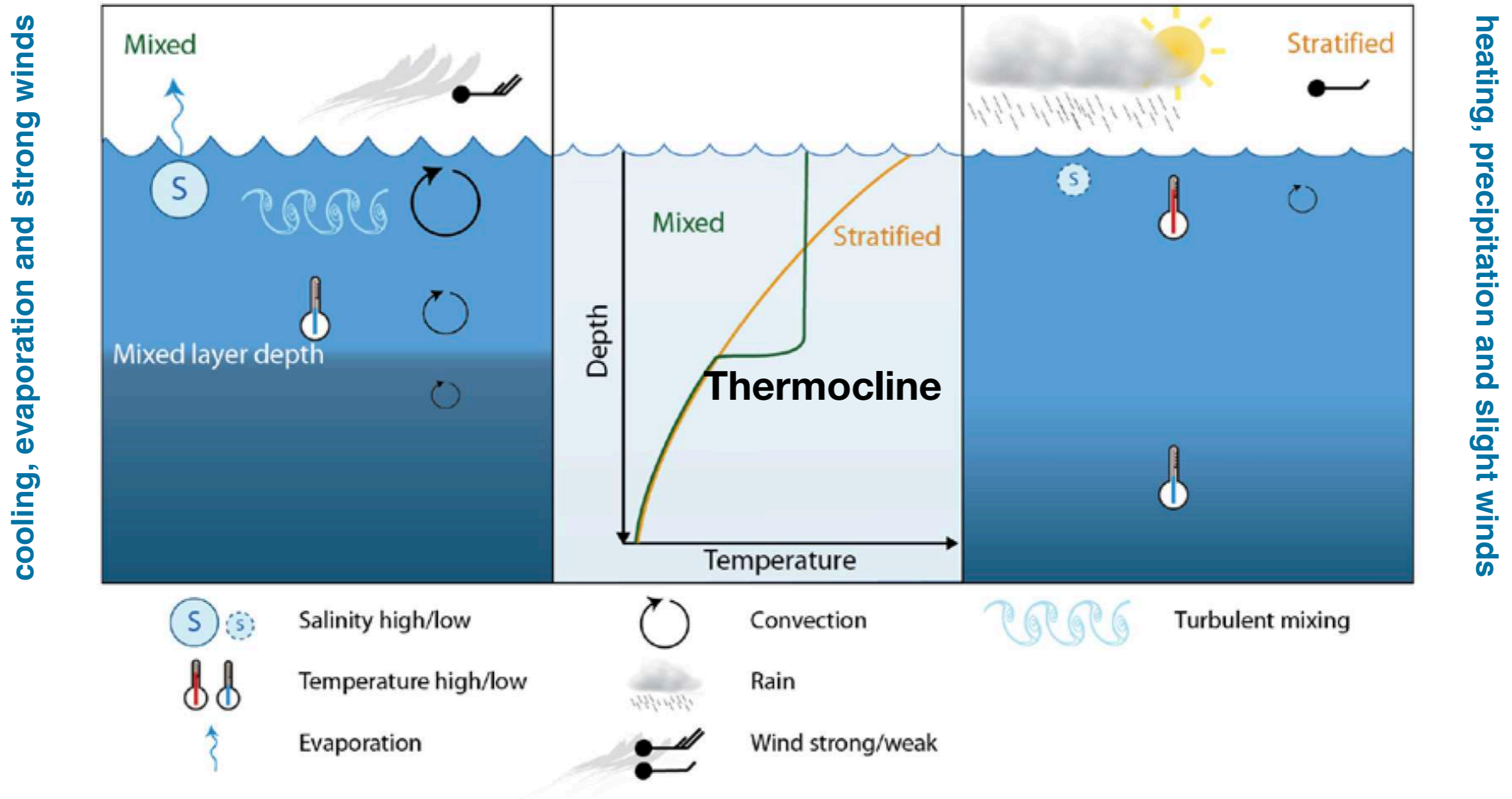
Williams (1984)

Ocean: Organic matter decomposition/respiration —> nutrient recycling

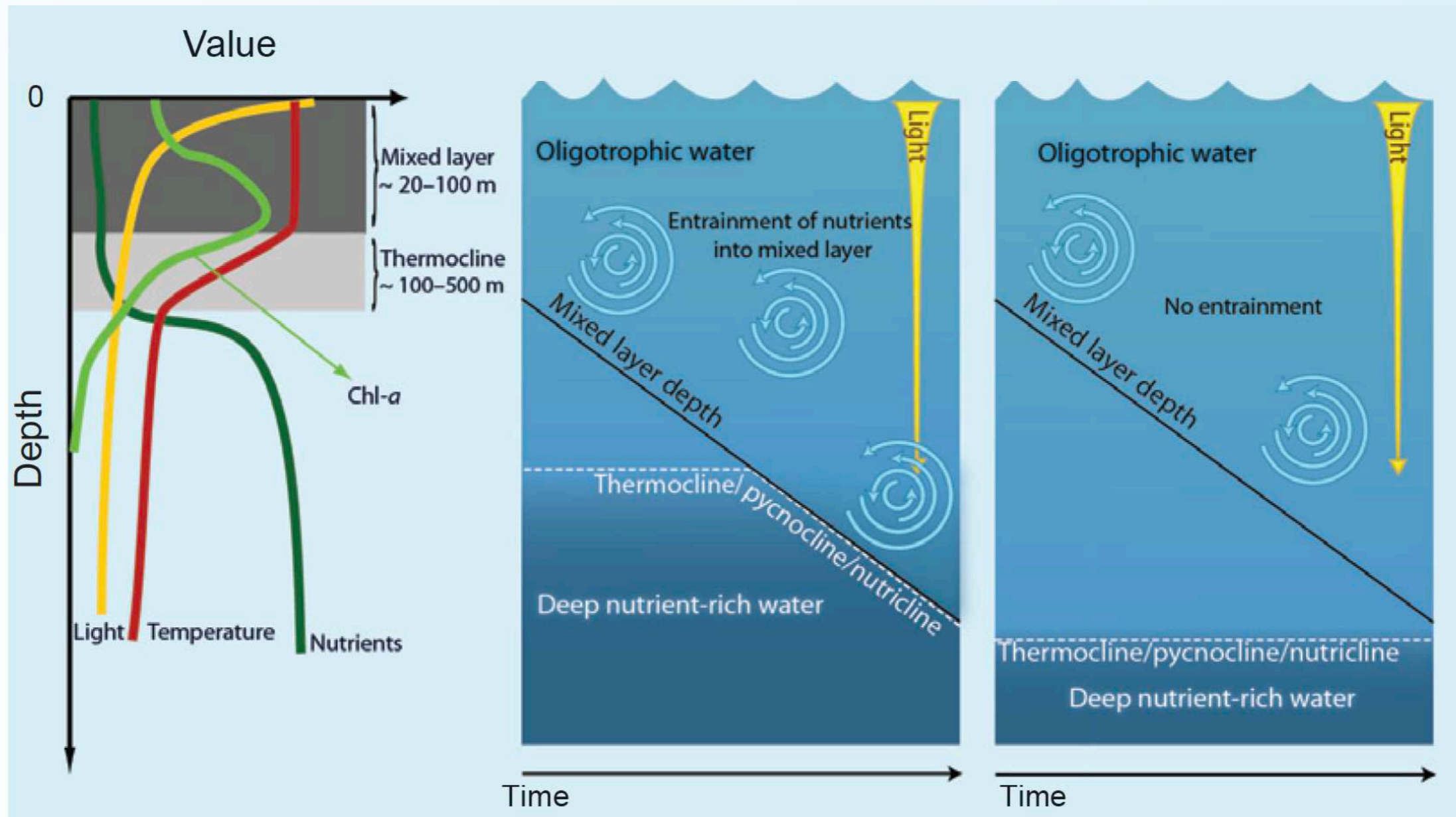
- Microbial degradation of organic matter in the water column generate nutrient fluxes available for primary producers (limiting nutrients based on cell demands)
- Microbial action with organic matter at the microscope structure ocean-basin scale



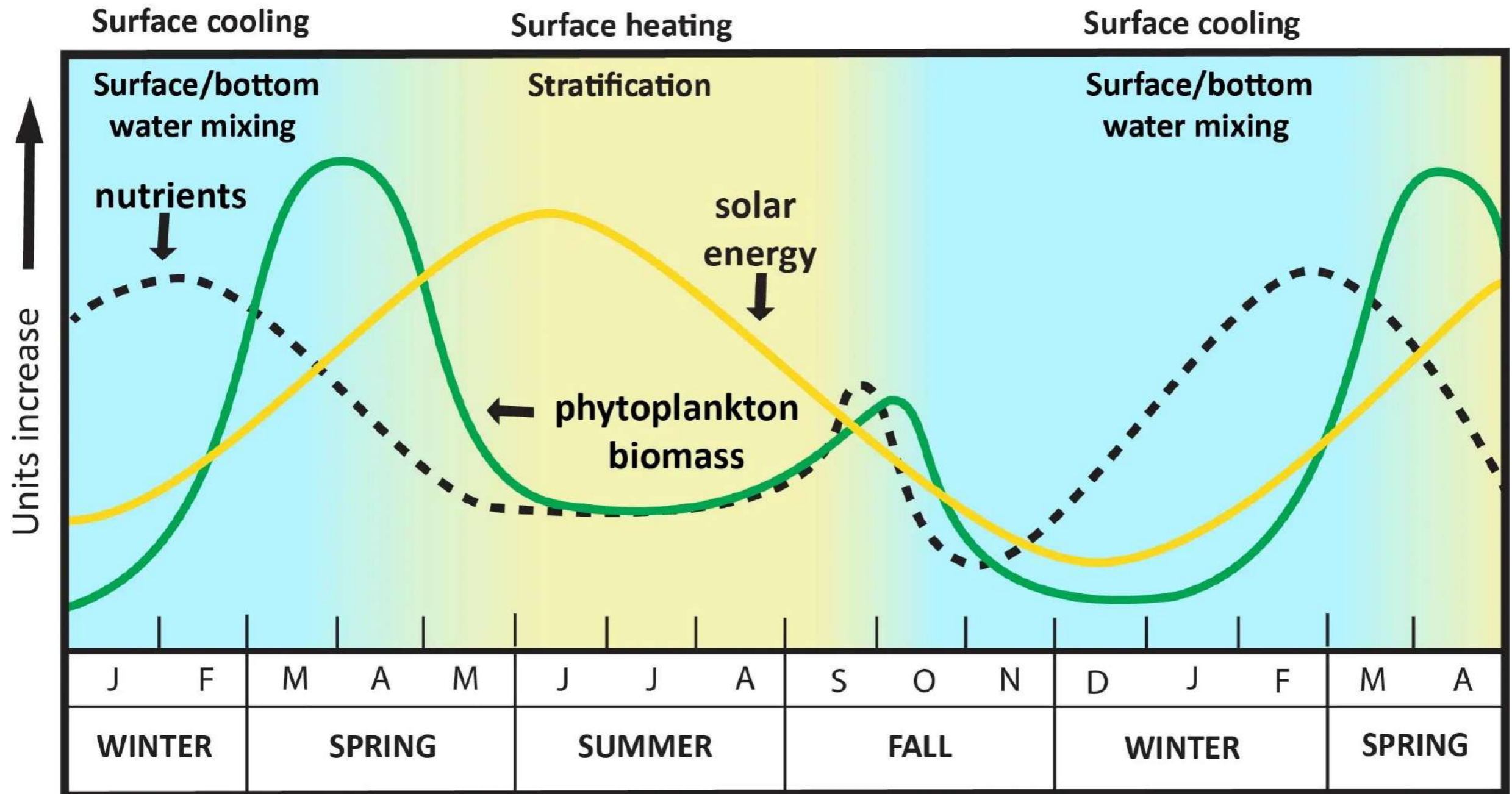
Seasonality in the ocean: factors affecting the mixed layer depth and C fixation in the food web and C biogeochemical cycle



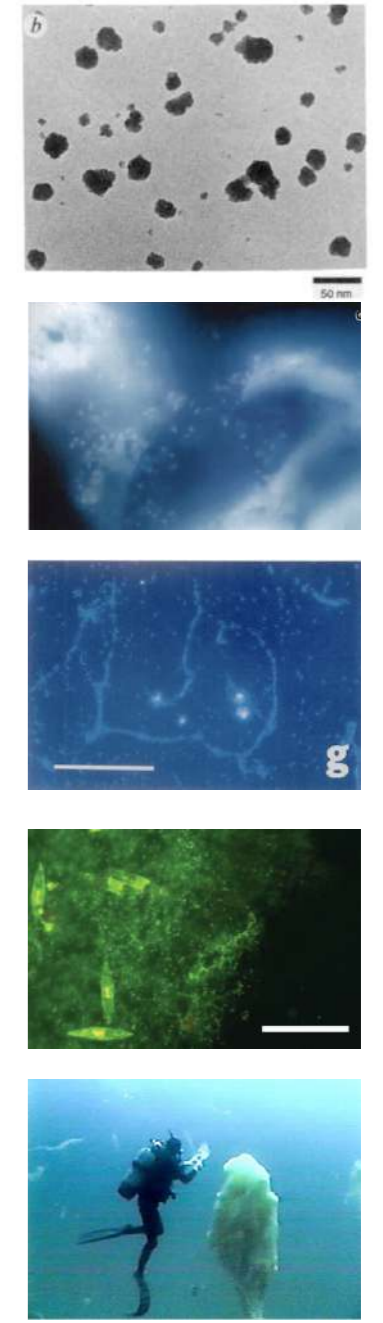
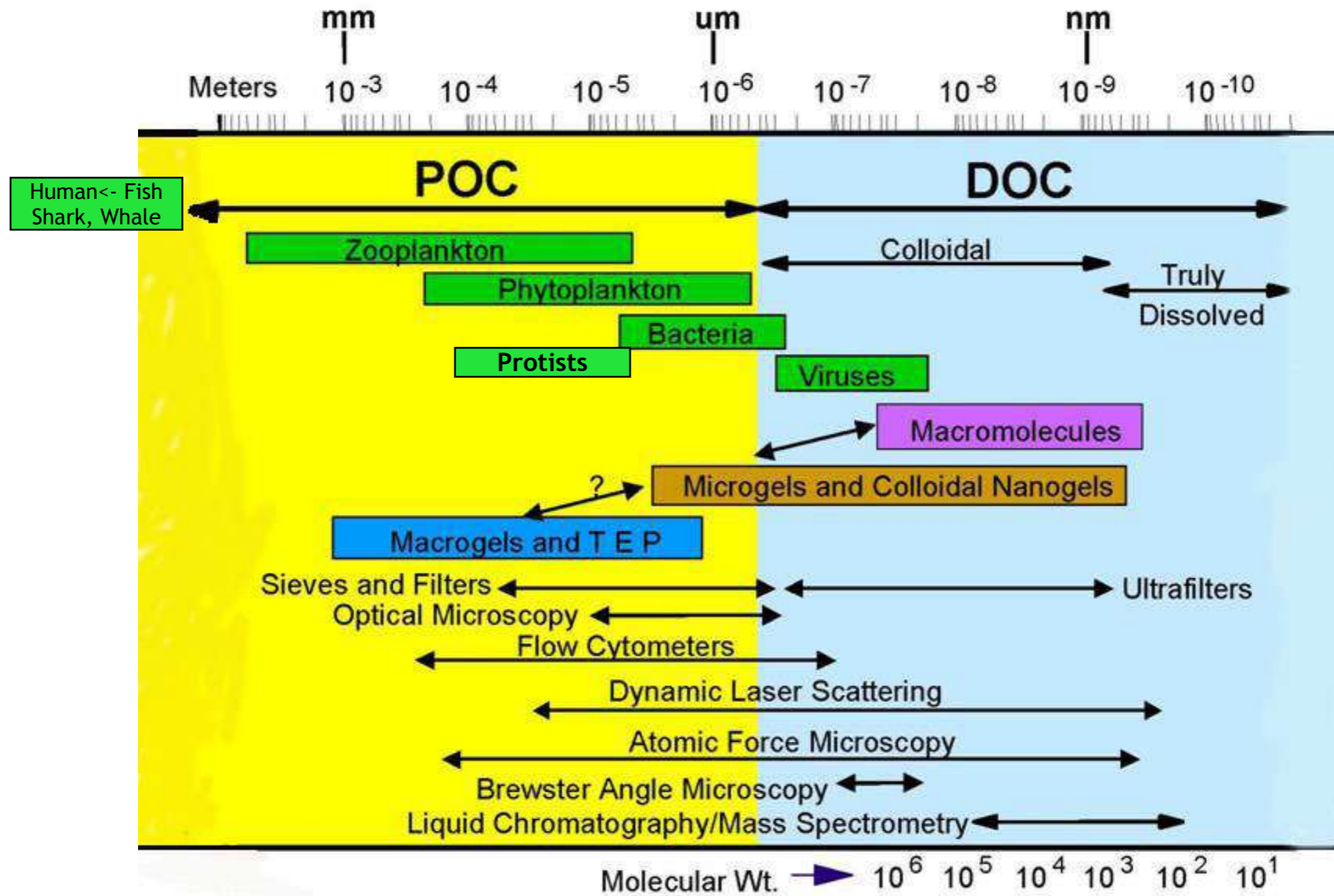
Influence of the thermocline and mixed layer depth on transfer of nutrients to surface waters for C fixation into organic matter by photoautotrophs



Seasonality in C fixation



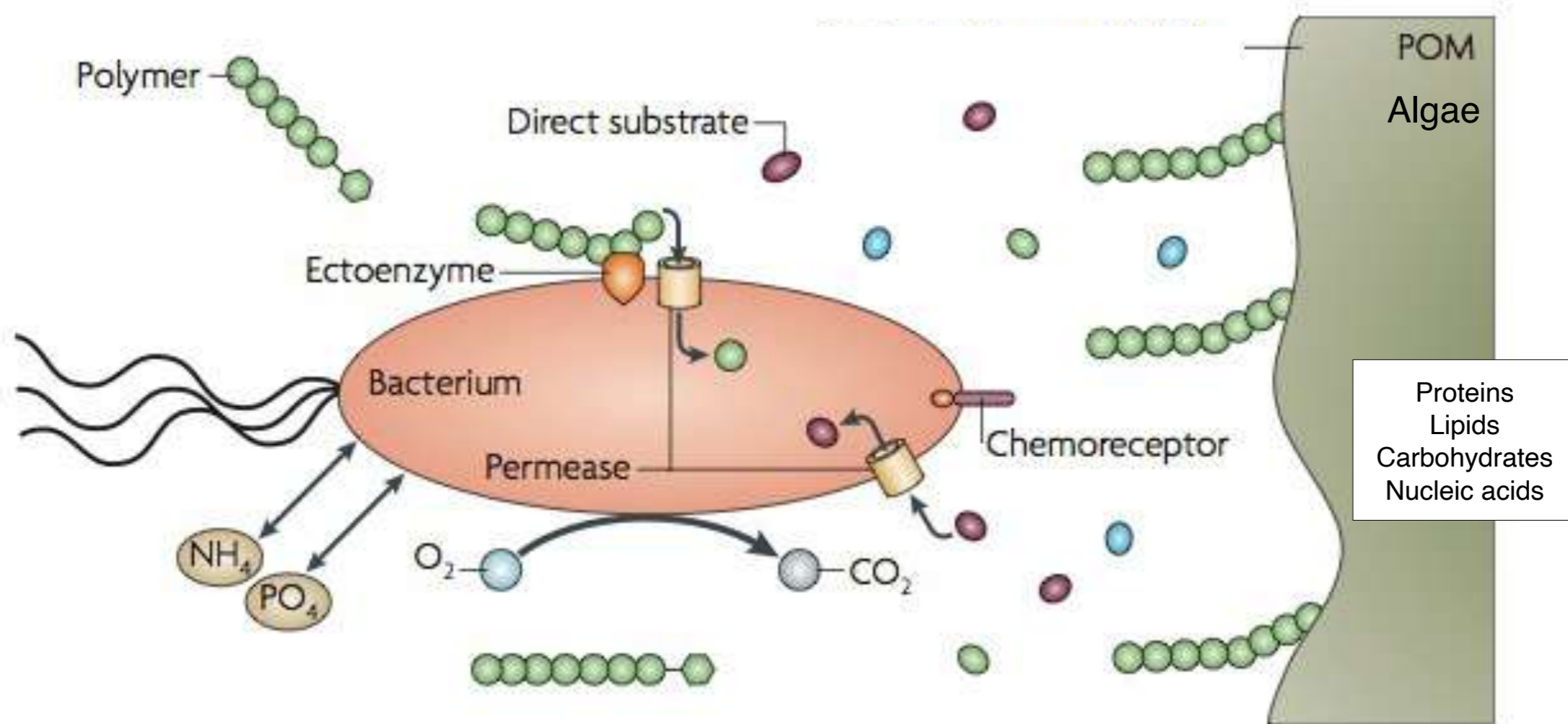
Organic matter continuum: a unifying concept to understand how bacteria perceive and interact with organic matter



By Farooq Azam

- Ocean is replete with transparent gel particles (~ million/liter) creating a gel-like milieu
- Primary production is the source of the great chemical and structural complexity

Microbial adaptive strategies for growth at small scale

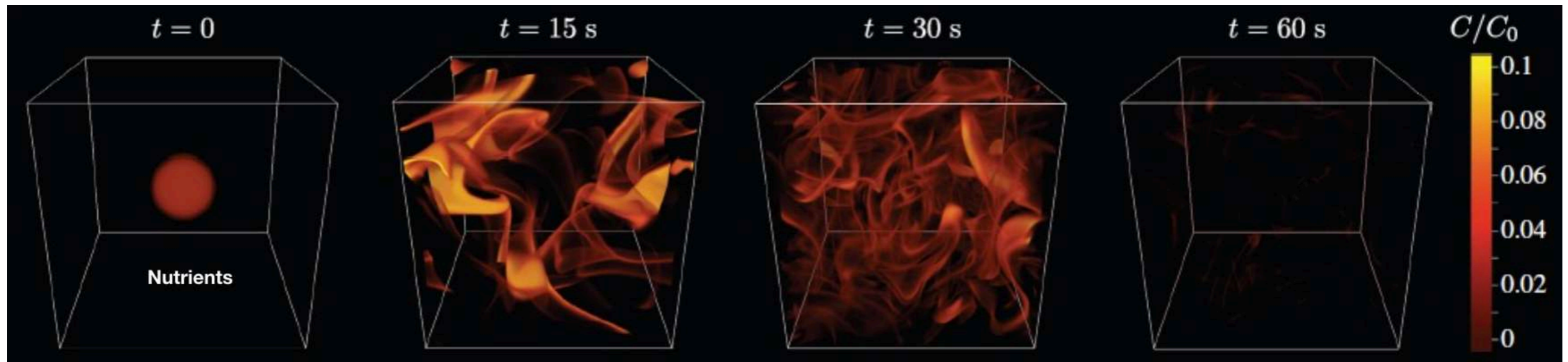


Azam and Malfatti, 2007 Nature Reviews Microbiology 10:782

- **Motility, environmental sensing (interacting with molecules)- MCPs**
- **Significance of spatial coupling hydrolysis-uptake (permease) on the cell**
- **Cell surface hydrolases; 10^2 - 10^4 x variability in cell-specific activity**
- **Coordinated behavior and biochemistry in space and time**

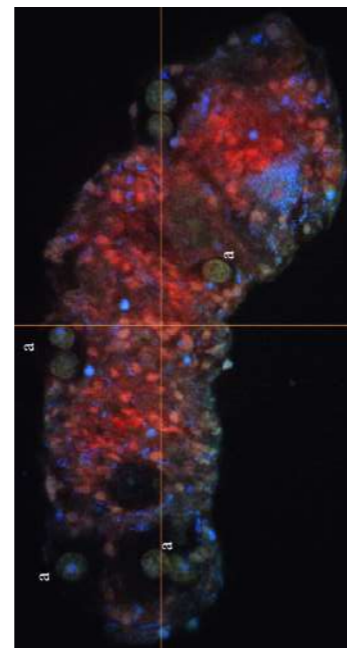
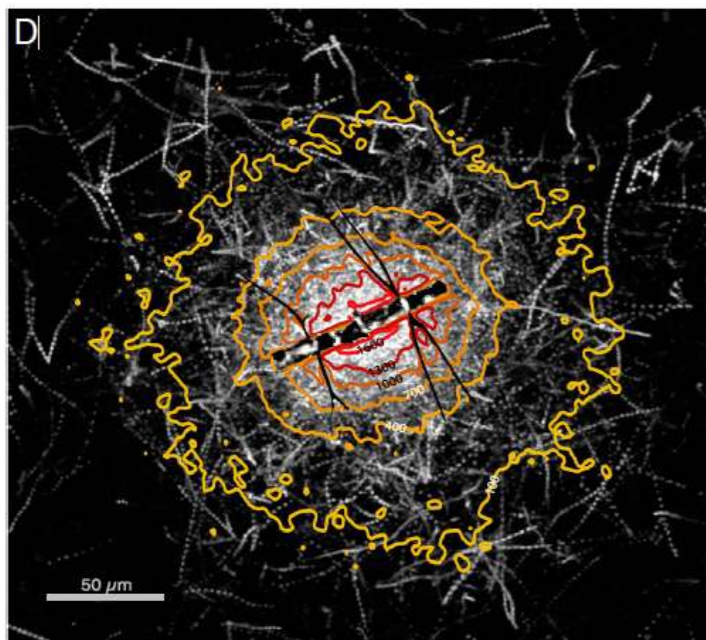
Nutrient patch evolution and sources

Taylor 2012, Science



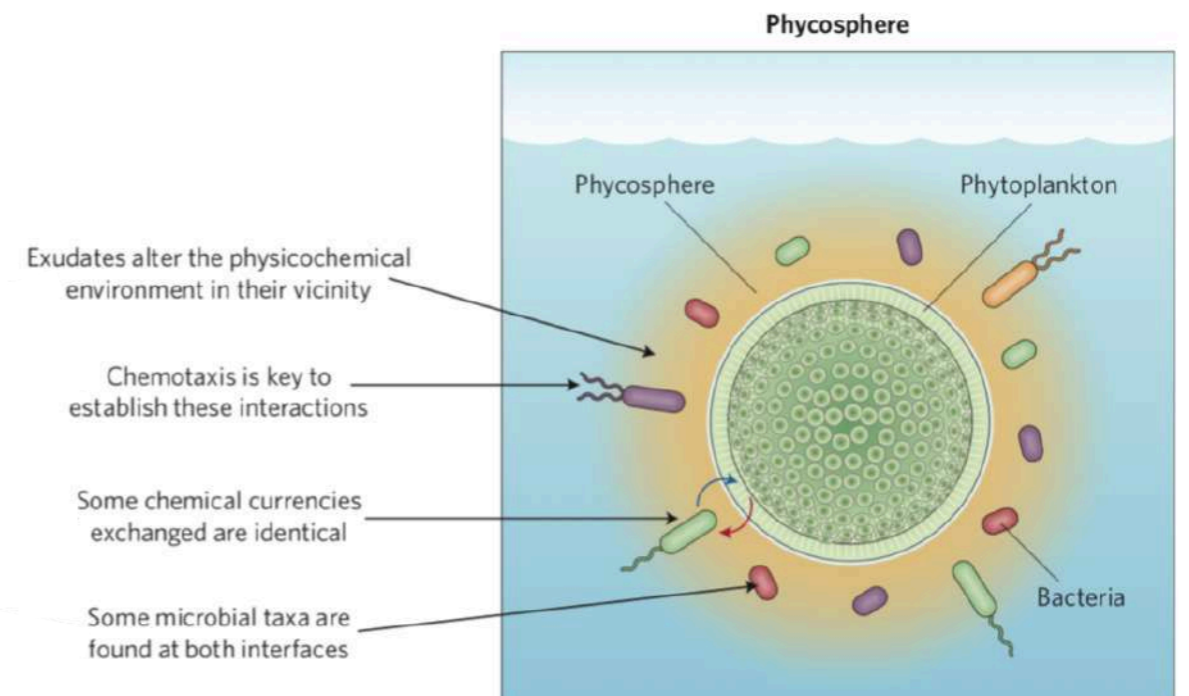
- Low Reynolds number \rightarrow high viscosity
- Modelling in time and space in a turbulent flow ($L = 5.65$ cm)
- Nutrient sources: living phytoplankton, dying phytoplankton, dying copepods, marine snow plume, fecal pellets

Smriga et al. 2016



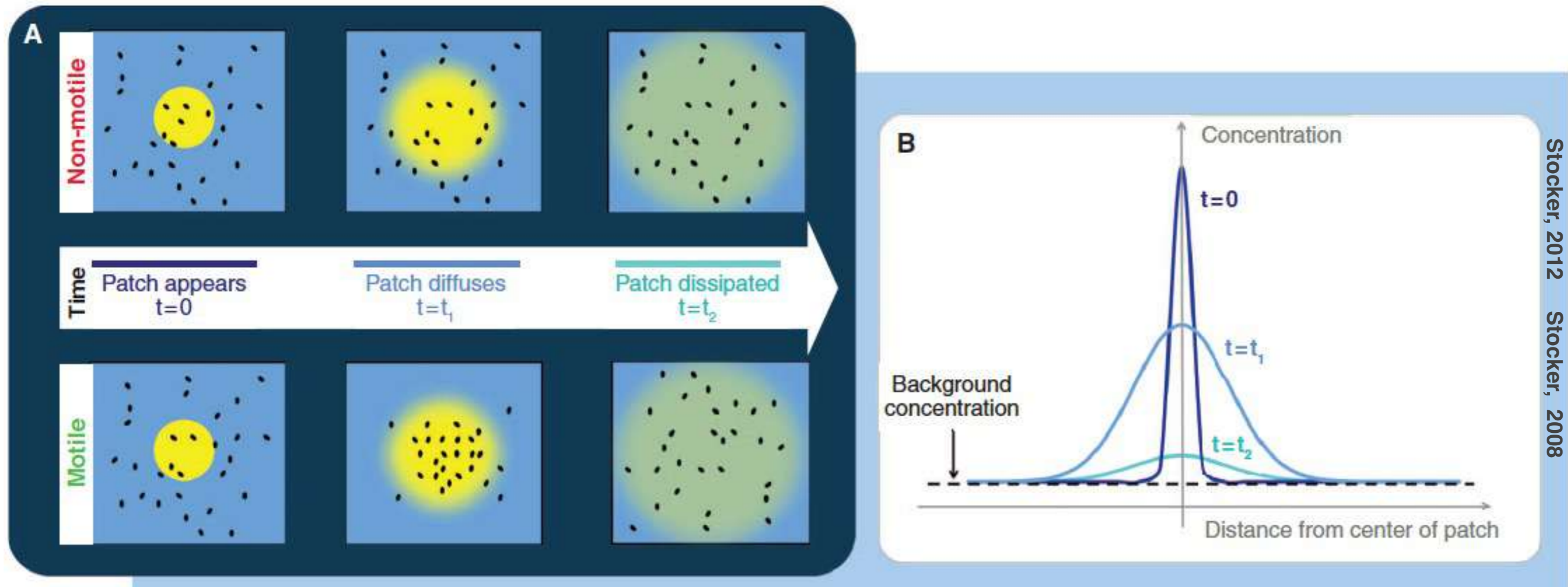
Malfatti unpub.

28



Seymour et al. 2017

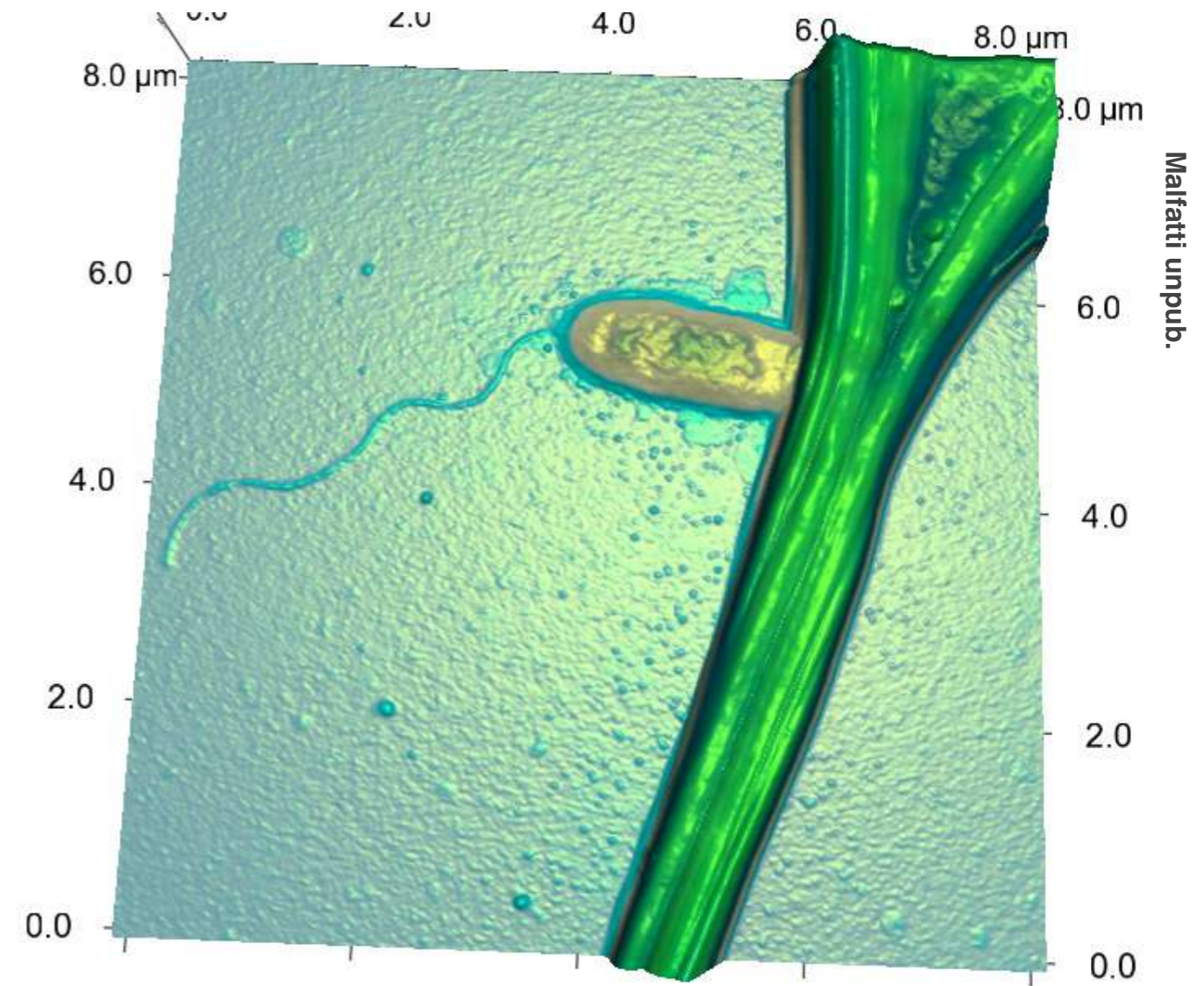
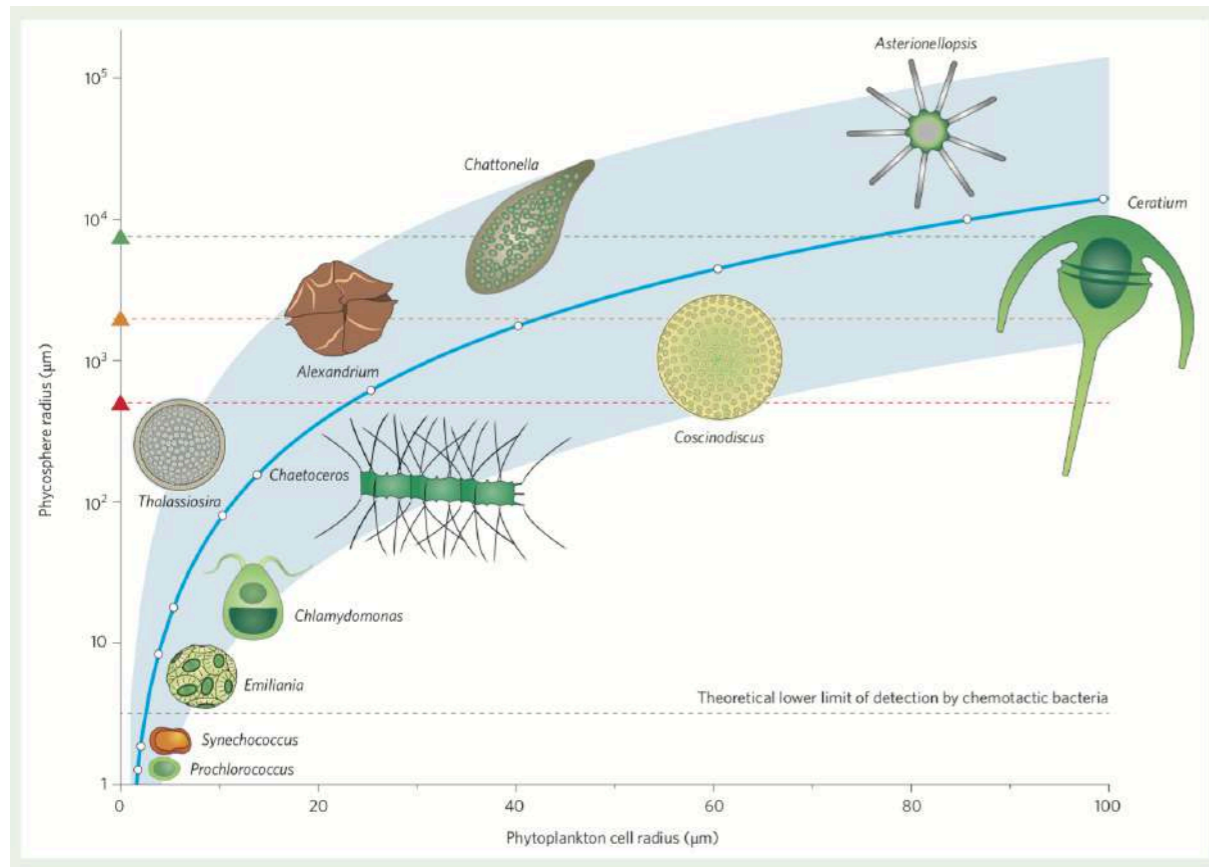
Marine bacteria can exploit ephemeral nutrient patches



Stocker, 2012 Stocker, 2008

- Motility as an adaptive strategy to respond to nutrient patches
- Exploration: non motile 0.45 pL (Brownian motion) vs motile 0.5 μ L every (motility 50 μ m/s) 10 min
- Daily exploration: non motile \sim 430 μ m cube vs motile \sim 1 cm cube chemotactic
- Copiotrophic populations outcompete nonmotile, oligotrophic populations during diatom blooms and bloom collapse conditions (more motile cells)

Bacteria-phytoplankton interactions

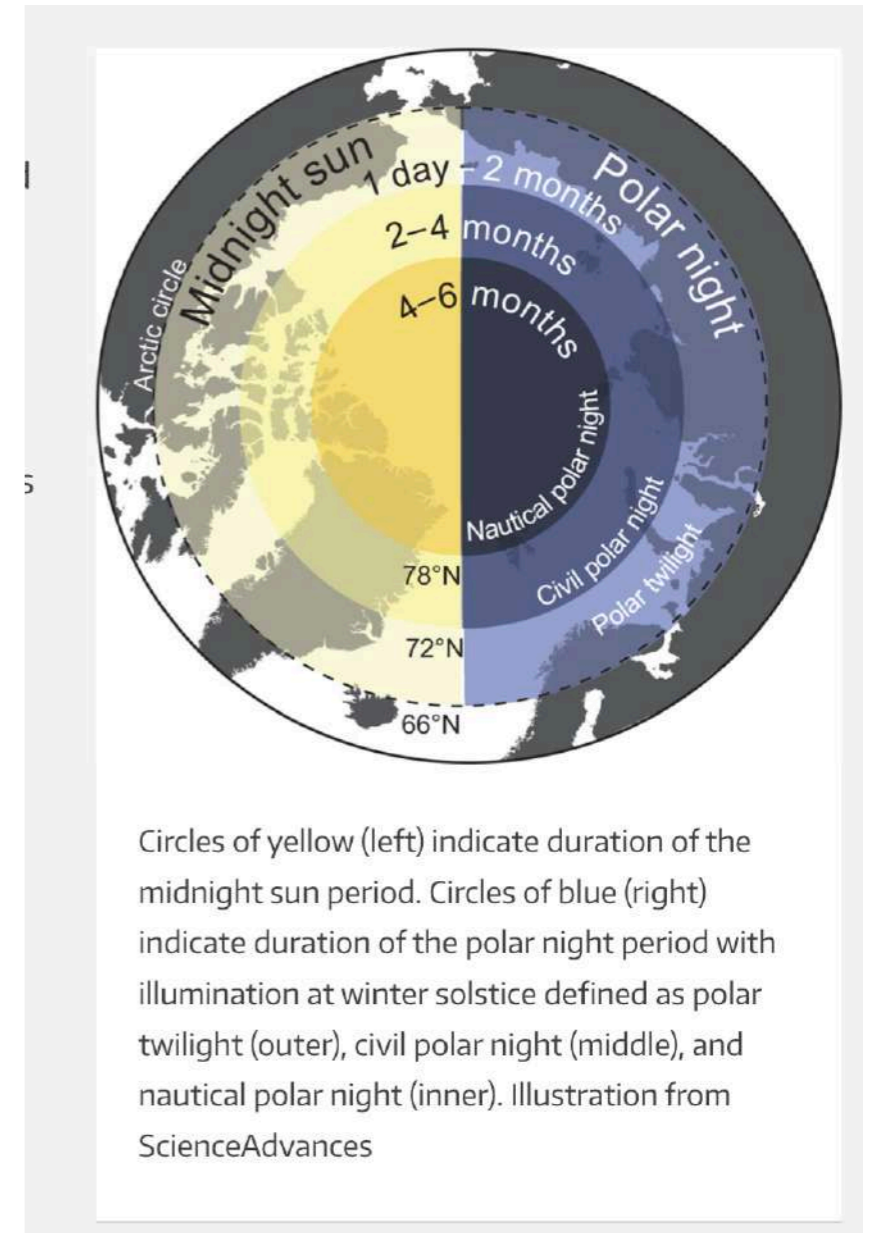


Malfatti unpub.

- The phycosphere, defined as the region surrounding a phytoplankton cell that is enriched in organic substrates exuded by the cell, is an important microenvironment for planktonic aquatic bacteria
- The phycosphere is defined as the region where the concentration is >50% above background

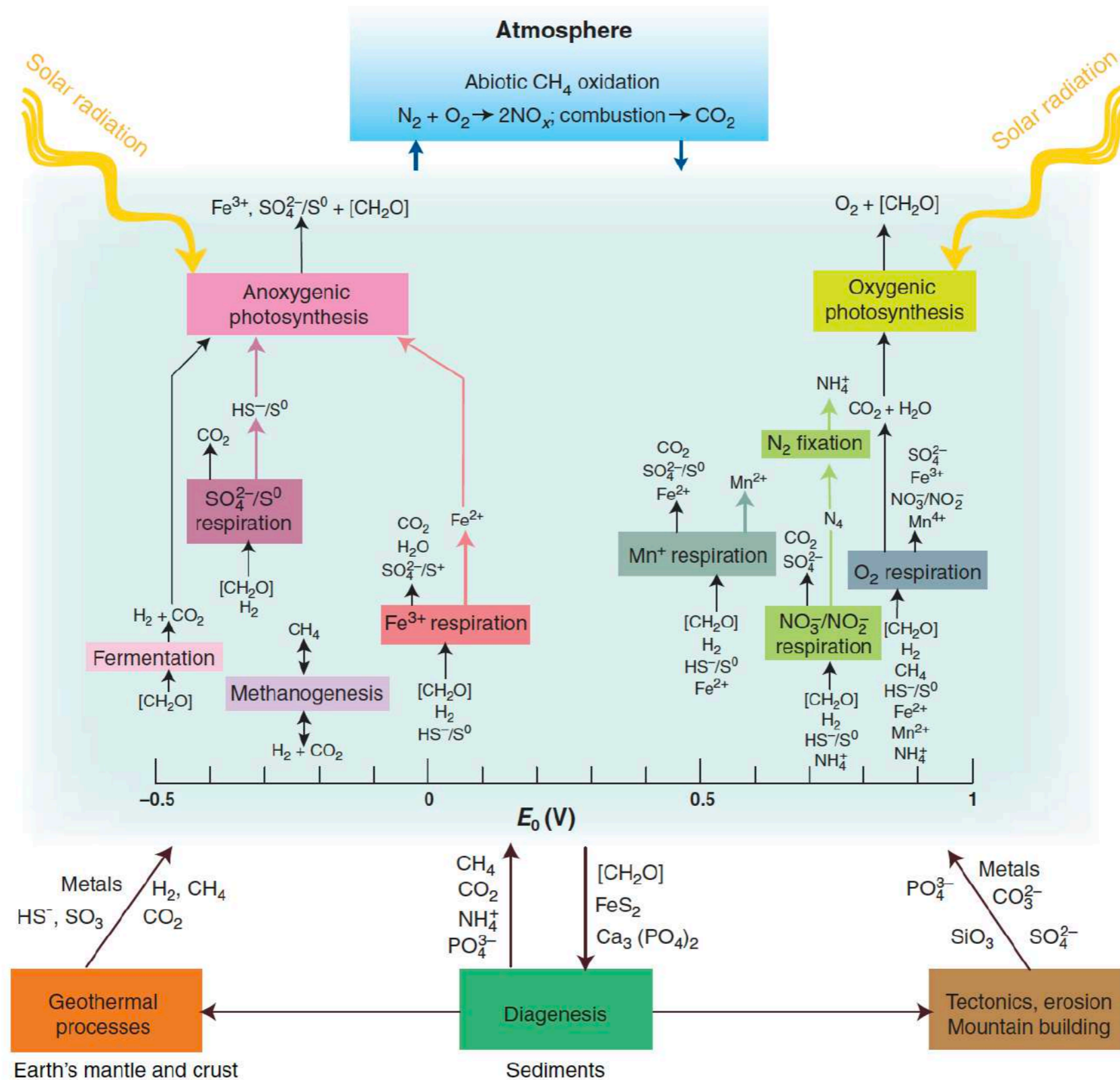
Polar winter

- Primary producers (microalgae, Euk) experience from lower or none solar irradiance
- Sea Ice formation
- Shift from autotrophy to mixotrophy
- Forming spores or resting structure
- Slowing down metabolisms and using storage reserve
- Producing cryoprotectant molecules to protect from freezing damage and desiccation
- Heterotrophic prokaryotes are active but slow



[https://
norwegianscitechnews.
com/2018/01/shedding-
light-zooplankton-dark/](https://norwegianscitechnews.com/2018/01/shedding-light-zooplankton-dark/)

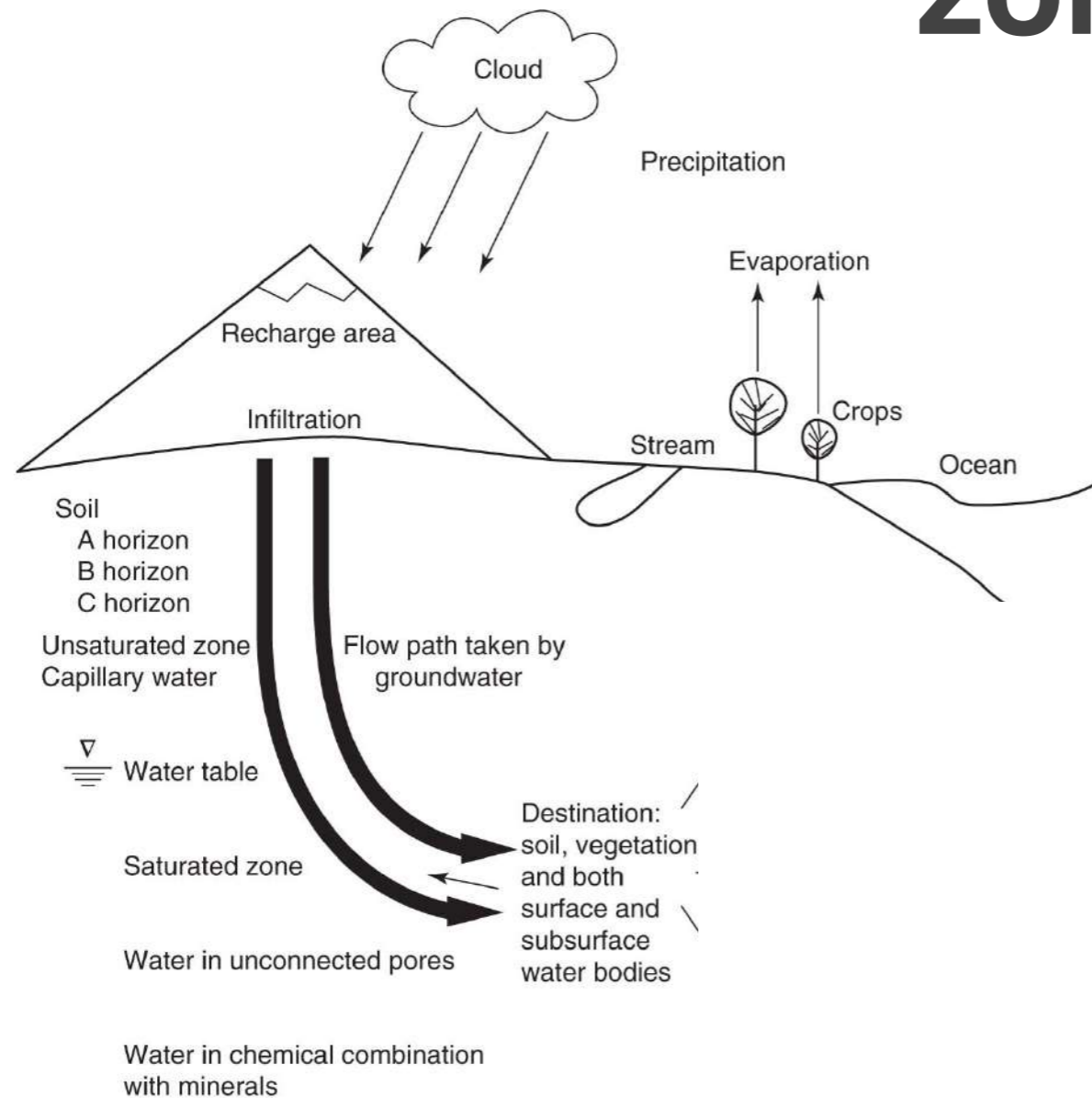
Biosphere model



Falkowski et al., 2008

FRESHWATER: LAKE RIVER

Hydrological cycle-soil-subsurface zone



- Water connects upper microbiomes (soil) with subsurface microbiome
- Lower microbiome at ~ 4 km at 125°C
- Underneath there are saturated and unsaturated subsurface zone
- Subsurface zone associated microbiome is important in dictating chemical composition of the water flowing in it (beside the influence of the soil and geological strata)

Figure 4.12 Conceptual flow system for understanding the role of soil and subsurface habitats in the hydrologic cycle. (Reprinted from Madsen, E.L. 1995. Impacts of agricultural practices on subsurface microbial ecology. *Adv. Agron.* **54**:1–67. Copyright 1995, with permission from Elsevier.)

About **30%** of all freshwater is **terrestrial ground water**, whereas the world's **lotic (streams and rivers)** and **lentic (lakes)** systems contribute only **0.3%**

Groundwater flow is governed largely by **recharge and discharge rates** and the **hydraulic properties of the saturated rocks**

Groundwater-flow paths are derived from hydraulic gradients between areas of recharge (e.g., precipitation in high topographic regions) and discharge (e.g., springs in low topographic regions)

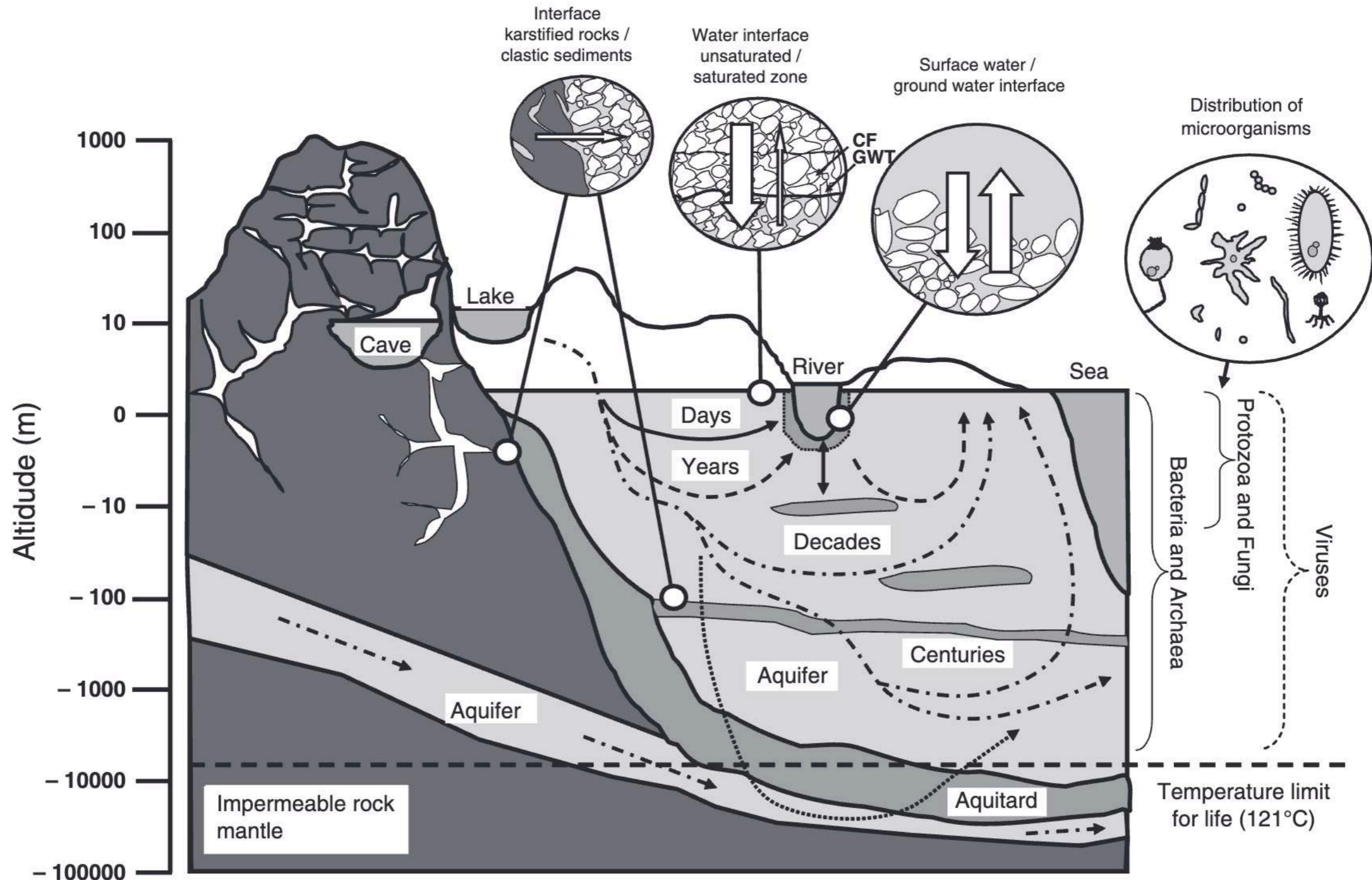
Groundwater environment are characterised by **continuous darkness and limited nutrients** —> **slow growing microbes**

Groundwater ecosystems deliver services that are of immense societal and economic value, such as:

- 1) purification of water and its storage in good quality for decades and centuries,
- 2) active biodegradation of anthropogenic contaminants and inactivation and elimination of pathogens,
- 3) nutrient recycling, and
- 4) mitigation of floods and droughts

Aquatic surface and subsurface microbial ecosystems

GRIEBLER AND LUEDERS, 2009



Arrows depict the flow of water carrying energy and matter through the subsurface, with boxes next to arrows indicating typical groundwater residence times.

Circles highlight transition zones between habitat types. Curly braces indicate the distribution of different microbial groups in the subsurface. CF, capillary fringe; GWT, groundwater table

Low microbial abundance in low C system

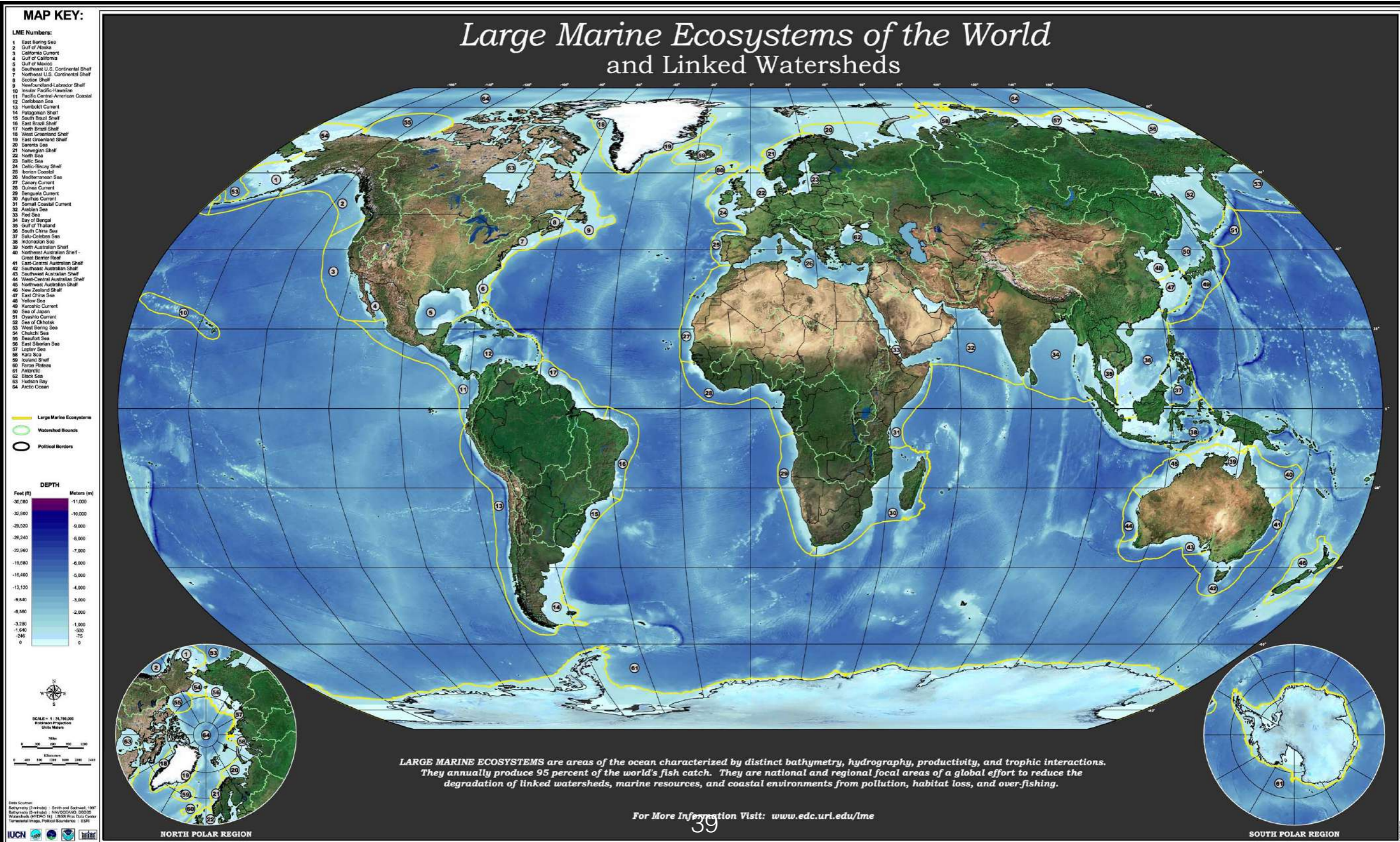
Table 2 Abundance of microbial groups in different subsurface habitats

Group	Habitat	Contamination	Abundance (cells cm ⁻³)	Reference
Prokaryota				
Bacteria	Water from karst and cave systems	No	10 ² –10 ⁵	Gounot (1994); Farnleitner <i>et al.</i> (2005)
	Sediment from cave waters	No	10 ⁴ –10 ⁸	Gounot (1994); Rusterholtz & Mallory (1994)
	Water from granite and basalt systems	No	10 ² –10 ⁵	Stevens & McKinley (1995); Pedersen (1997)
	Ground water	No	10 ³ –10 ⁶	Ghiorse & Wilson (1988); Madsen & Ghiorse (1993); Pedersen (2000); Griebler (2001)
		Yes	10 ³ –10 ⁷	
	Groundwater-saturated porous sediment	No	10 ⁵ –10 ⁸	up to 10 ¹⁰
		Yes	up to 10 ¹⁰	
Archaea	Vadose zone sediment	No	10 ⁴ –10 ⁸	Brockman <i>et al.</i> (1992); Kieft <i>et al.</i> (1993) Detmers <i>et al.</i> (2004)
		Yes or no	up to 20% of total cell counts	
Protozoa				
Heterotrophic Flagellata	Ground water	No	10 ⁰ –10 ²	Hirsch <i>et al.</i> (1992); Madsen & Ghiorse (1993); Novarino <i>et al.</i> (1997)
		Yes	up to 10 ⁵	
	Groundwater-saturated porous sediment	No	10 ³ –10 ⁵	Novarino <i>et al.</i> (1997) Novarino <i>et al.</i> (1997)
		Yes	up to 10 ⁸	
Amoebae	Ground water	Yes or no	10 ⁻¹ –10 ¹	Hirsch <i>et al.</i> (1992); Madsen & Ghiorse (1993); Novarino <i>et al.</i> (1997)
Ciliata	Ground water (near surface)	Yes or no	10 ⁻¹ –10 ⁰	
Heliozoa	Ground water (near surface)	No	10 ⁻¹ –10 ⁰	

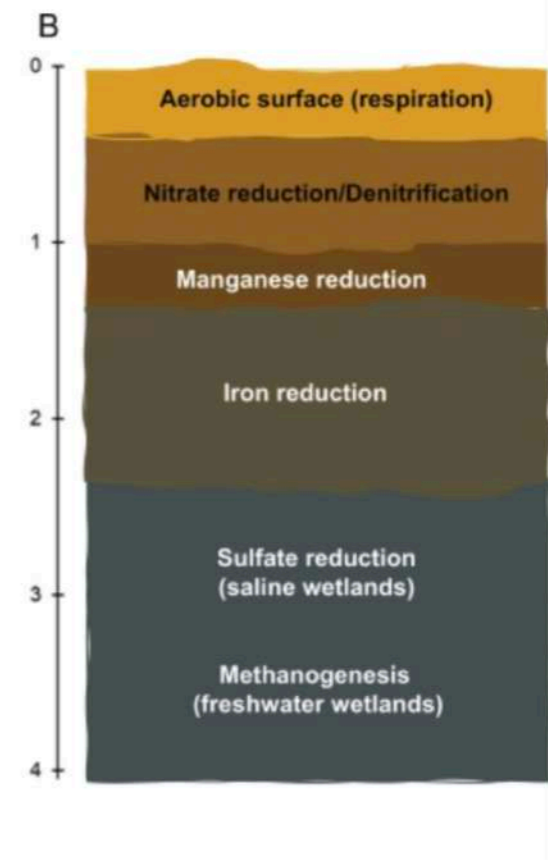
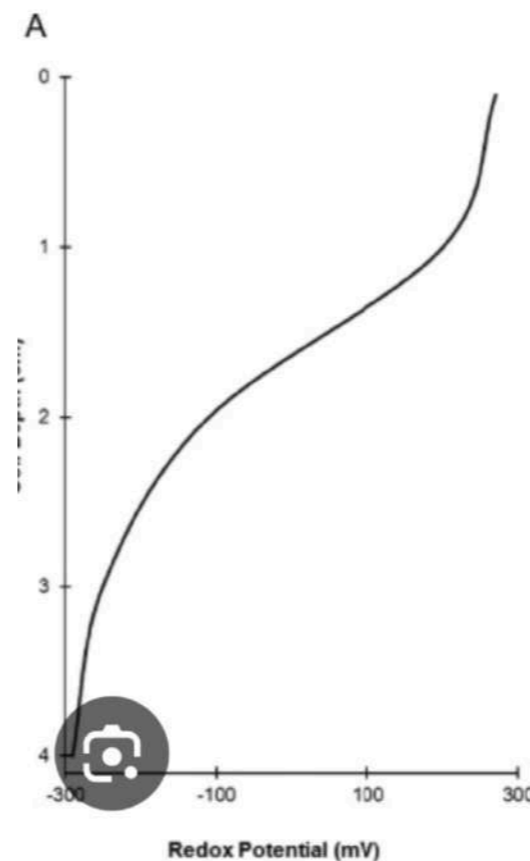
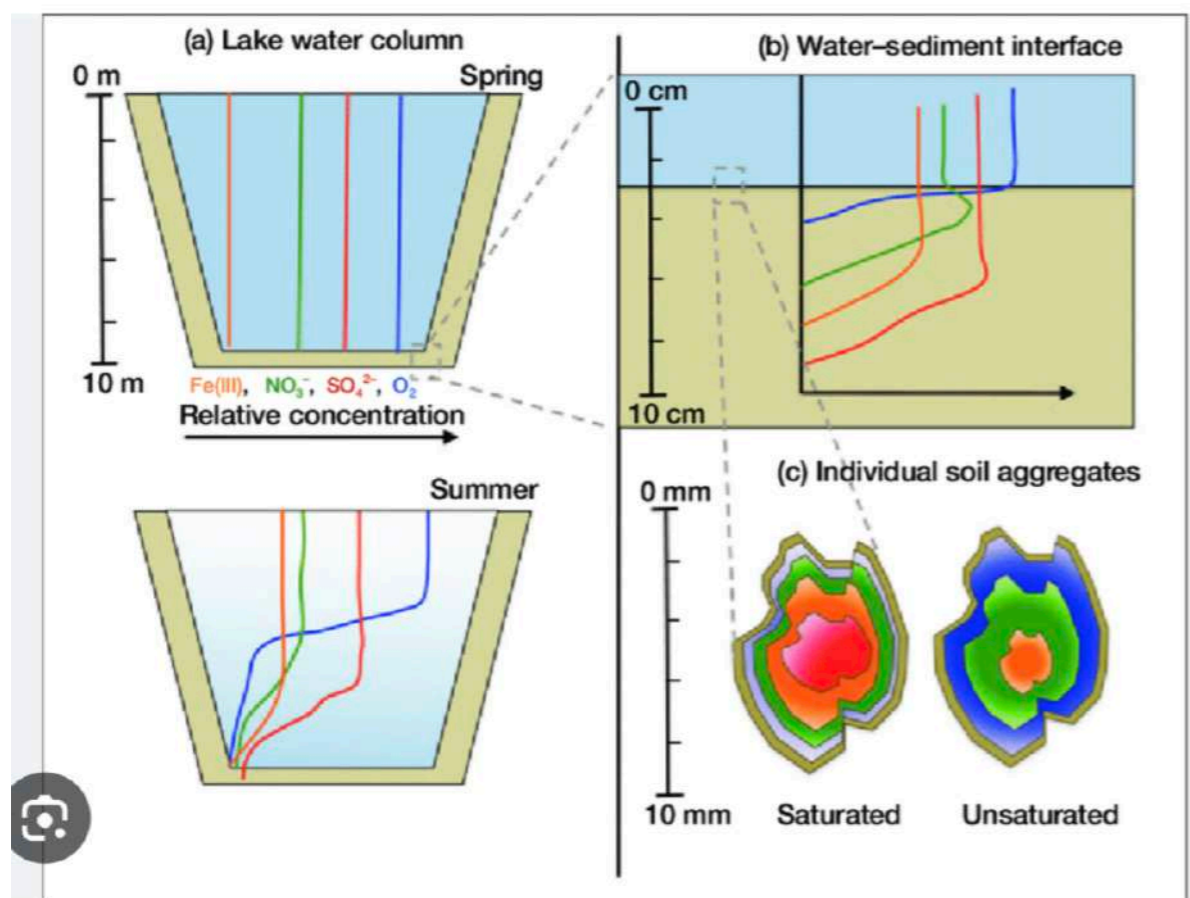
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Large marine ecosystems and watersheds

- On land watersheds don't communicate with other watersheds
- Characteristic chemical and biological fingerprint of each watershed



Steep microbial gradients in lake and sediment shape microbial community and metabolism



Mobilian and Craft, 2022

The **redox potential** is used to describe a system's overall reducing or oxidizing capacity

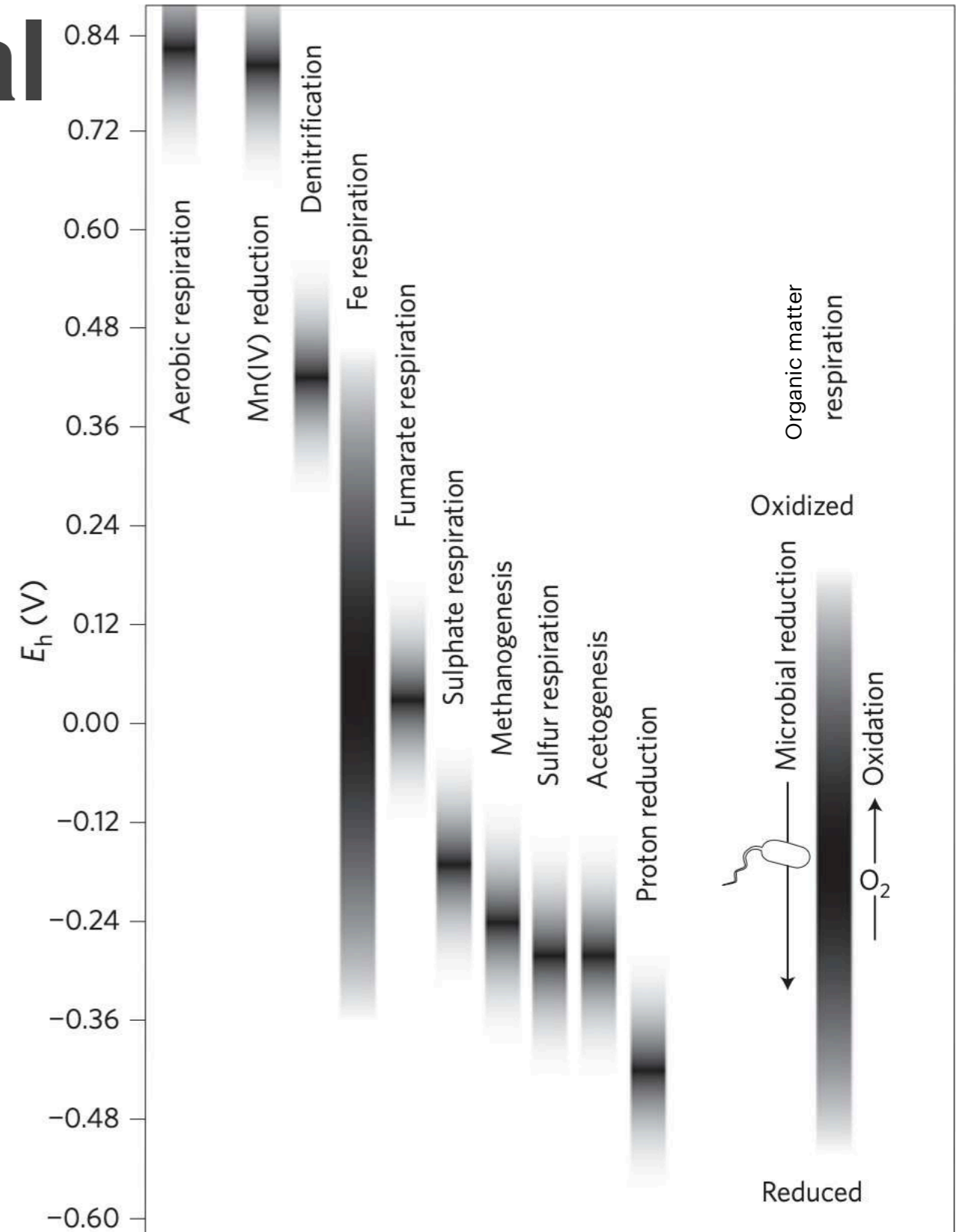
The redox potential is measured in millivolts (mV) relative to a standard hydrogen electrode and is commonly measured using a platinum electrode with a saturated calomel electrode as reference

In well-oxidized water, with oxygen concentrations $> \sim 1 \text{ mg O}_2 \text{ L}^{-1}$, the redox potential will be $> 300\text{--}500 \text{ mV}$

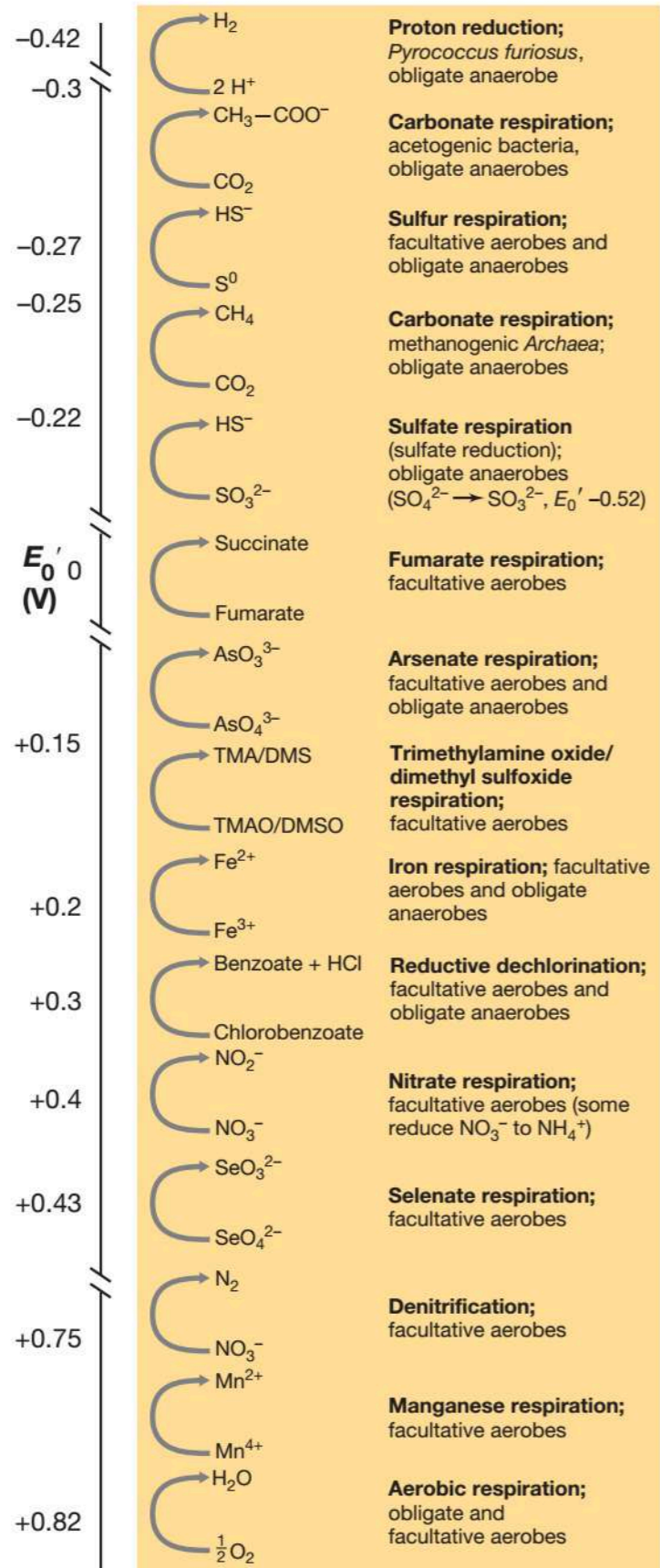
In reduced environments, such as in the deep water of stratified lakes or the sediment of eutrophic lakes, the redox potential will be $< 100 \text{ mV}$ or even negative

Reduction potential ranges of microbial respiration

- The achievable energy yield of ETC depends on the difference in electrical potential between electron donor and acceptor
- Microbes able to respire in multiple ways will always choose available acceptors with the **biggest potential difference** to the donor (e.g., *E. coli* $O_2 > NO_3^- > \text{fumarate}$)



Anaerobic respiration



Microbially mediated reactions

Microaerophiles

$4\text{Fe}^{2+} + 10\text{H}_2\text{O} + \text{O}_2 \rightarrow 4\text{Fe}(\text{OH})_3 + 8\text{H}^+$
Gallionella spp., *Leptothrix* spp.,
Mariprofundus spp., *Sideroxydans* spp.

Photoferrotrophs

$\text{HCO}_3^- + \text{Fe}^{2+} + 10\text{H}_2\text{O} \xrightarrow{h\nu} (\text{CH}_2\text{O}) + 4\text{Fe}(\text{OH})_3 + 7\text{H}^+$
Rhodospseudomonas palustris TIE-1
Rhodobacter sp. SW2
Chlorobium ferrooxidans (KoFox)
Thiodictyon sp. F4

NO₃⁻-reducing Fe(II)-oxidizers

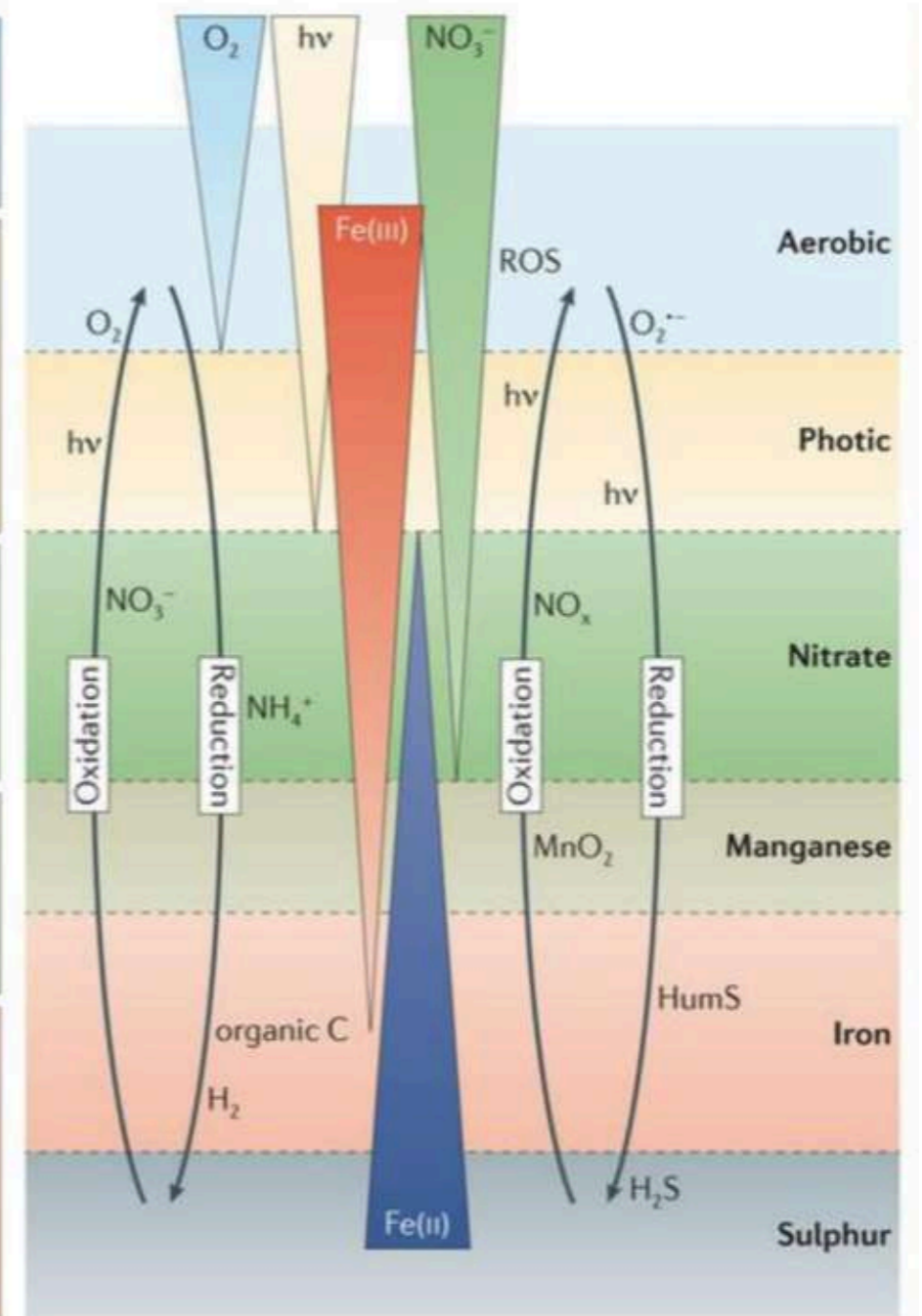
$10\text{Fe}^{2+} + 2\text{NO}_3^- + 24\text{H}_2\text{O} \rightarrow 10\text{Fe}(\text{OH})_3 + \text{N}_2 + 18\text{H}^+$
Acidovorax spp., KS, 2002
Thiobacillus denitrificans

Fe-ammoX

$\text{NH}_4^+ + 6\text{FeOOH} + 10\text{H}^+ \rightarrow \text{NO}_2^- + 6\text{Fe}^{2+} + 10\text{H}_2\text{O}$
 Unknown

Fe(III)-reducing organic C and/or H₂-oxidizers

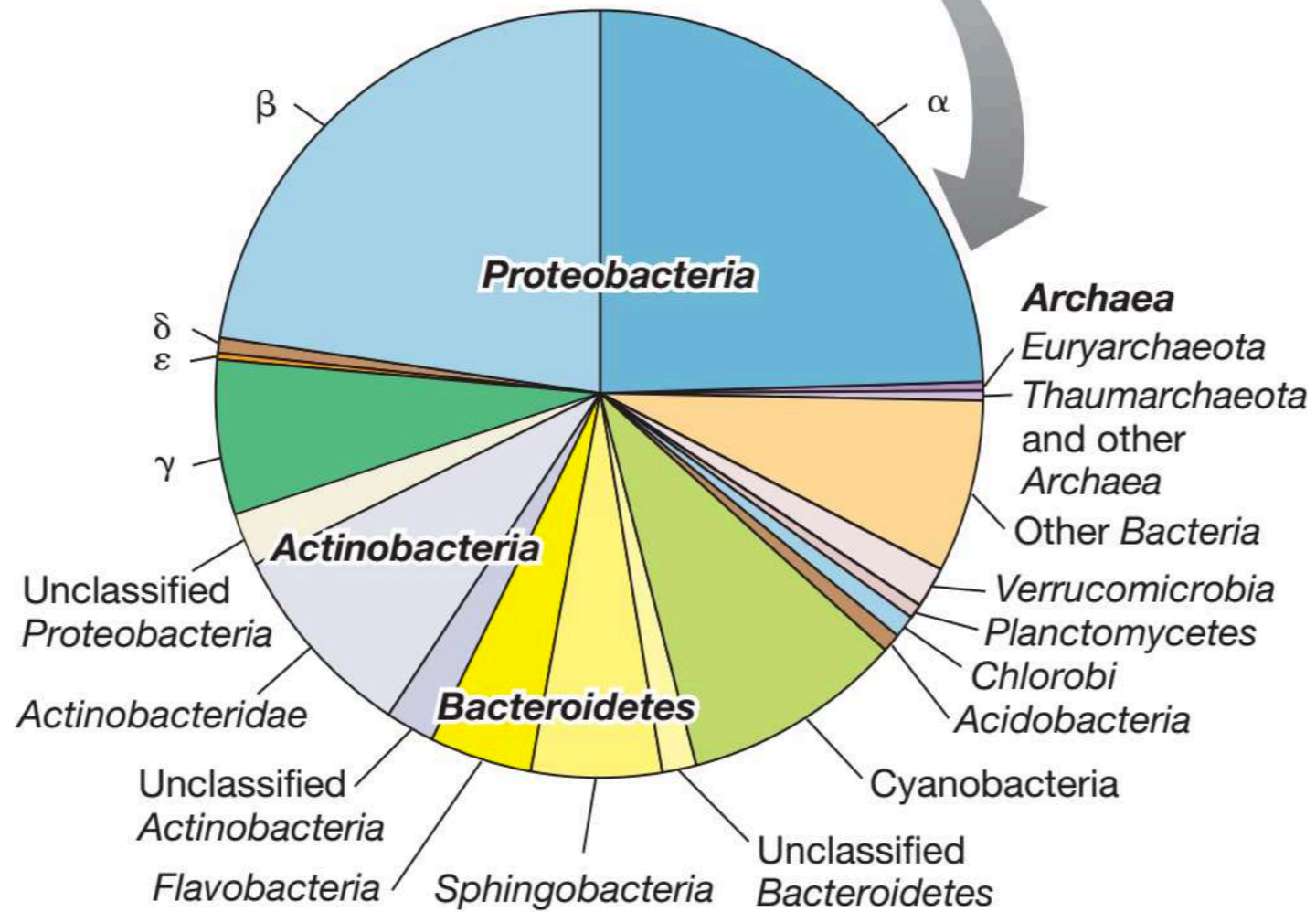
$4\text{FeOOH} + \text{CH}_3\text{CHOHCOO}^- + 7\text{H}^+ \rightarrow 4\text{Fe}^{2+} + \text{CH}_3\text{COO}^- + \text{HCO}_3^- + 6\text{H}_2\text{O}$
 $2\text{Fe}(\text{OH}) + \text{H}_2 \rightarrow 2\text{Fe}^{2+} + 2\text{H}_2\text{O}$
Geobacter spp., *Shewanella* spp.,
Albidoferax ferrireducens, *Geothrix* spp.



Freshwater microbial diversity



1. Extract lake microbial community DNA
2. Isolate, sequence, and analyze 16S rRNA genes



Madigan et al. 2020

- 16S rRNA gene based diversity

Biogeochemical processes

Table 1 | **Examples of physiological processes catalysed by microorganisms in biosphere habitats**

Process	Nature of process	Typical habitat
Carbon cycle		
Photosynthesis	Light-driven CO ₂ fixation into biomass	Ow, Fw, FwS, Os
C respiration	Oxidation of organic C to CO ₂	All
Cellulose decomposition	Depolymerization, respiration	Sl
Methanogenesis	CH ₄ production	Sw, FwS, Os
Aerobic CH ₄ oxidation	CH ₄ becomes CO ₂	All
Anaerobic CH ₄ oxidation	CH ₄ becomes CO ₂	Os
Biodegradation		
Synthetic organic compounds	Decomposition, CO ₂ formation	All
Petroleum hydrocarbons	Decomposition, CO ₂ formation	All
Fuel additives (MTBE)	Decomposition, CO ₂ formation	Sl, Sw, Gw
Nitroaromatics	Decomposition, CO ₂ formation	Sl, Sw, Gw
Pharmaceuticals, personal care products	Decomposition	Sl, Sw, Gw
Chlorinated solvents	Compounds are dechlorinated through respiration in anaerobic habitats	Sl, Sw, Gw

Madsen, 2005

As, arsenic; C, carbon; CH₄, methane; CO₂, carbon dioxide; Fe, iron; FeS, iron sulphide; Fw, freshwater; FwS, freshwater sediment; Gw, groundwater; H₂, hydrogen; Hg, mercury; Hg²⁺, mercuric ion; MTBE, methyl tertiary butyl ether; N₂, nitrogen; NH₃, ammonia; NH₄⁺, ammonium; NO₂⁻, nitrite; NO₃⁻, nitrate; Os, ocean sediments; Ow, ocean waters; S⁰, elemental sulphur; S²⁻, sulphide; Sl, soil; SO₄²⁻, sulphate; Sw, sewage; U, uranium.

Nitrogen cycle

N ₂ fixation	N ₂ gas becomes NH ₃	Sl, Ow
NH ₄ ⁺ oxidation	NH ₃ becomes NO ₂ ⁻ , NO ₃ ⁻	Sl, Sw
Anaerobic NH ₄ ⁺ oxidation	NO ₂ ⁻ and NH ₃ become N ₂ gas	Sw, Os
Denitrification	NO ₃ ⁻ is used as an electron acceptor and converted to N ₂ gas	Sl, Sw

Sulphur cycle

S ₂ oxidation	S ²⁻ and S ⁰ become SO ₄ ²⁻	Os, FwS
SO ₄ ²⁻ reduction	SO ₄ ²⁻ is used as an electron acceptor and converted to S ⁰ and S ²⁻	Os, Sw, Gw

Other elements

H ₂ oxidation	H ₂ is oxidized to H ⁺ , electrons reduce other substances	Sw, Sl, Os, FwS
Hg methylation and reduction	Organic Hg is formed and Hg ²⁺ is converted to Hg	FwS, Os
(per)chlorate reduction	Oxidants in rocket fuel and other sources are converted to chloride	Gw
U reduction	U oxyanion is used as an electron acceptor; therefore immobilized	Gw
As reduction	As oxyanion is used as an electron acceptor; therefore toxicity is diminished	FwS, Gw
Fe oxidation, acid mine drainage	FeS ores are oxidized, strong acidity is generated	FwS, Gw

Madsen, 2005

As, arsenic; C, carbon; CH₄, methane; CO₂, carbon dioxide; Fe, iron; FeS, iron sulphide; Fw, freshwater; FwS, freshwater sediment; Gw, groundwater; H₂, hydrogen; Hg, mercury; Hg²⁺, mercuric ion; MTBE, methyl tertiary butyl ether; N₂, nitrogen; NH₃, ammonia; NH₄⁺, ammonium; NO₂⁻, nitrite; NO₃⁻, nitrate; Os, ocean sediments; Ow, ocean waters; S⁰, elemental sulphur; S²⁻, sulphide; Sl, soil; SO₄²⁻, sulphate; Sw, sewage; U, uranium.